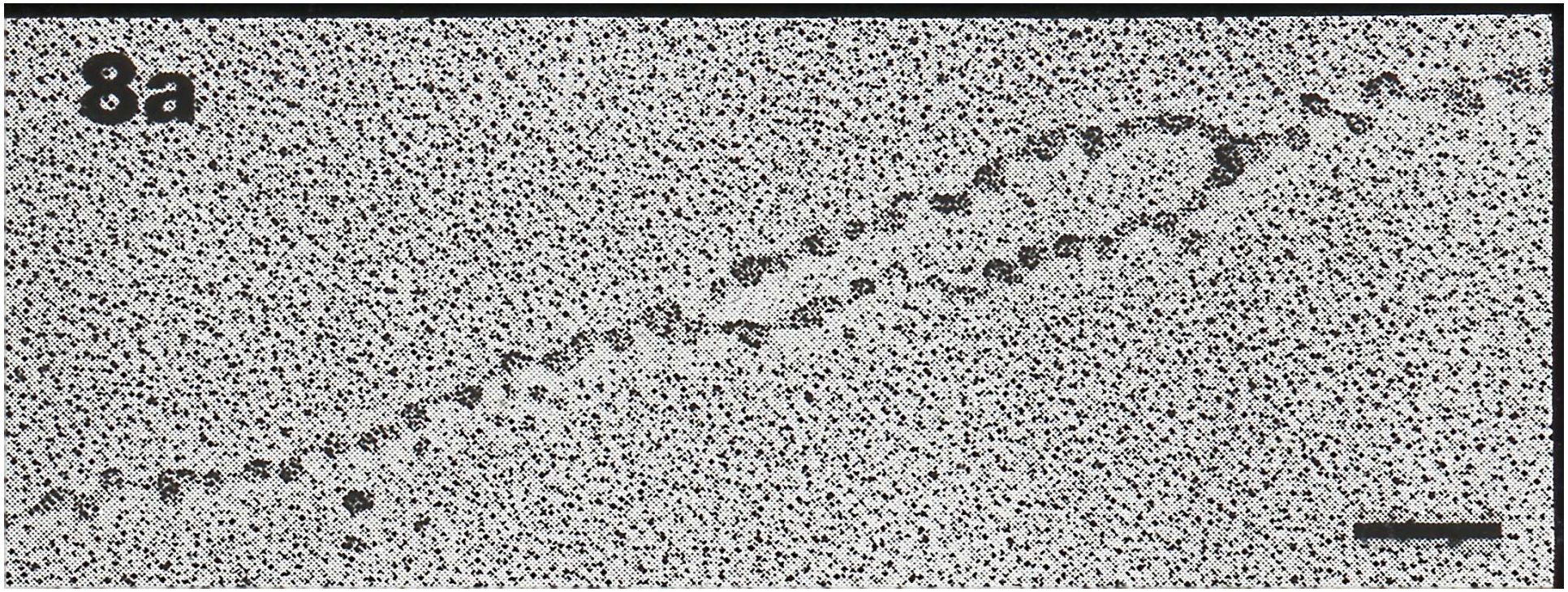
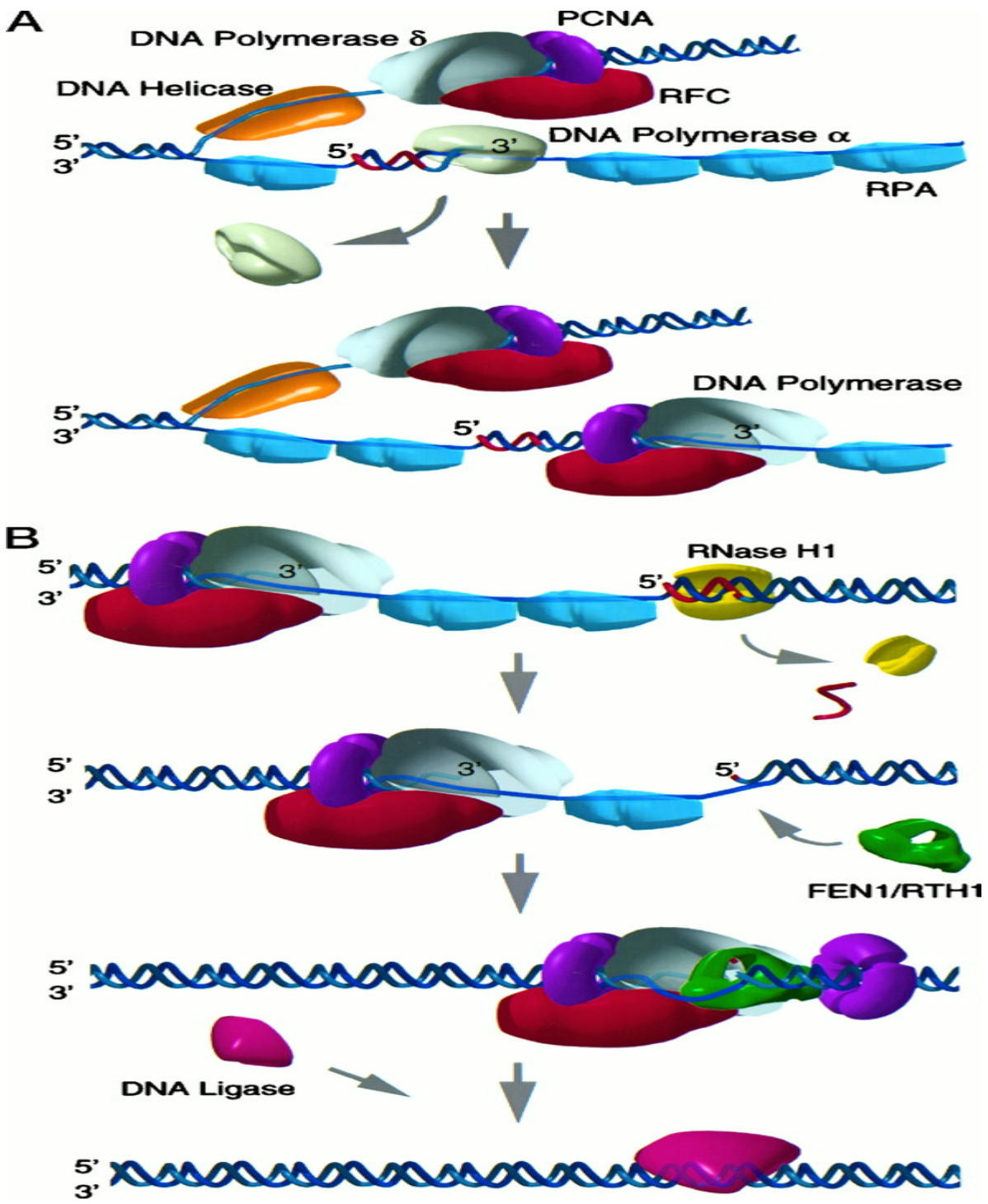
