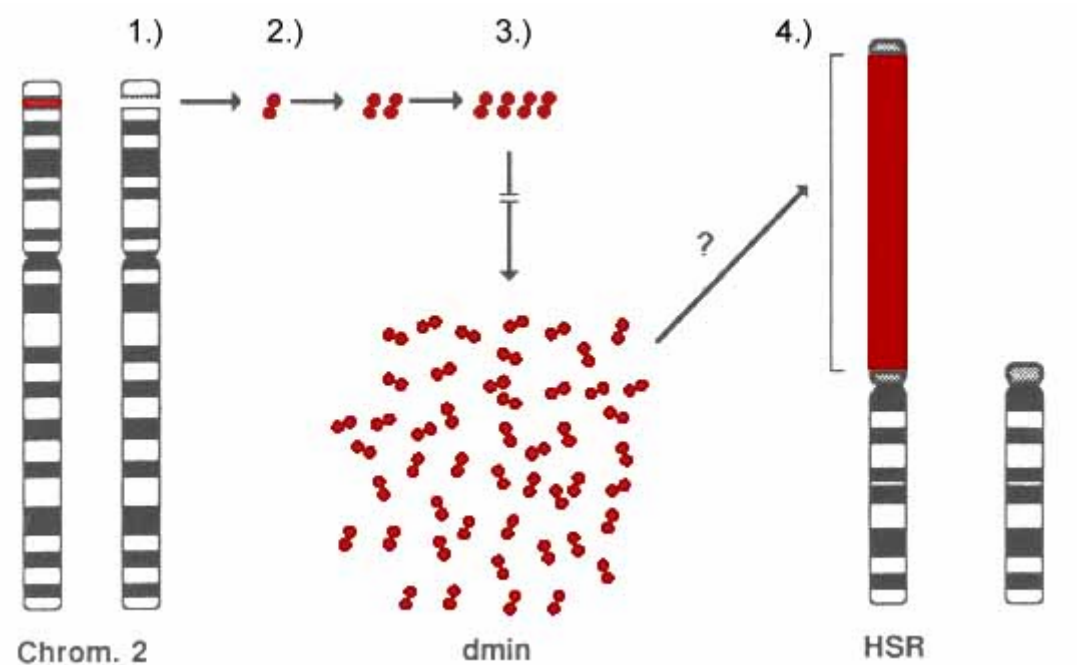
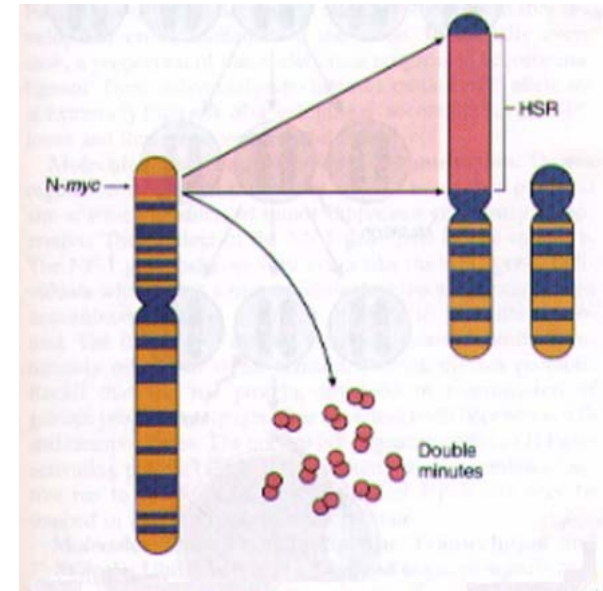
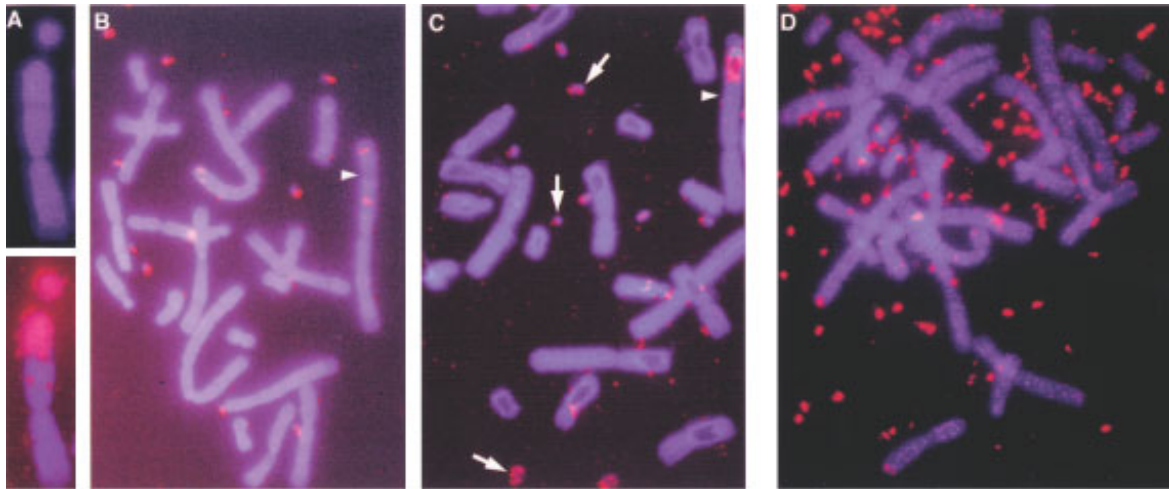


„Double minutes“ und HSR (homogeneously staining region)





HSR → dmin upon DS break induced by a homing endonuclease (I-SceI).

Arnaud Coquelle, Lorène Rozier, Bernard Dutrillaux and Michelle Debatisse
 ONCOGENE, 31 October 2002, Volume 21, Number 50, Pages 7671-7679
 Induction of multiple double-strand breaks within an hsr by meganuclease I-SceI expression
 or fragile site activation leads to formation of double minutes and other chromosomal rearrangements

DNA-Amplifikation in Tumorzellen

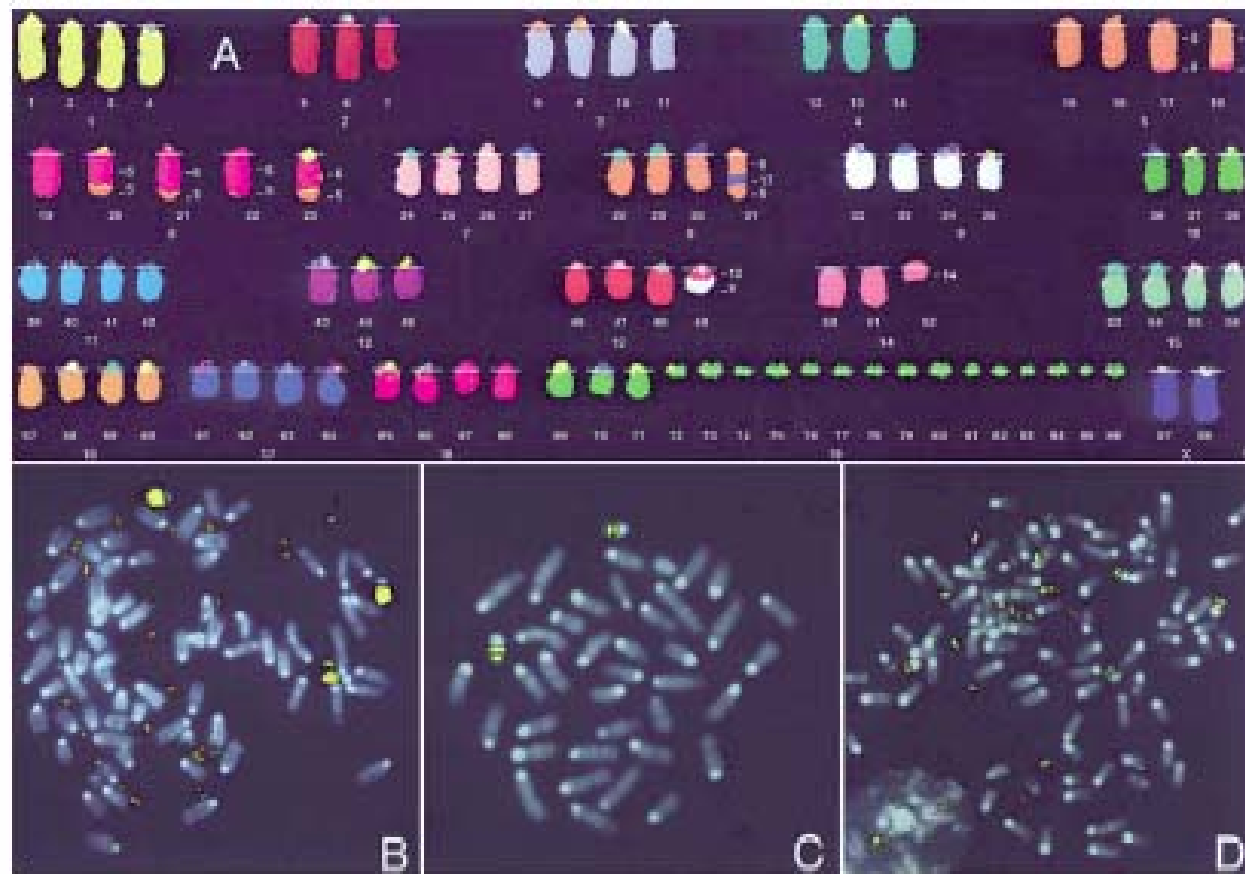
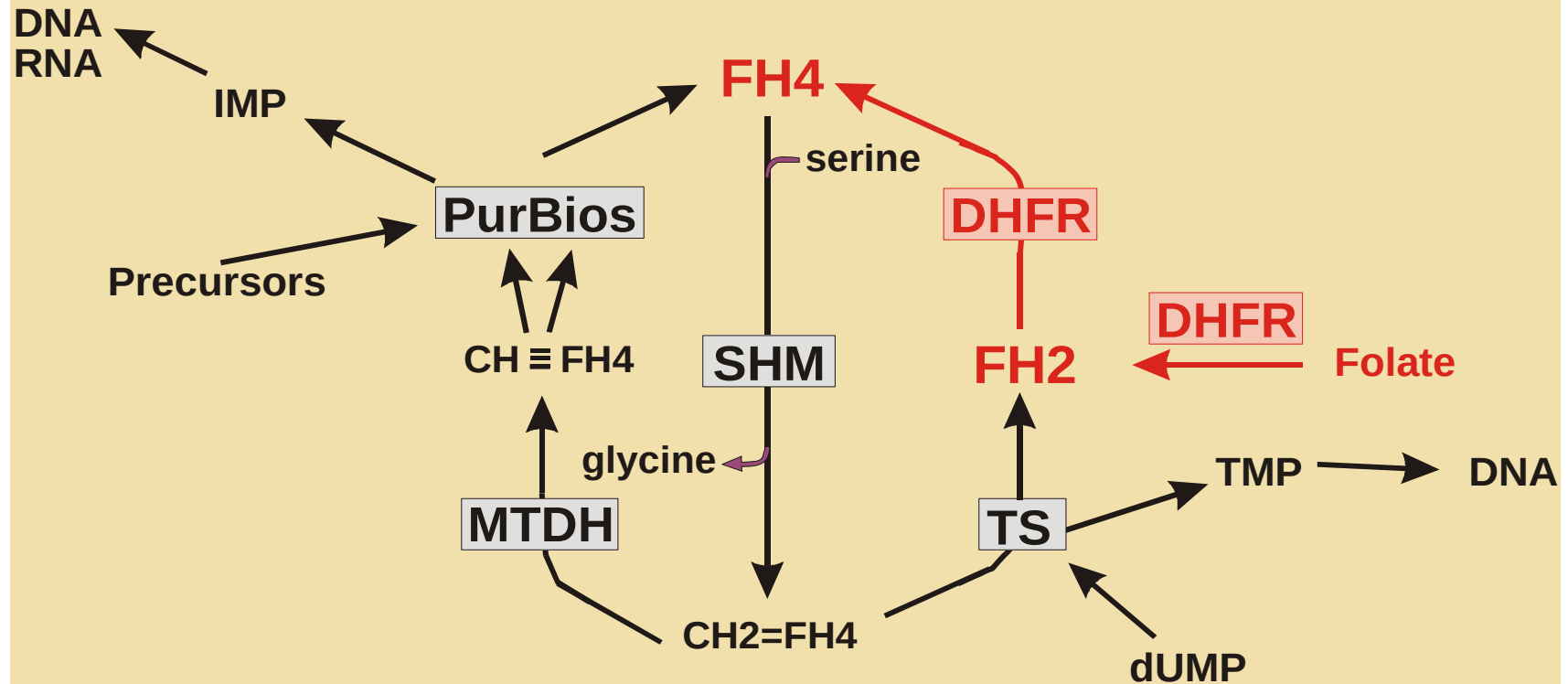


Fig. 1. SKY and FISH analysis of the HCC-myc 3 cell line. (A) Spectral classified karyotype of a hypodiploid cell showing numerical and structural abnormalities among which are the presence of 15 pairs of double minutes derived from chromosome 19. (B) Metaphase after hybridization with chromosome 19 painting probe confirming the origin of DM. Four copies of chromosome 19 and DM are labeled with the painting probe. (C) Normal metaphase from stimulated mouse spleen cell culture hybridized with a biotinylated PCR amplified DNA probe of microdissected DM showing fluorescent signal at regions B1.1-B1.3 and D1-D2.1 of both chromosomes 19. (D) FISH with microdissected DM probe of a metaphase from HCC-myc-3 cell line showing labeling at two sites of chromosomes 19 and DM.