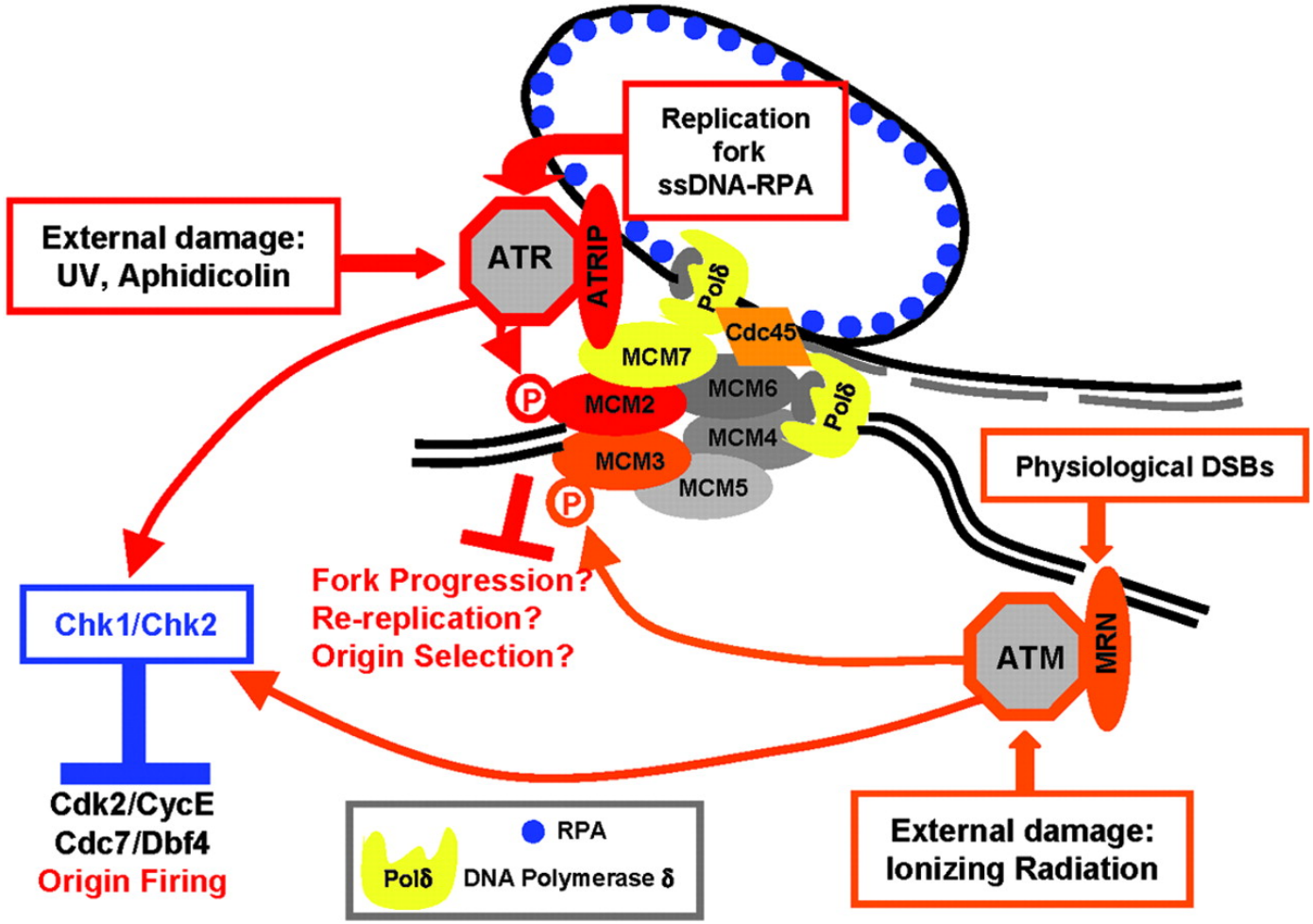
Bei Eukaryoten ist die DNA während der Replikation als Chromatin verpackt



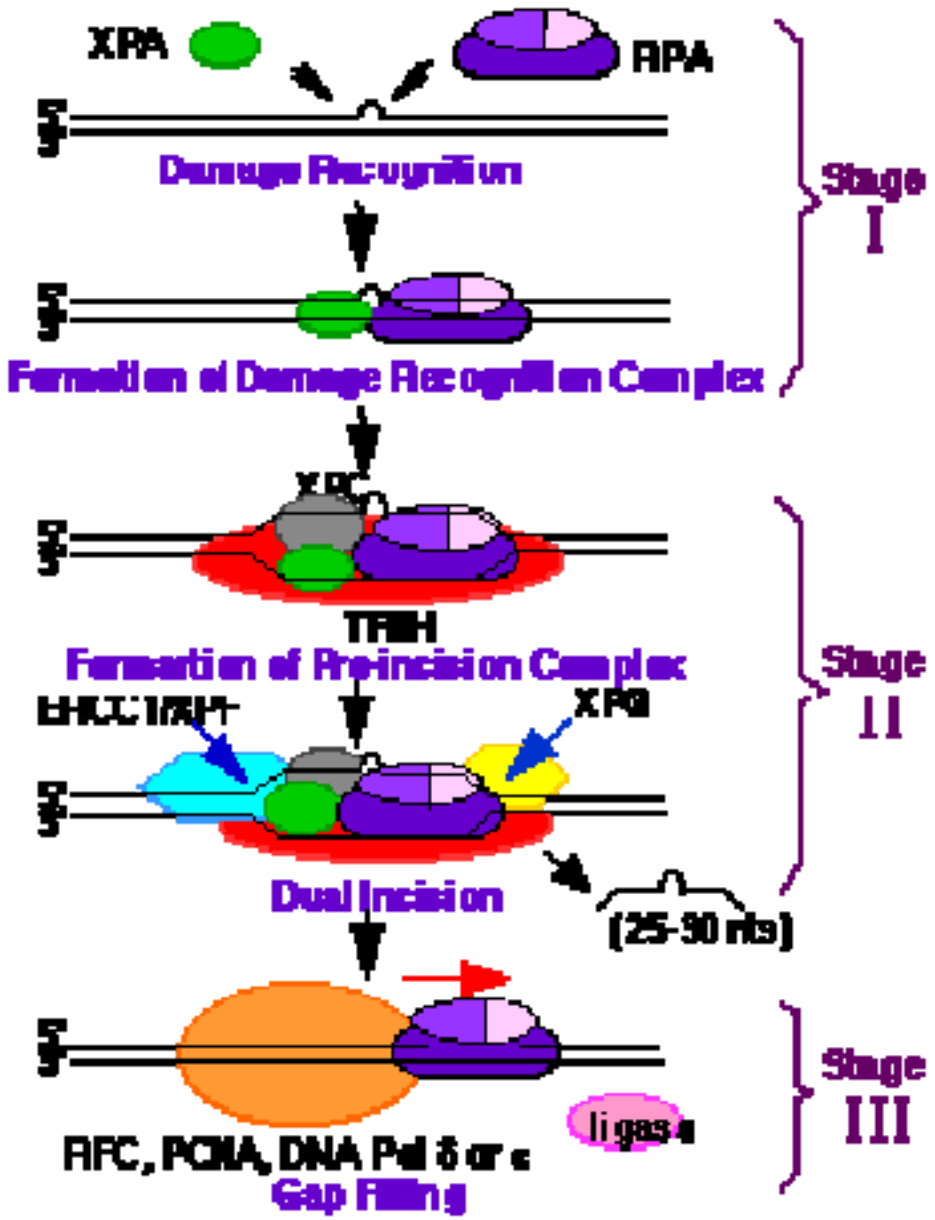

A
PCNA
DNA Polymerase δ
DNA Helicase
RFC
5'
3'
5'
3'
DNA Polymerase α
RPA
DNA Polymerase
5'
3'
5'
3'
B
RNase H1
5'
3'
3'
5'
S
5'
3'
3'
5'
FEN1/RTH1
5'
3'
DNA Ligase
5'
3'