Reduction of folate to tetrahydrofolate



DHFR:

Dihydrofolate reductase

SHM:

Serine hydroxymethyltransferase

TS:

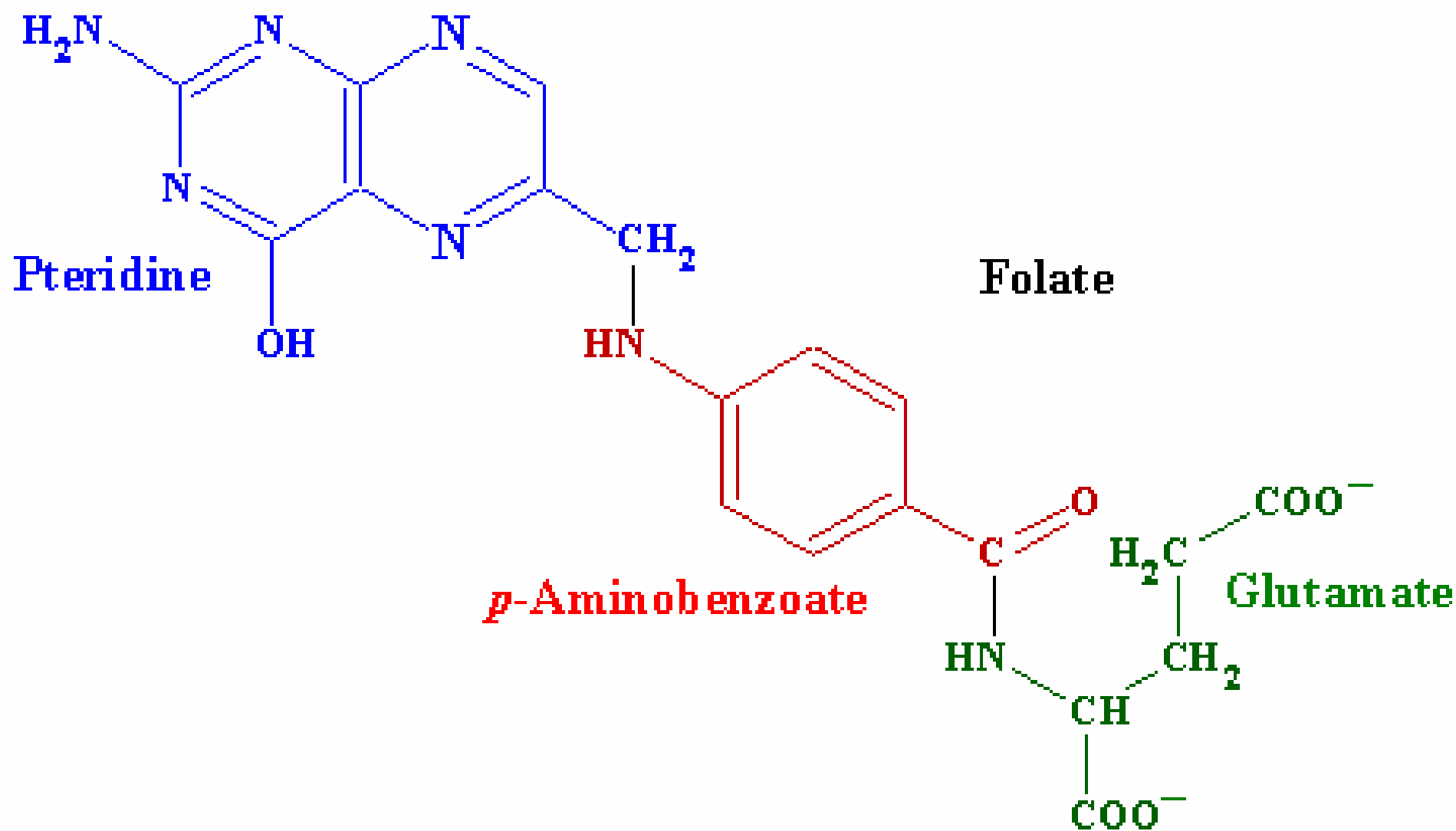
Thymidylate synthetase

PurBios:

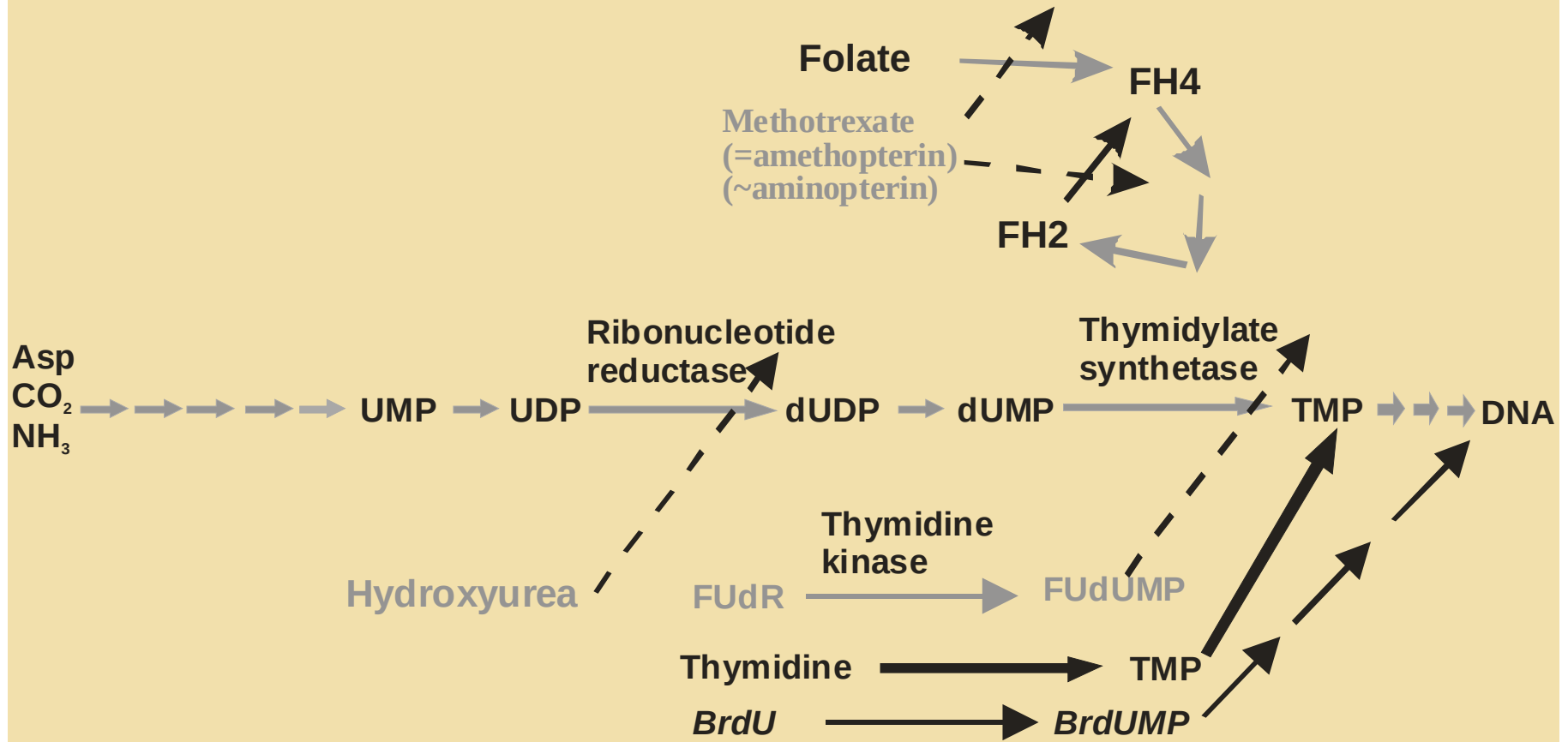
Purine nucleotide biosynthetic pathway

MTDH:

Methylenetetrahydrofolate dehydrogenase



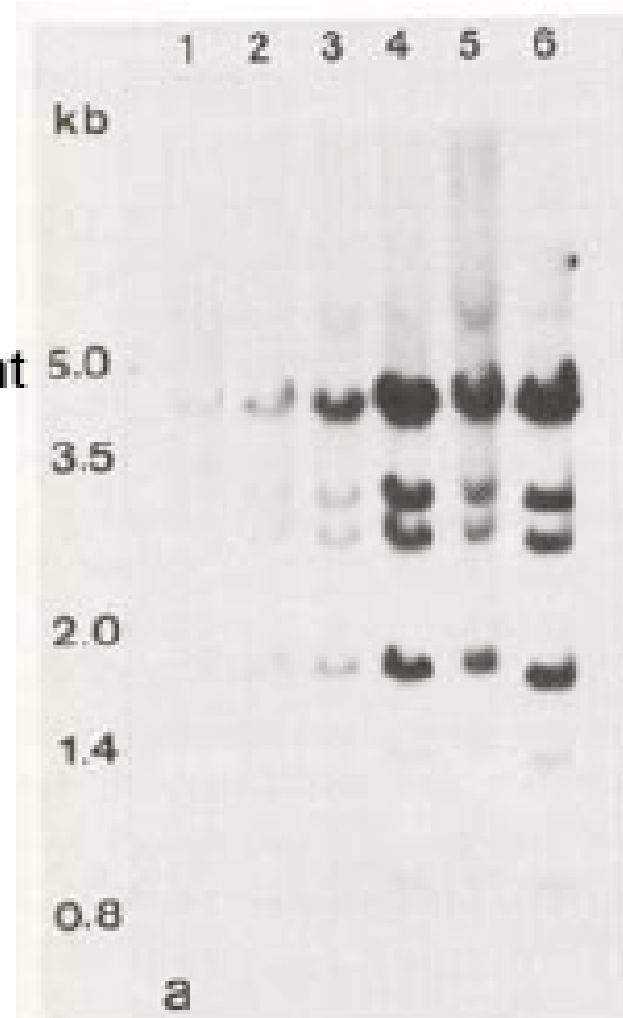
Thymidylate biosynthesis, salvage pathways, and inhibitors



Methotrexate Resistance by DHFR Amplification

CHO cells
↓
mtx
mtx
mtx
↓
Colonies of mtx resistant
CHO cells
↓
Prepare Genomic DNA
↓
*Eco*RI digest
↓
Southern Blot using
DHFR cDNA probe

Lane 1 Parental CHO cells
Lanes 2-6 mtx resistant cells



Copy Number
Per **haploid**
Genome

Lane 1	1.0
Lane 2	3.0
Lane 3	6.7
Lane 4	39
Lane 5	17
Lane 6	53

Adapted from:
Johnston et al
PNAS 80, 3711

Adapted from:
Lehninger
"Principles of
Biochemistry"

Causes of Cancer: Replication Errors

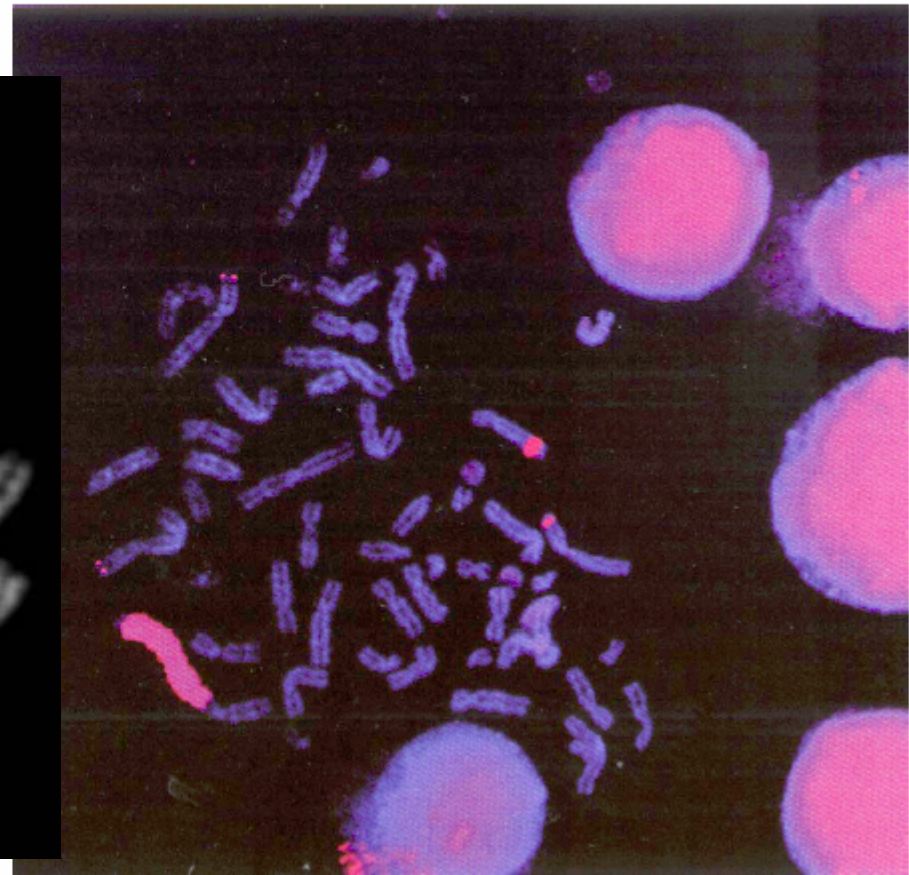
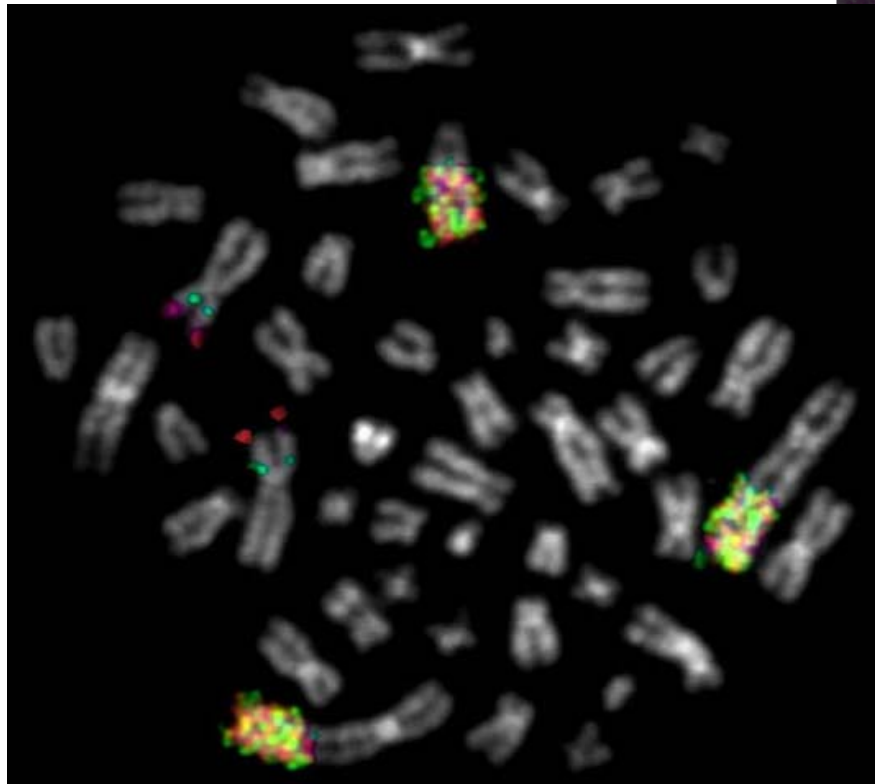
- Gene amplification
 - Over expression of oncogenes