Protein	Funktion 1	Literaturlink
DNA-Polymerase α	DNA-Synthese „lagging strand“	
DNA-Polymerase δ	DNA-Synthese „leading stand“	
ORC	Erkennung/Aktivierung Ori: Assembly of preRC	
Cdt	Licensing protein; oncogene	
RFC	Clamp loader of PCNA	http://cat.inist.fr/?aModele=afficheN&cpsidt=15405201
RPA	Single stranded binding protein	http://nar.oxfordjournals.org/cgi/content/full/34/15/4126
MCM	Origin assambly factor; später Helikase	http://www-rcf.usc.edu/~forsburg/MCM.html
PCNA (=Cyclin)	Processivity factor of DNA-Pol delta (stimul. Repl.>10x)	http://www.ncbi.nlm.nih.gov/entrez/dispomim.cgi?id=176740
RNAse HI	Primerentfernung	http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=84926
FEN1	„flap endonuclease“; Entfernung letztes Primernukleotid und 5‘ „flaps“	http://cat.inist.fr/?aModele=afficheN&cpsidt=1189753

Fig. 1. A schematic view of the signaling pathways inhibiting DNA replication. ssDNA-RPA intermediates and DSBs arise as a consequence of external insults (irradiation and polymerase inhibitors) or during normal replication



Shechter, David and Gautier, Jean (2004) Proc. Natl. Acad. Sci. USA 101, 10845-10846



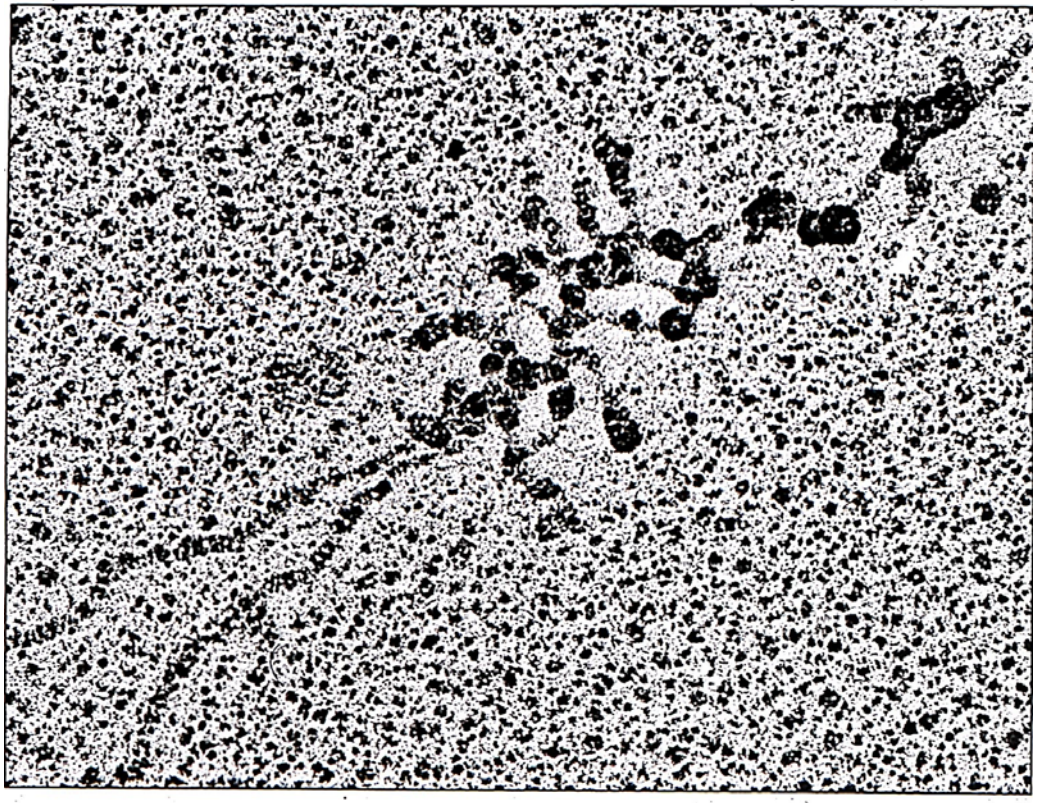
Model of Nucleotide Excision Repair (NER)

Transkription und Replikation

ASM News

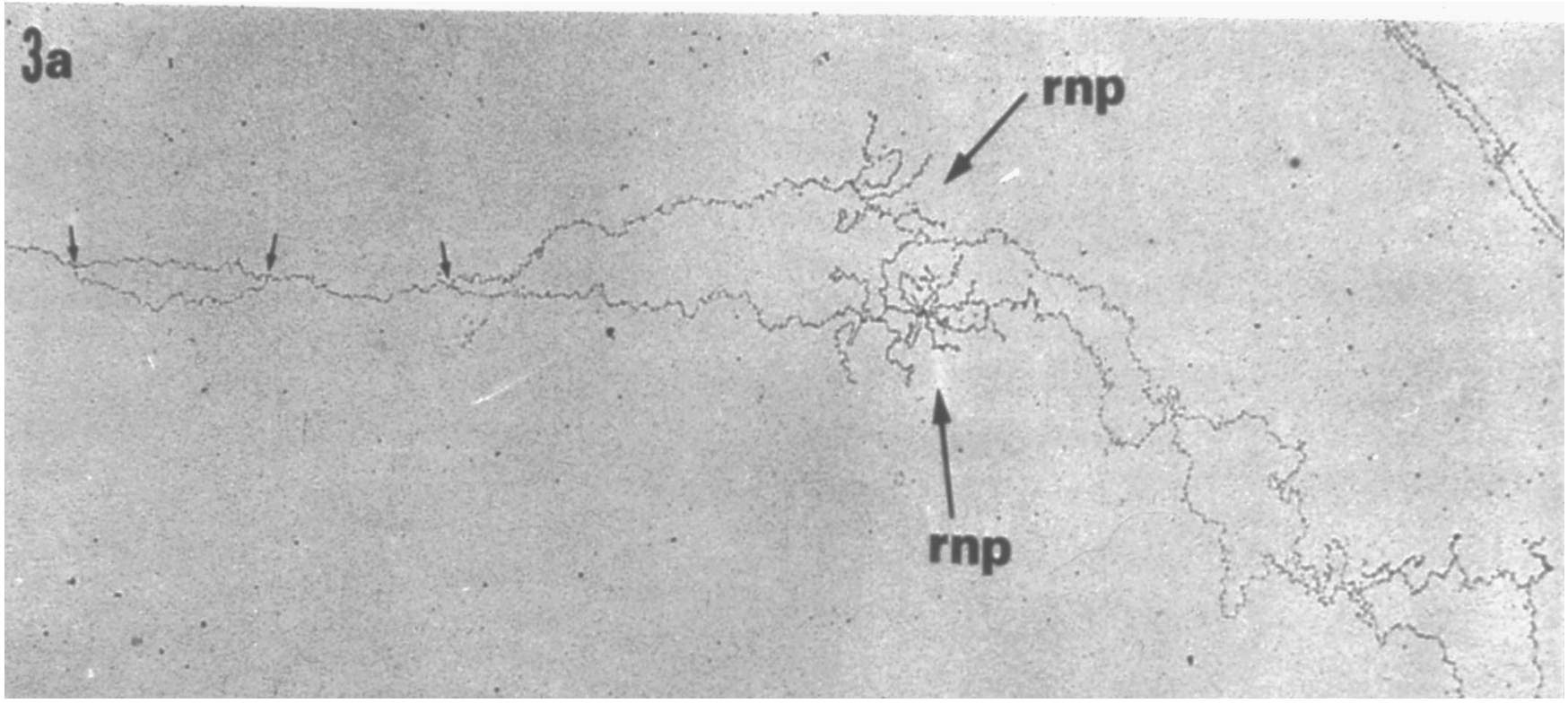
Volume 59
Number 9
September 1993

American Society for Microbiology