Oncogene	Tumour	Amplification
MYC	Promyelocytic leukemia	20x
MYC	Small-cell lung carcinoma	5→30x
MYCN	Neuroblastoma	>100x
MYB	Acute myeloid leukemia	10x
MYB	Colon carcinoma	10x
EGFR	Epidermal carcinoma	30x
KRAS	Lung cancer	10x

- Leads to...
 - Homogenously Stained Regions (HSR)
 - Double minutes (DM)

Causes of Cancer

- Homogenously Stained Regions (HSR)
 - NMYC amplification



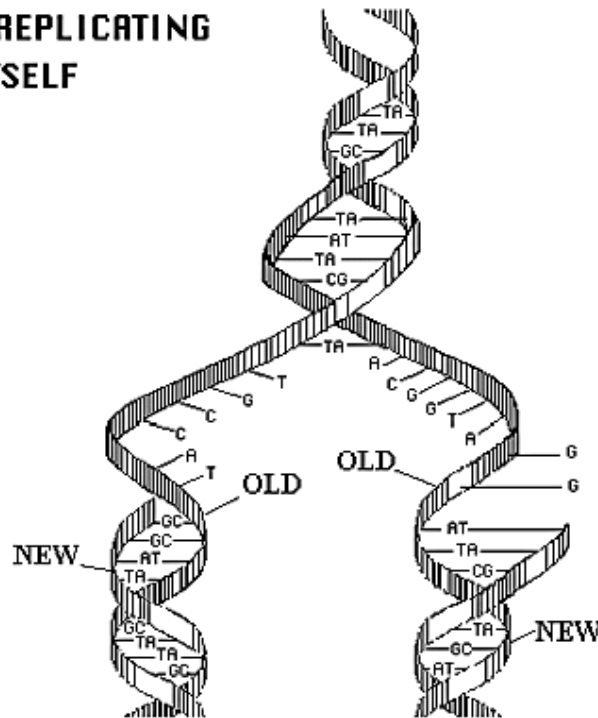
'Molecular and Genetic Analysis of Human Traits' Maroni (2001)

Andere amplifizierte Gene

Adenosine deaminase	ADA	deoxycoformycin	MTX + adenosine
Ornithine decarboxylase	ODC	difluoromethyl-ornithine	Polyamine-free
Asparagine synthetase	AS		Asparagine-free
Ribonucleoside reductase	RR	hydroxyurea	Deoxynucleoside-free
Tri-functional pyrimidine synthetic enzyme	CAD	PALA N-(phosphonacetyl)-L-aspartate	Pyrimidine-free
Thymidylate synthetase	TS	FUdr	Thymidine-free
Dihydrofolate reductase	DHFR	MTX	Gly-, TdR-, purine-free
Glutamine synthetase	GS	Methionine sulfoxime	Gln-free

DNA-Replikation bei Pro-und Eukaryoten

DNA REPLICATING
ITSELF



Die DNA-Replikation findet in der S-Phase des Zellzyklus statt

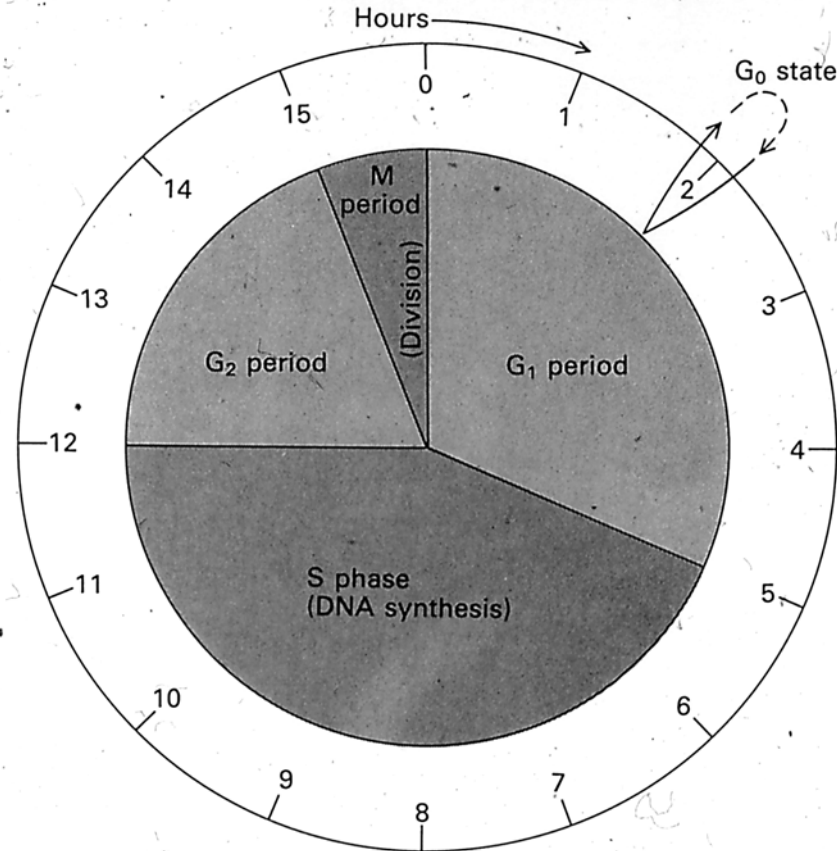


Figure 5-13 The cell cycle in a mammalian cell having a generation time of 16 h. The three periods spanning the first 15 h or so—the G₁ (first gap) period, the S (synthetic) phase, and the G₂ (second gap) period—make up the interphase, during which DNA and other cellular macromolecules are synthesized. The remaining hour is the M (mitosis) period; it is during this period that the cell actually divides.

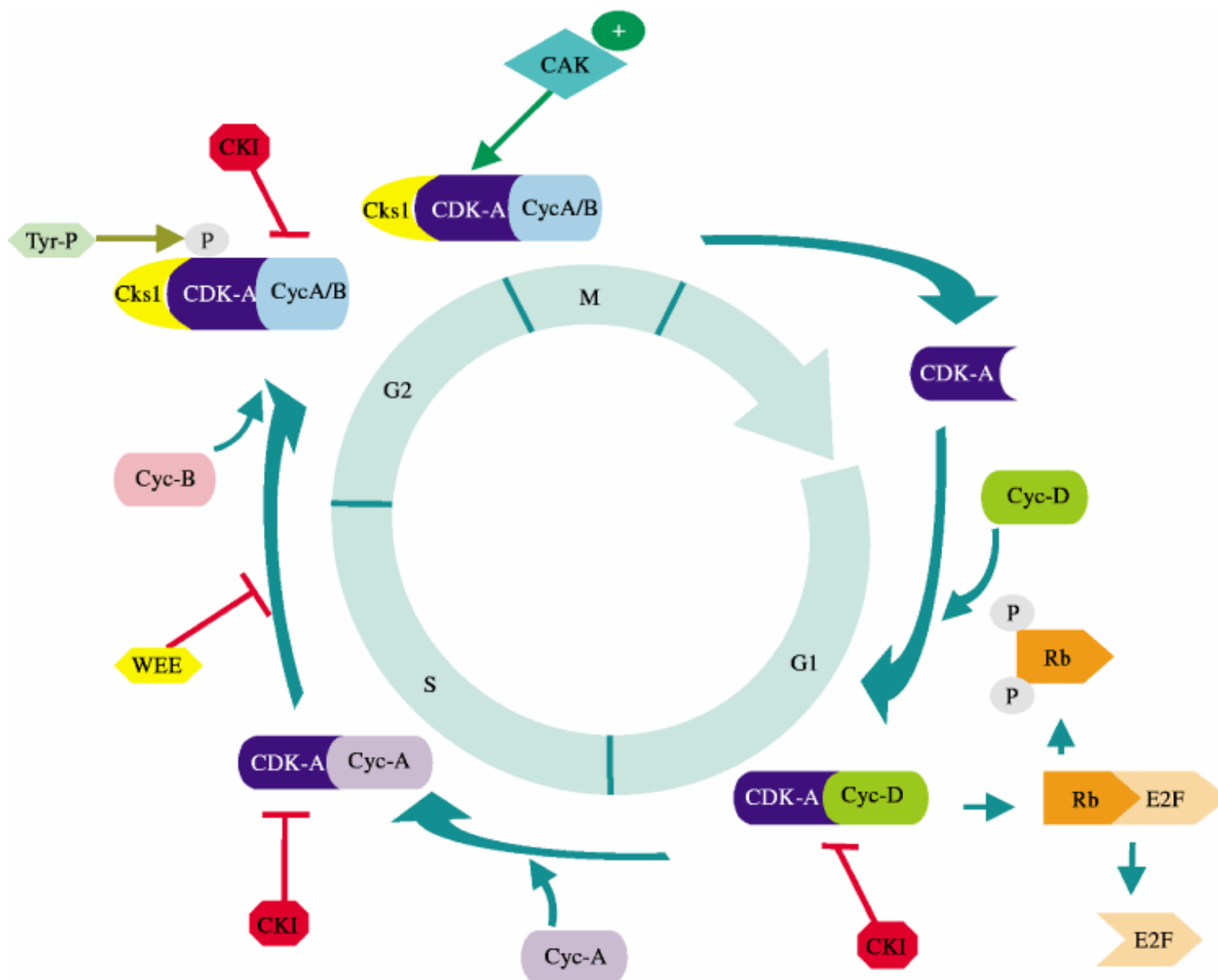
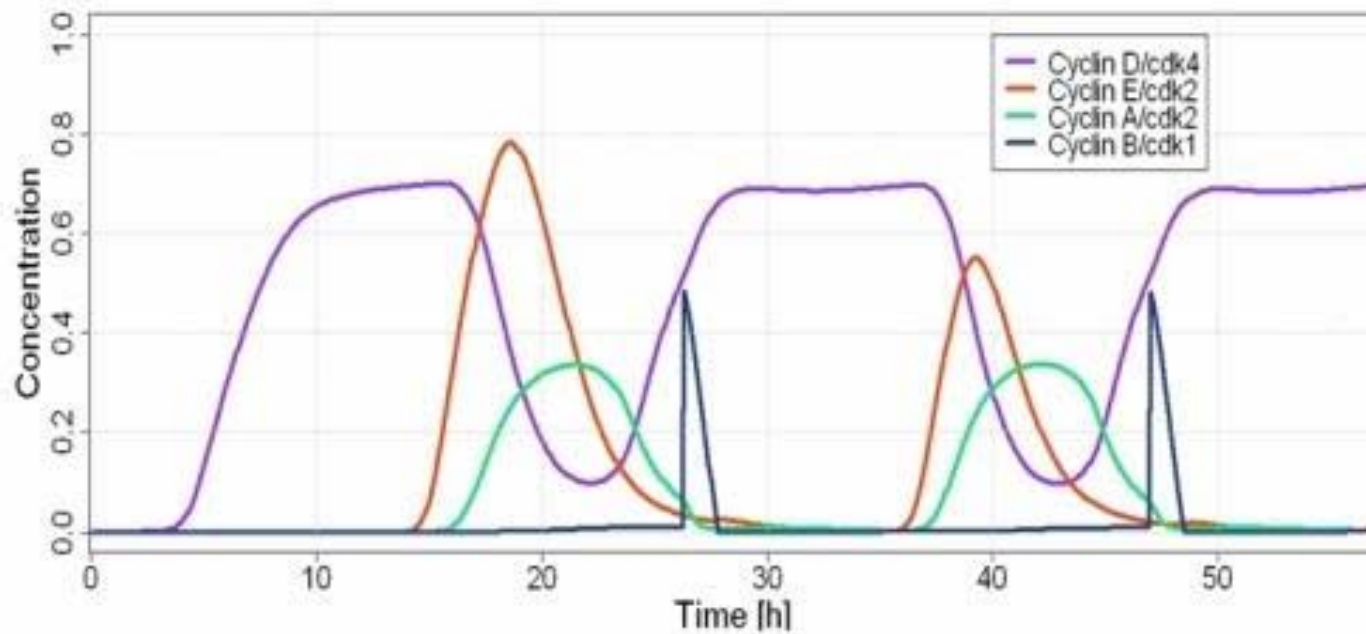
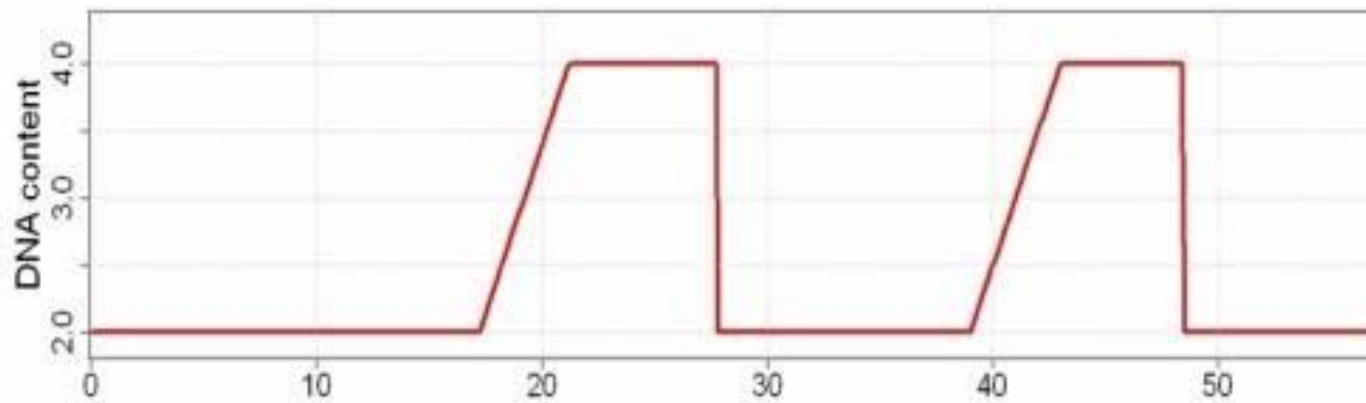
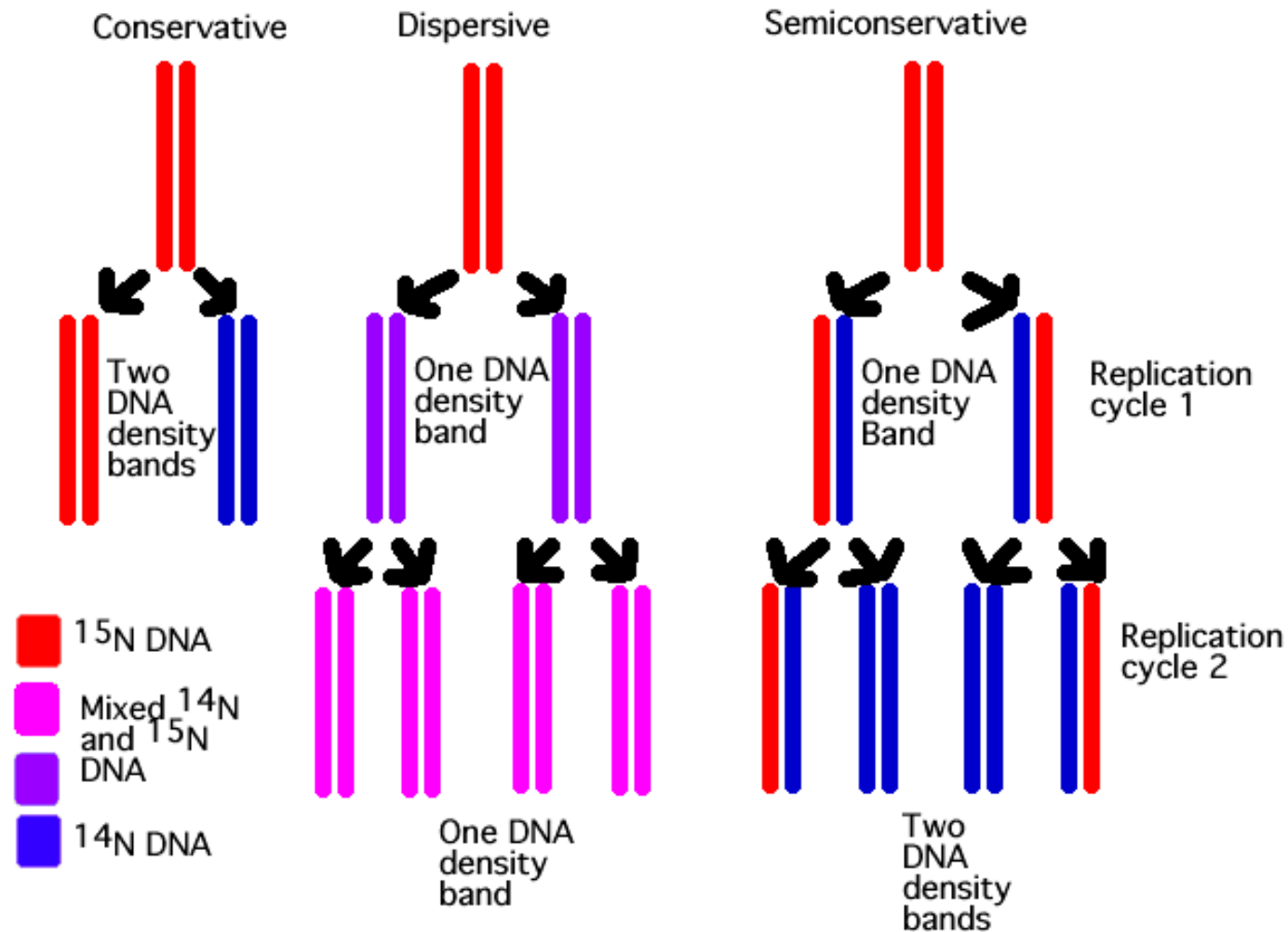


Figure 1 - Schematic representation of cell cycle control in plants. Progression through the mitotic cycle involves the successive formation, activation and subsequent inactivation of cyclin-dependent protein kinases (CDKs). The kinases bind sequentially to a series of cyclins, which are responsible for differential activation of the kinase during the cell cycle. The G1 to S transition is thought to be controlled by CDKs containing D-type cyclins that phosphorylate the retinoblastoma protein, releasing E2F transcription factors. E2F are involved in the transcription of genes needed for the G1 to S transition. The G2 to M transition is carried by CDK complexes containing CycA and CycB cyclins. CDK complexes are kept in inactive state by phosphorylation by the Wee1 kinase, and by interaction with inhibiting proteins (CKIs). At the G2 to M boundary activation of the kinase is brought about by release of the CKI protein, by positive phosphorylation (by CAK kinase), and by a still unidentified protein phosphatase.

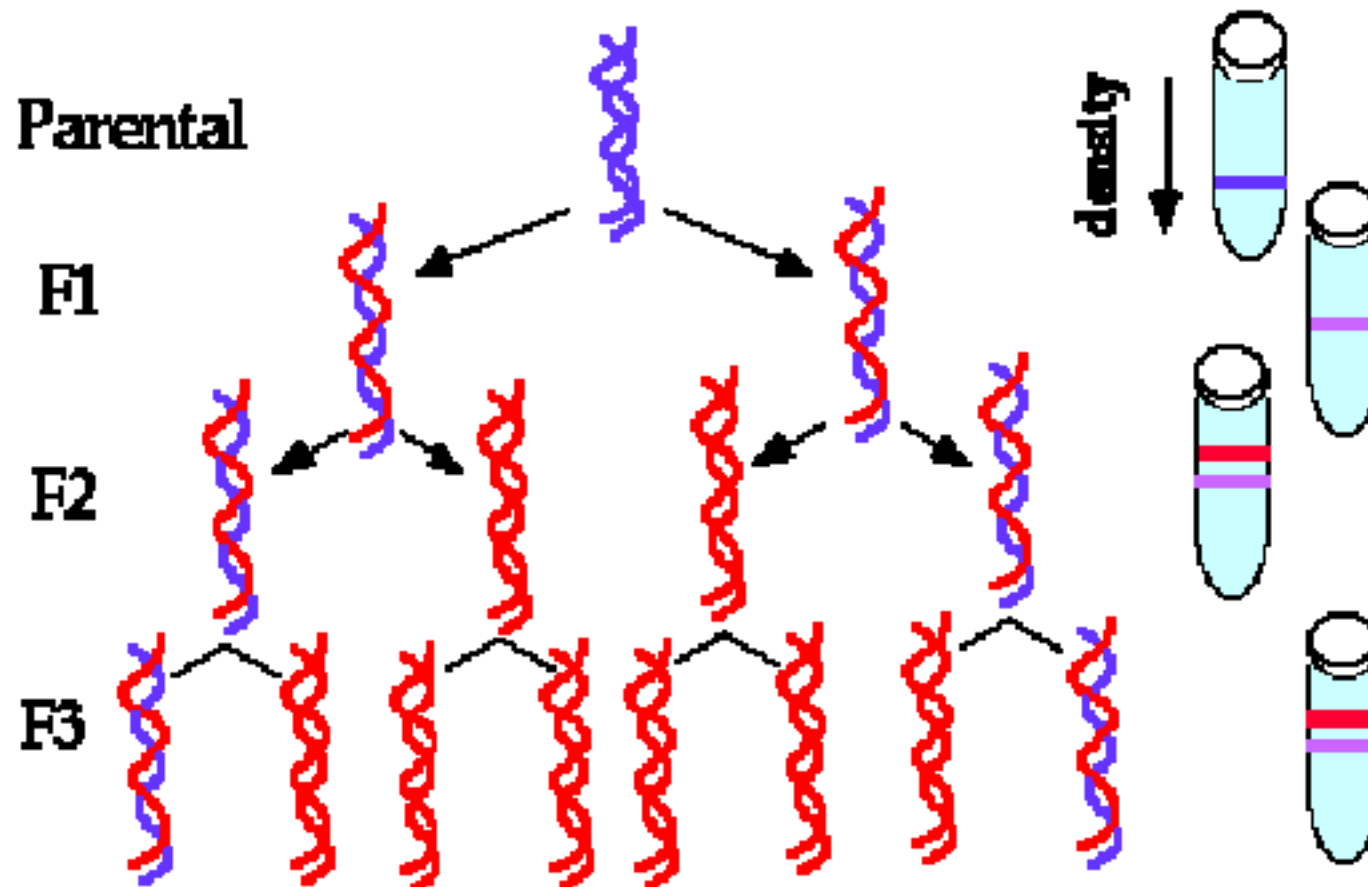
Steuerung des Zellzyklus durch Cycline



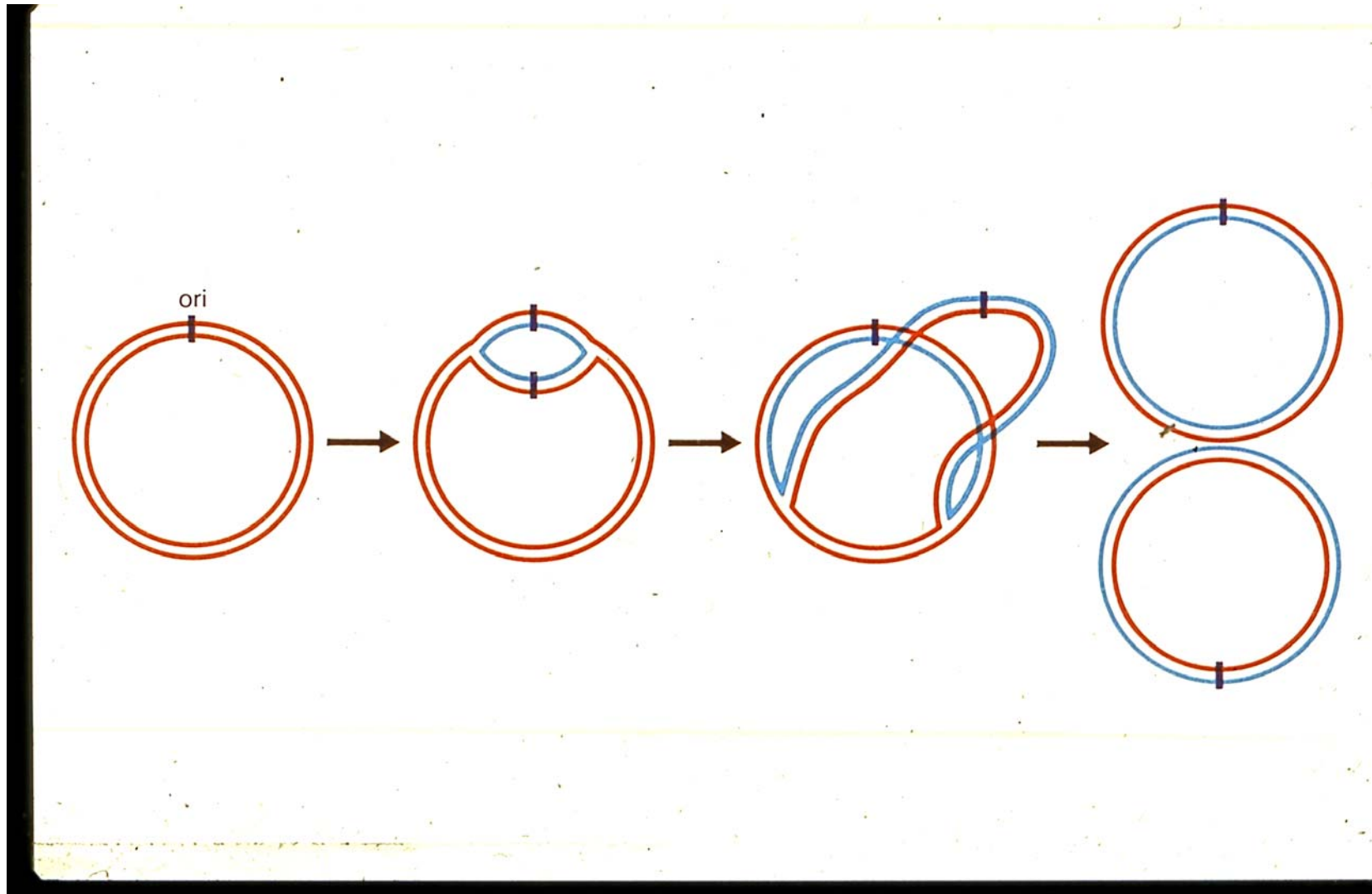
Die DNA-Replikation verläuft semi-konservativ, bewiesen durch Meselson und Stahl, Nobelpreis 1962



**Die DNA-Replikation verläuft
semi-konservativ,
bewiesen durch Meselson und
Stahl, Nobelpreis 1962**

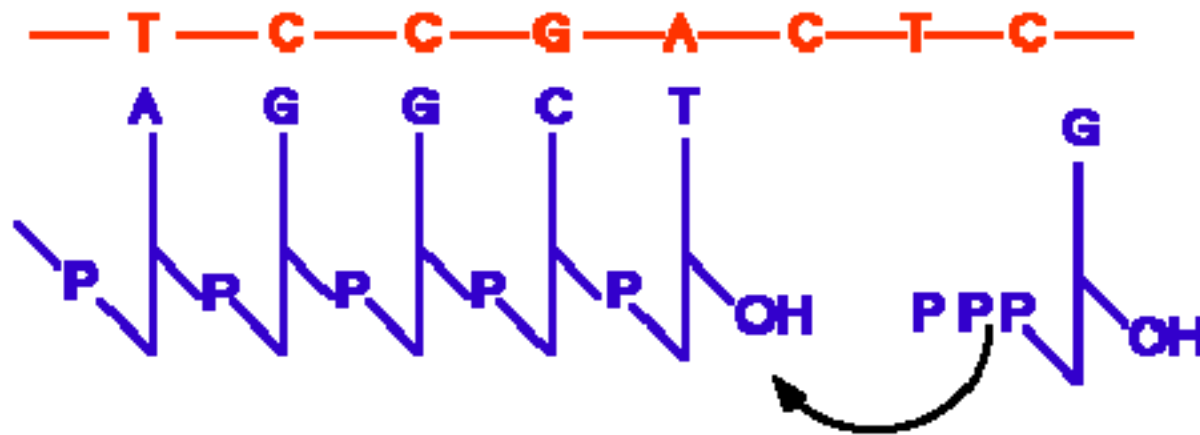


Die DNA-Replikation verläuft normalerweise „bidirektional“

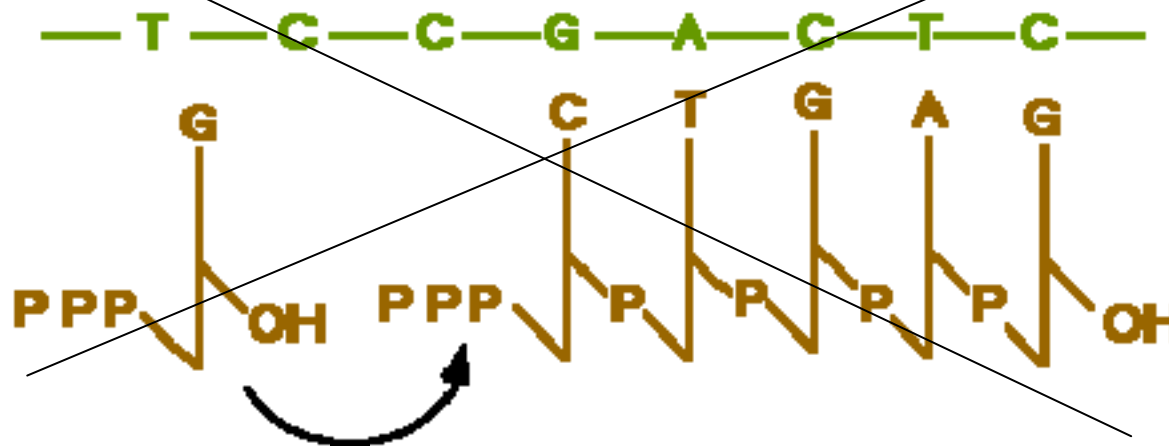


Die DNA-Synthese läuft immer 5' - 3' ab

5' to 3' synthesis



3' to 5' synthesis



DNA-Replikation

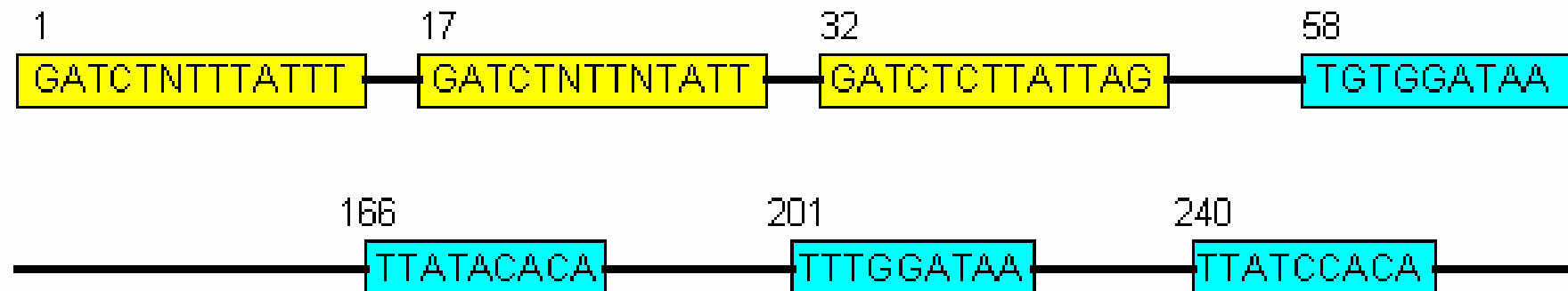
- Ein Prozess in drei Stufen
 1. Initiation
 2. Elongation
 3. Termination

Die DNA-Replikation beginnt an
einem speziellen
Sequenzelement:

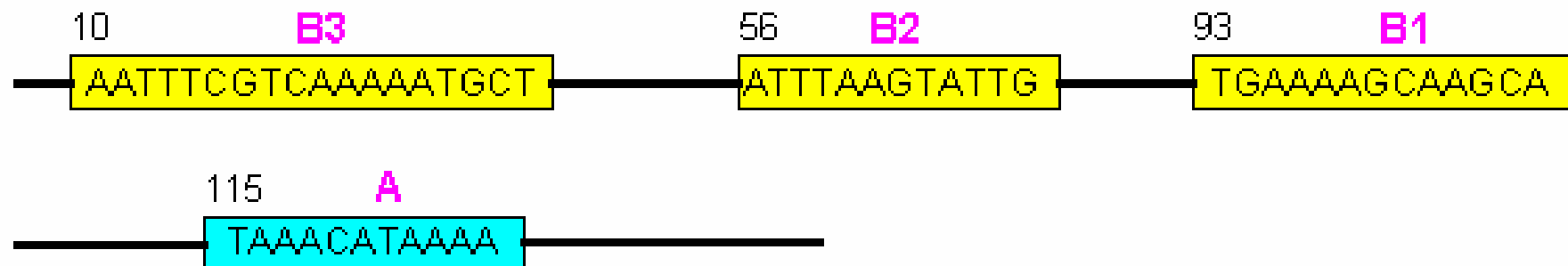
„origin of replication“
(ori)
(Replikationsursprung)

Sequenzen bekannter Origins

(a) Bacterial replication origin



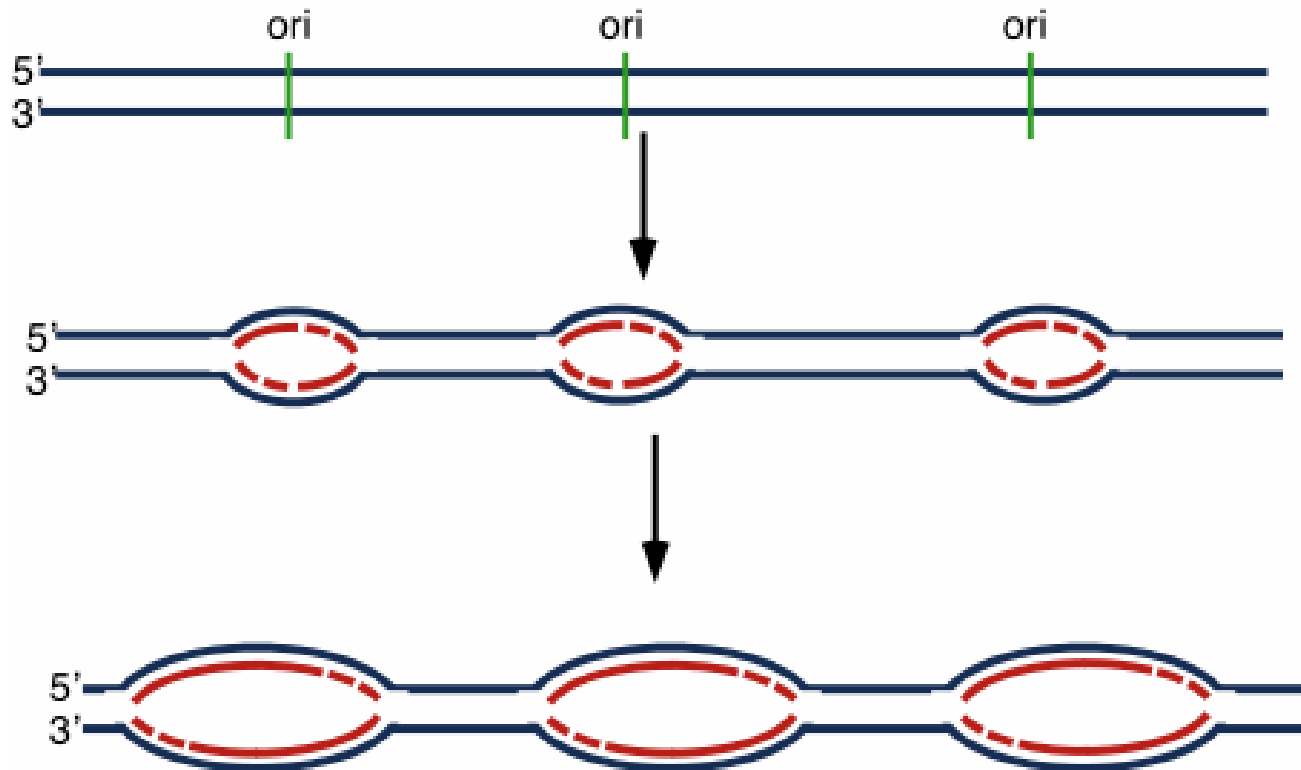
(b) Yeast replication origin (ARS1)



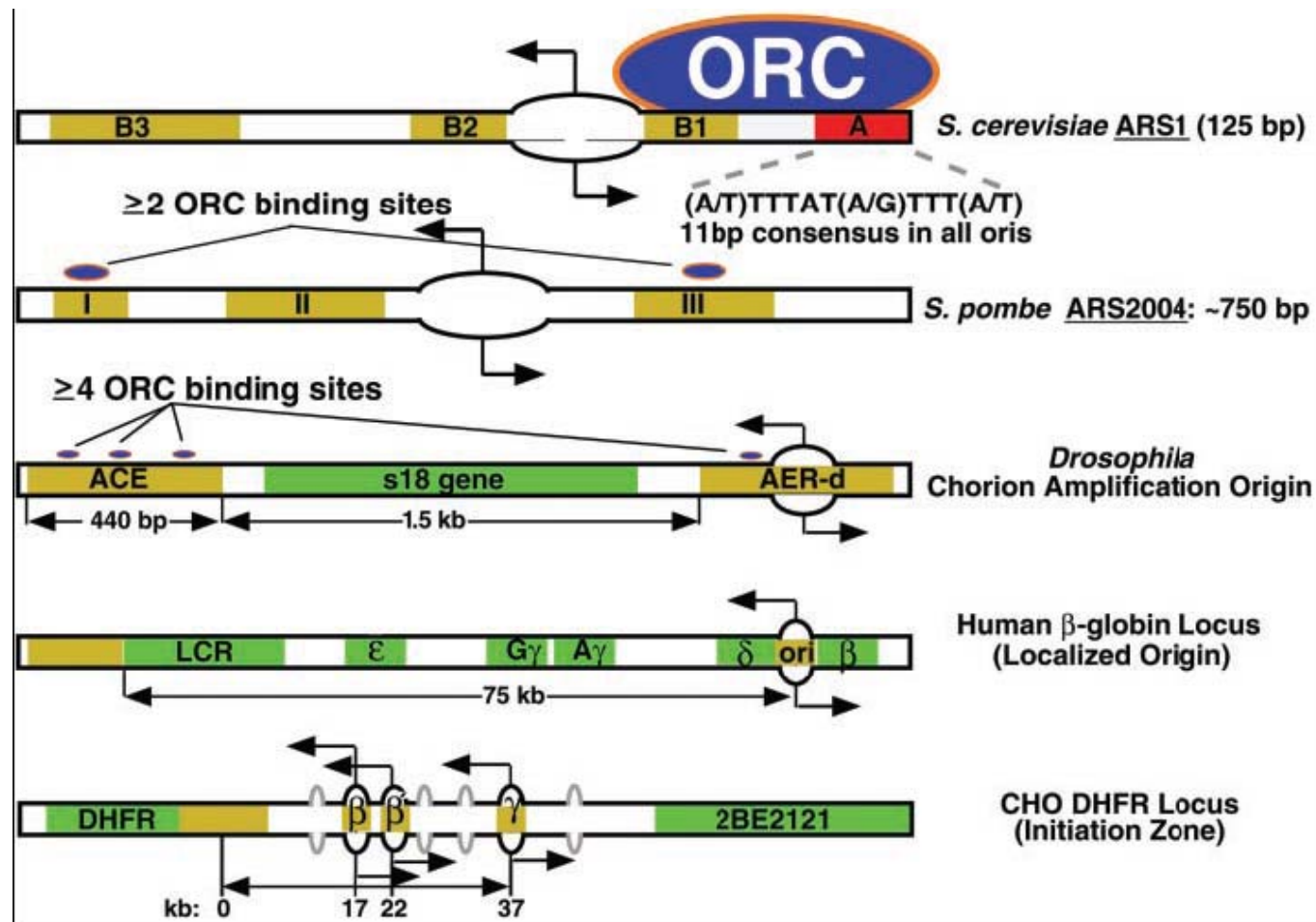
**Prokaryoten besitzen meist nur einen
„vegetativen“ Origin of Replication;**

Eukaryoten haben dagegen multiple Origins

**Large eukaryotic DNA has
multiple origins of replication**

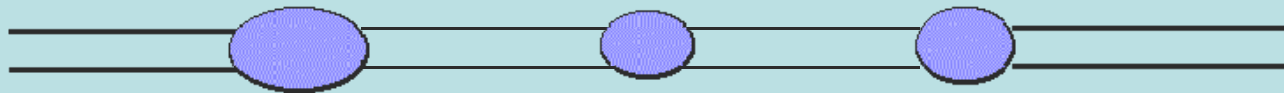


Eukaryotische Replikationsursprünge

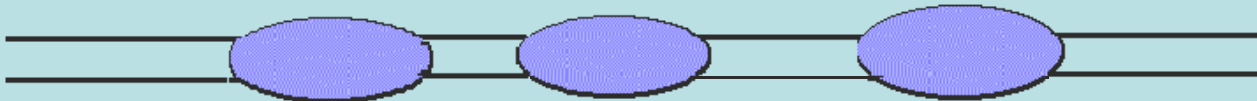


SCIENCE VOL 294 5 OCTOBER 2001

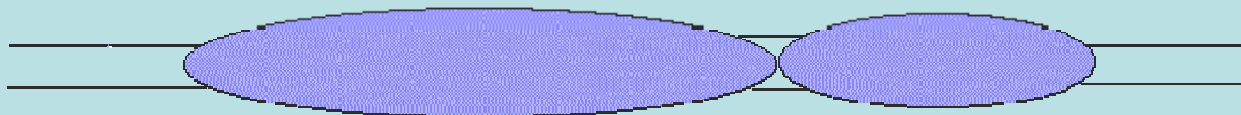
Replication bubbles begin at different points along the DNA helix.



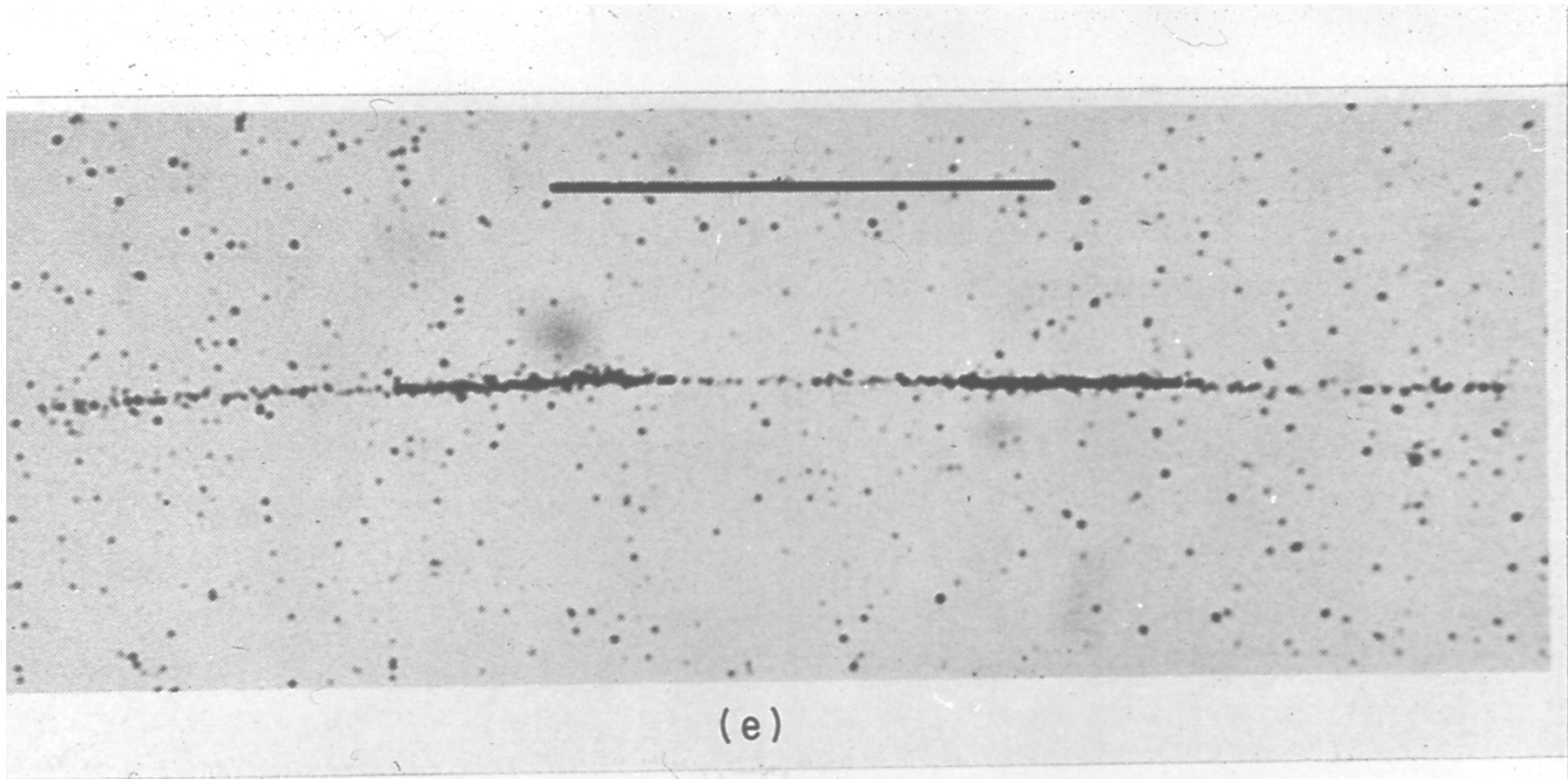
The replication bubbles are "growing" as the replication forks proceed in opposite directions.



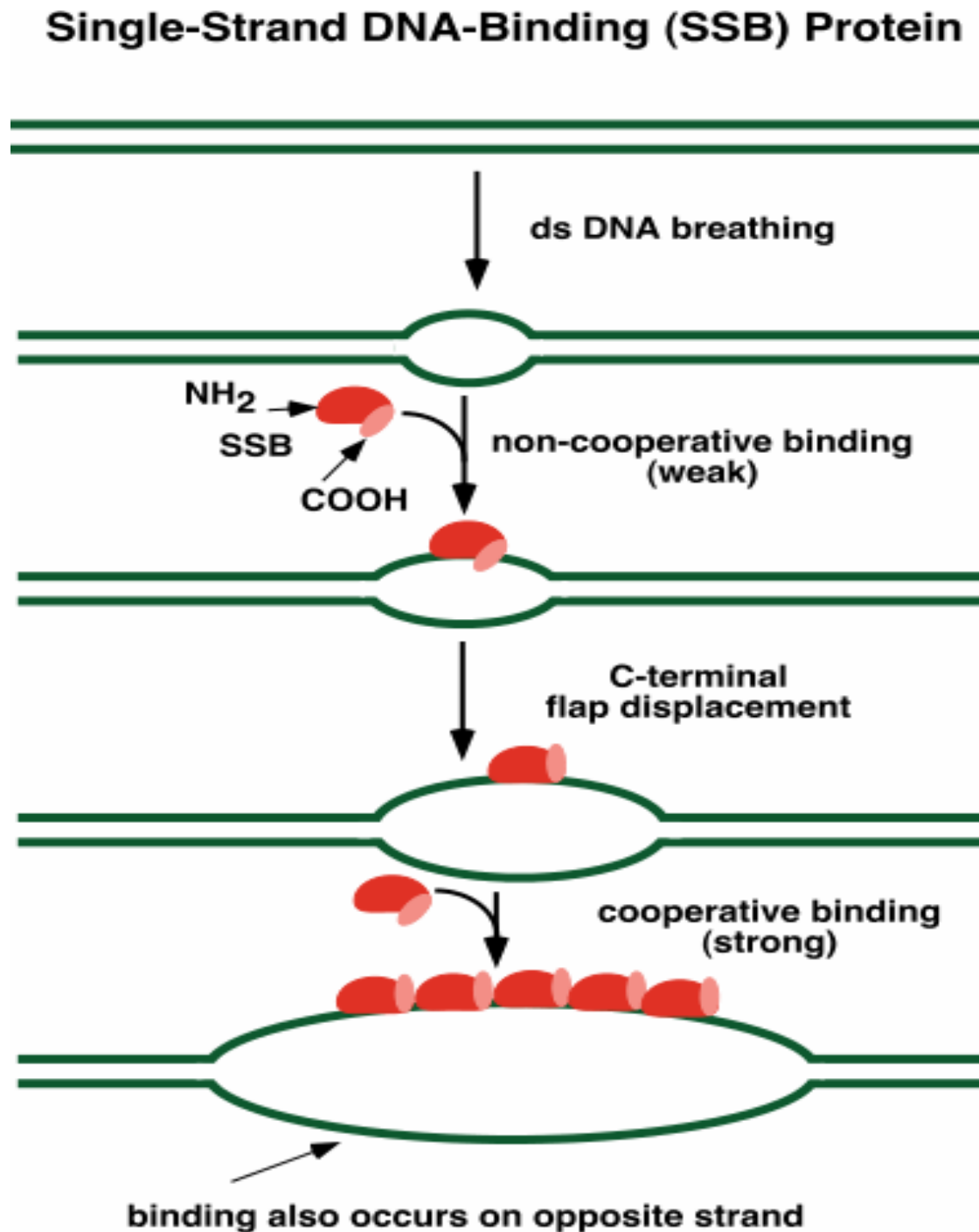
Eventually, the replication bubbles join together as the entire DNA helix is replicated.

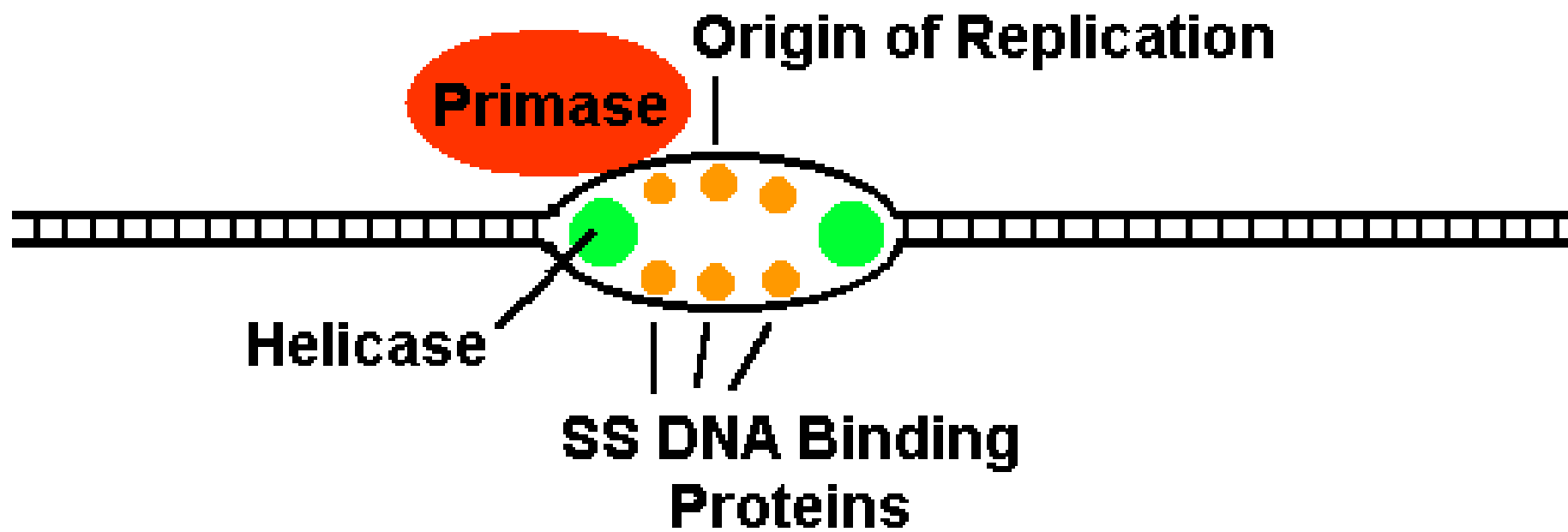


Nachweis von Replikationsstartpunkten (Oris) durch Fiber-Autoradiografie

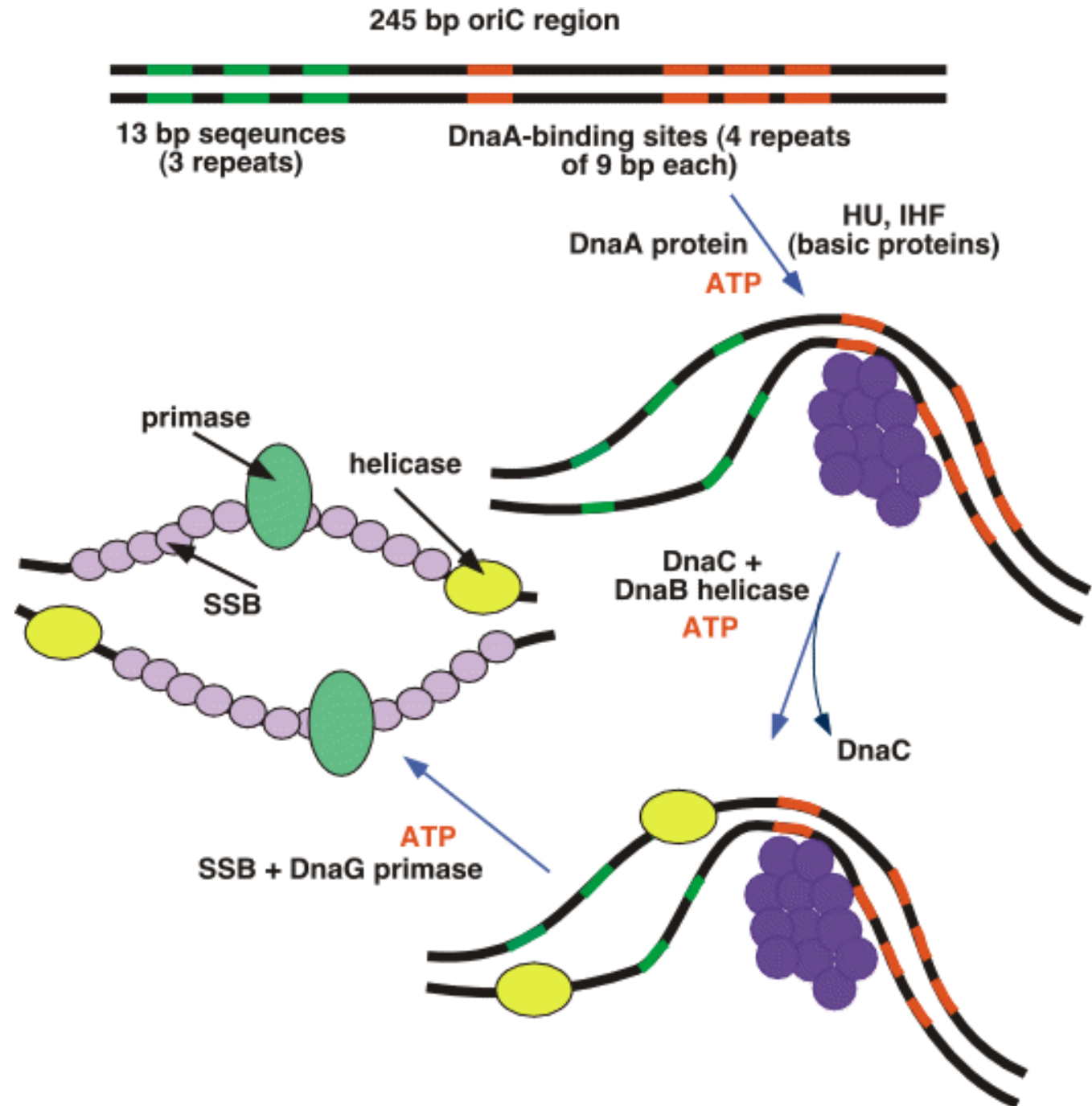


Initiation

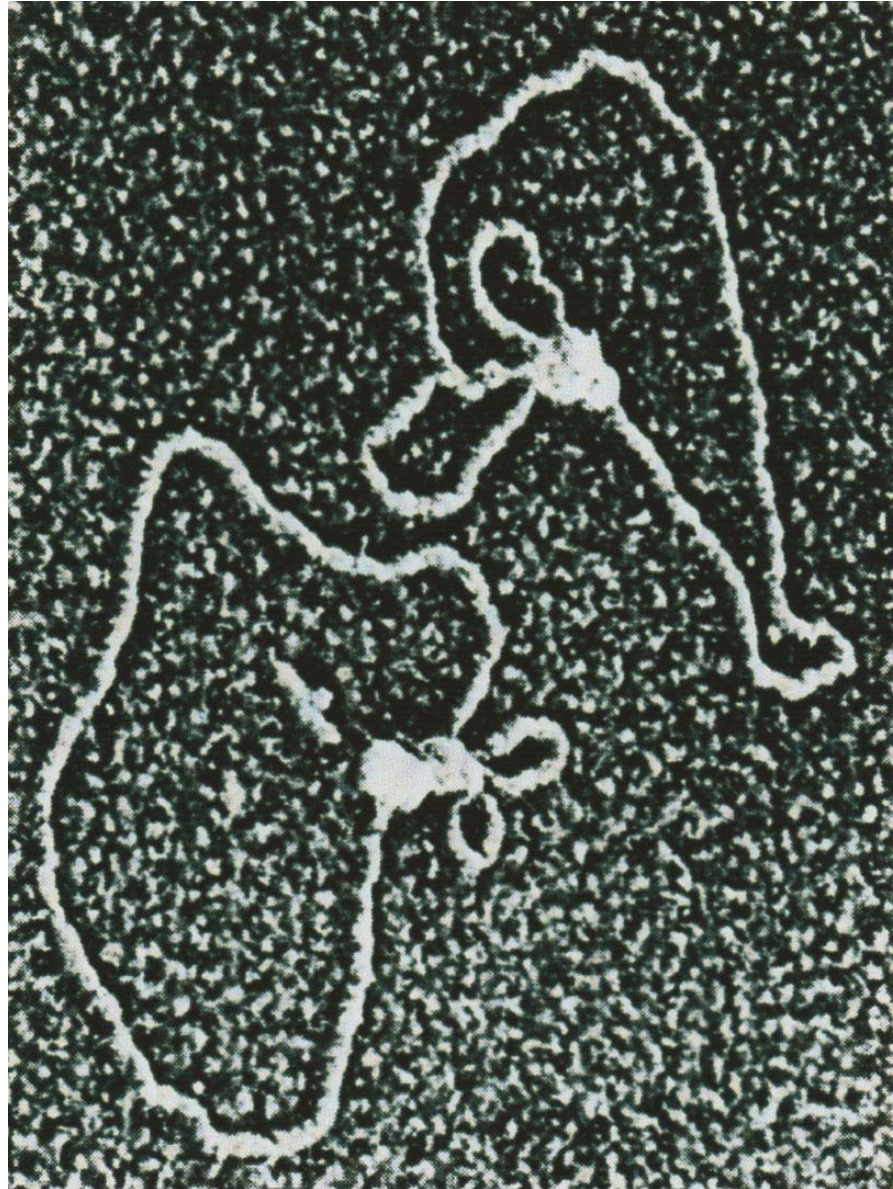




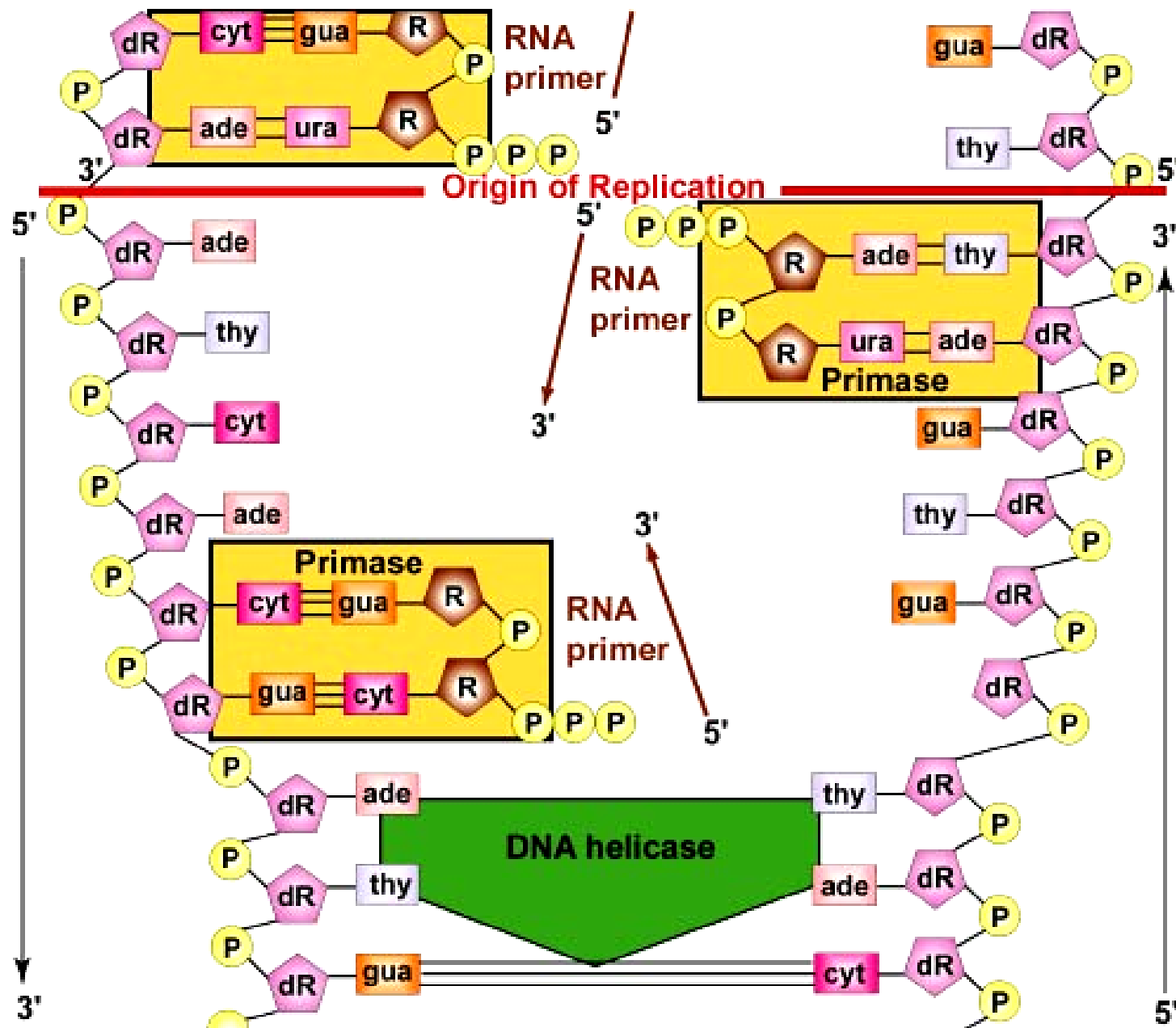
Initiation

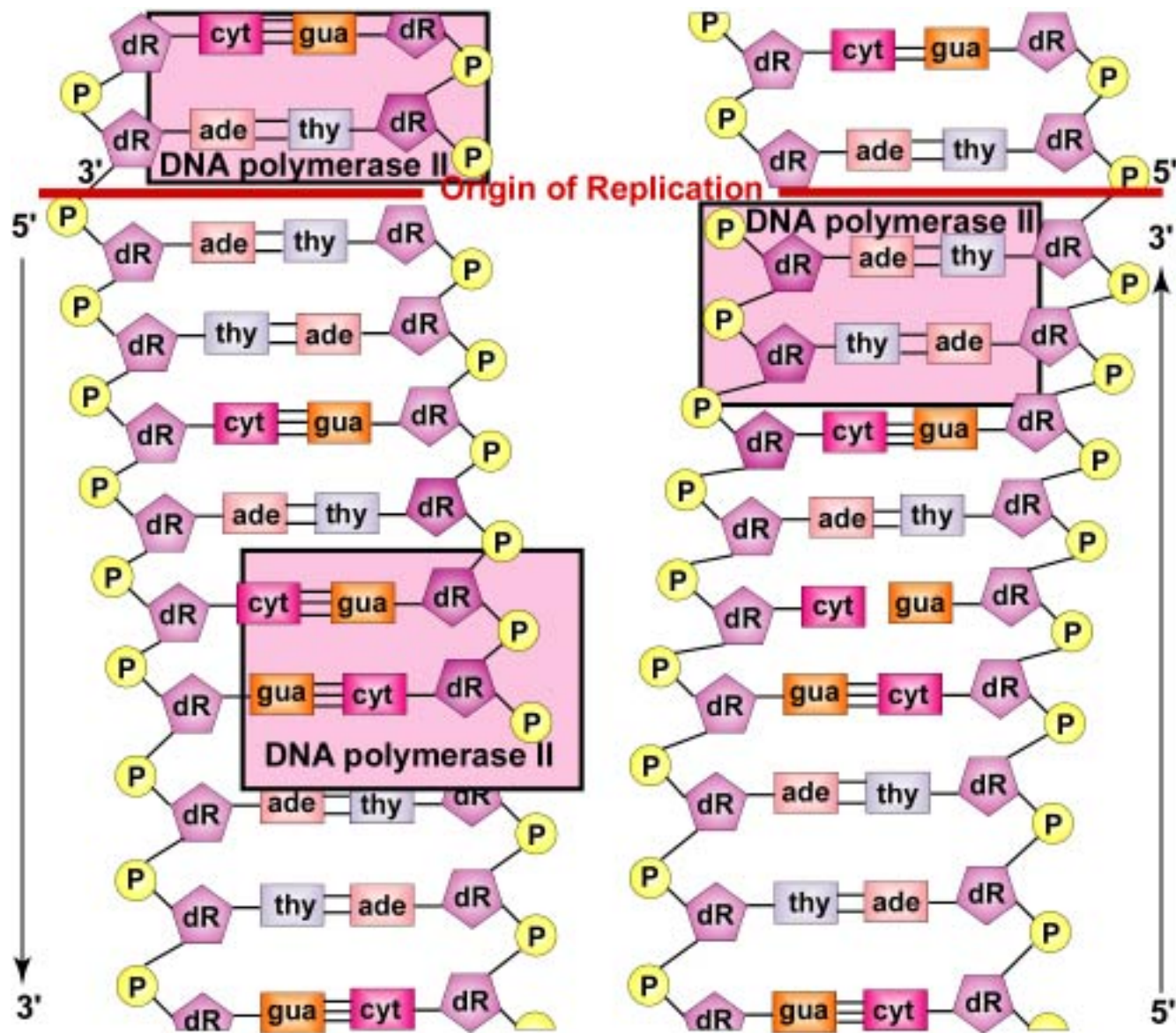


Initiationskomplex bei PhiX-Phage

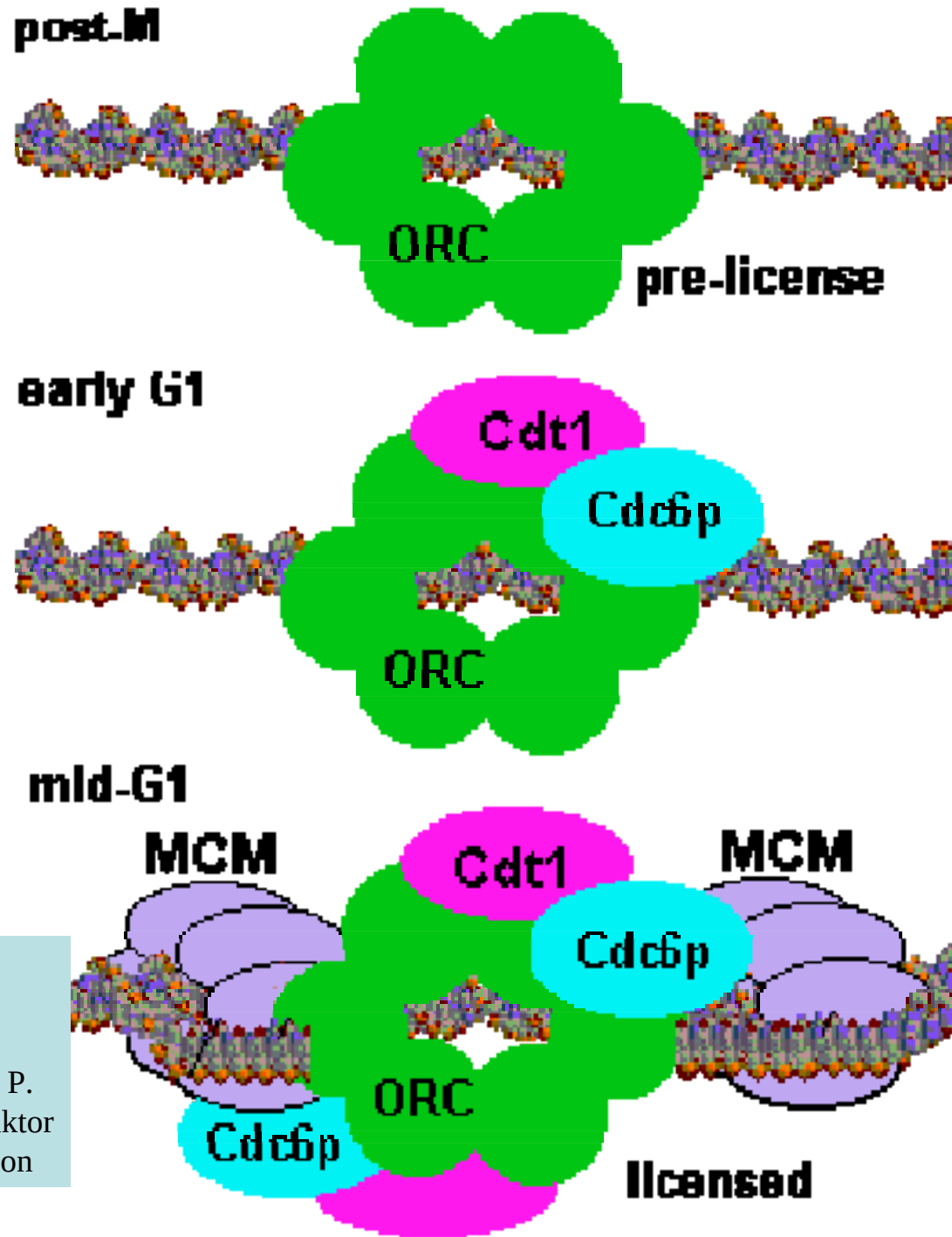


1. Syntheseschritt: Synthese des „Primer“ durch Primase





Die **Initiation** der
DNA-Replikation
bei **Eukaryoten**
am ori erfolgt erst
nach der
„Lizensierung“
durch ORC und
weitere Proteine



ORC= Origin recognition complex
Cdk= Cyclin dependent kinase
Cdc= Cell division cycle Protein
Mcm= Minichromosome maintenance P.
Cdt1= Replikationsfaktor/LicensingFaktor
OBR= Origin of bidirectional replication

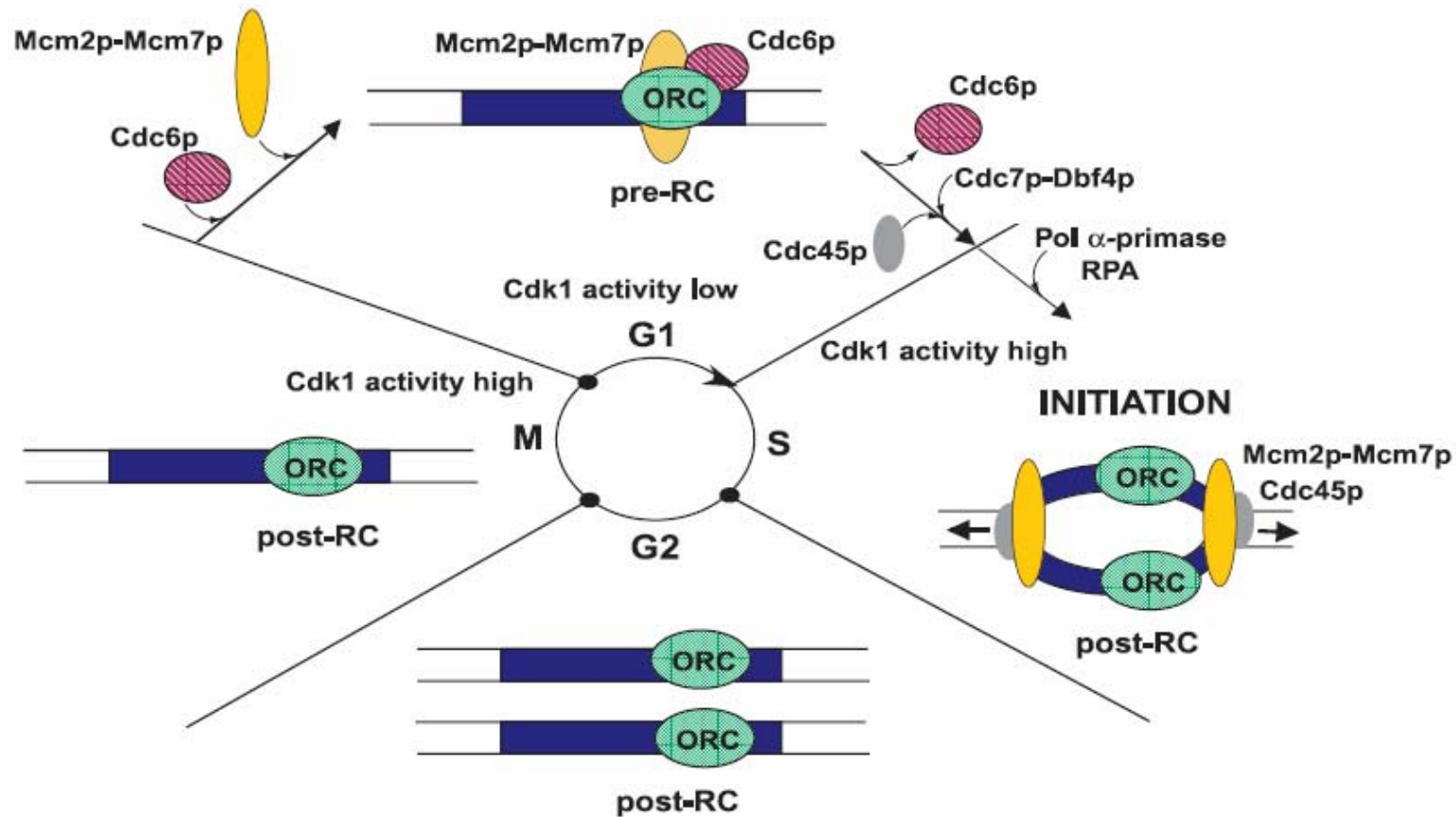
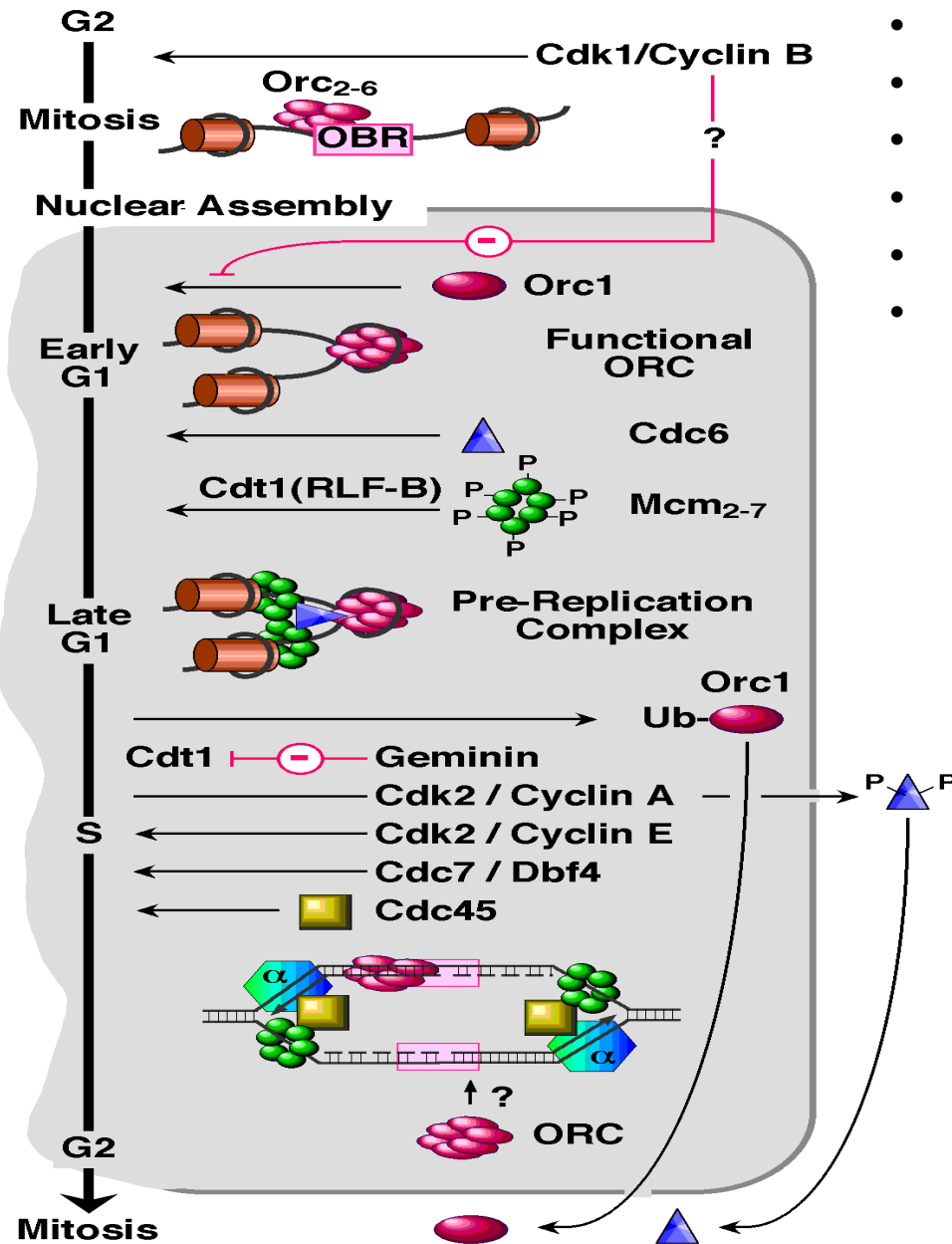


Fig. 1. Events leading to origin activation in budding yeast. Origin recognition complex (ORC) binds to an ARS element in yeast (blue rectangle). For the stepwise assembly of the pre-RC during G₁ phase when Cdk1p activity is low, ORC recruits Cdc6p, which in turn loads Mcm2p-Mcm7p. When Cdk1p activity rises at the G₁/S transition, the pre-RC is disassembled. The post-RC remains stable until the end of mitosis, and owing to high Cdk1p activity the pre-RC cannot re-associate during this time but must await the subsequent G₁ phase.



- ORC= Origin recognition complex
- Cdk= Cyclin dependent kinase
- Cdc= Cell division cycle Protein
- Mcm= Minichromosome maintenance P.
- Cdt1= Replikationsfaktor
- OBR= Origin of bidirectional replication

Initiator-Proteine

Table 2. Initiation Proteins

Function	<i>E. coli</i>	Phage λ	Phage T4	SV40/Human	Yeast
Initiator protein	DnaA	λ O	none	T antigen	ORC
Loading and remodeling factor(s)	DnaC	λ P, DnaJ, DnaK	gp59	Cellular chaperone?	Cdc6 protein
DNA helicase	DnaB	DnaB	gp41	T antigen	MCM proteins?