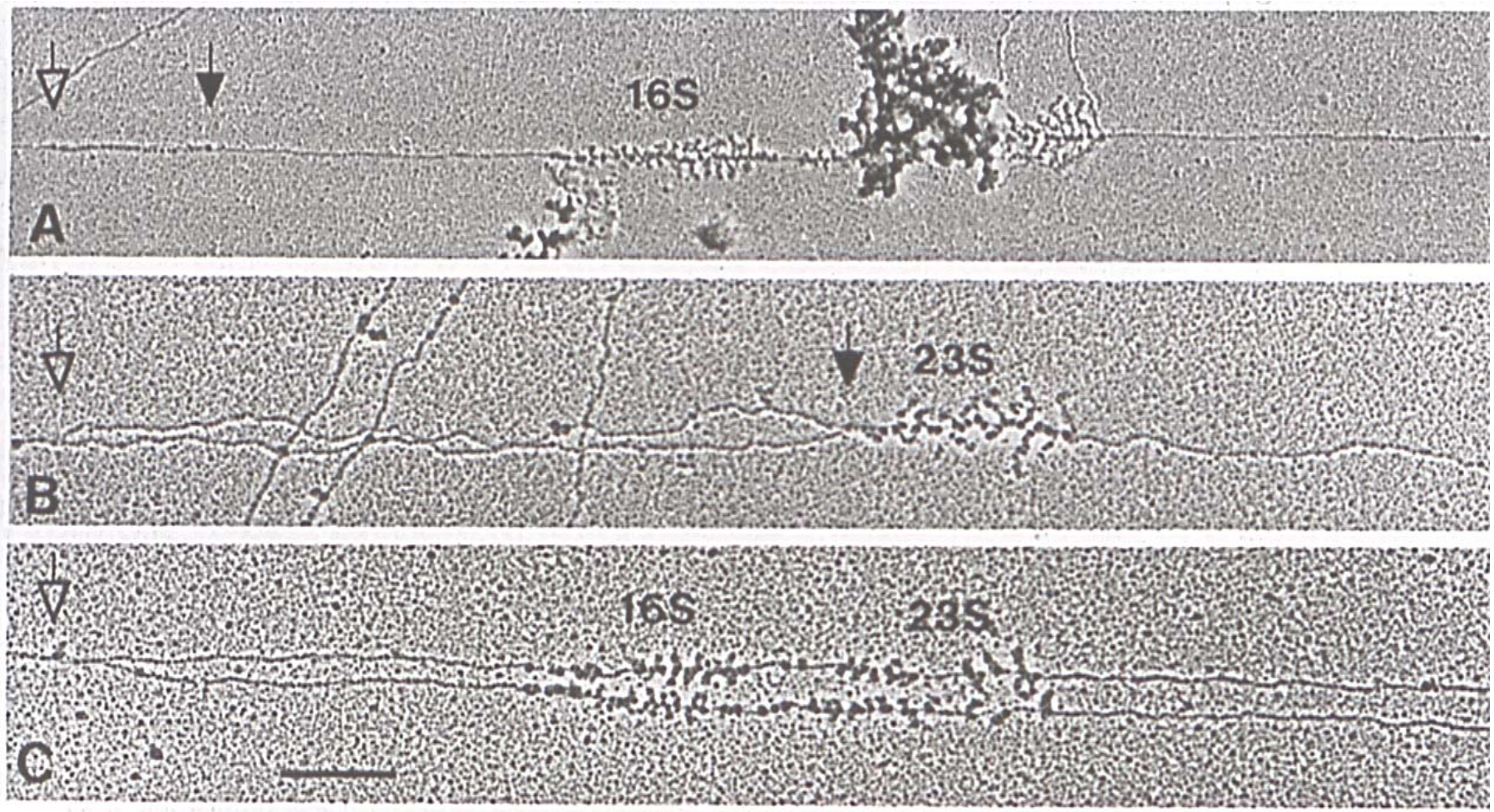


Right of Way in Replication and Transcription

Transkription und Replikation, gleichsinnig

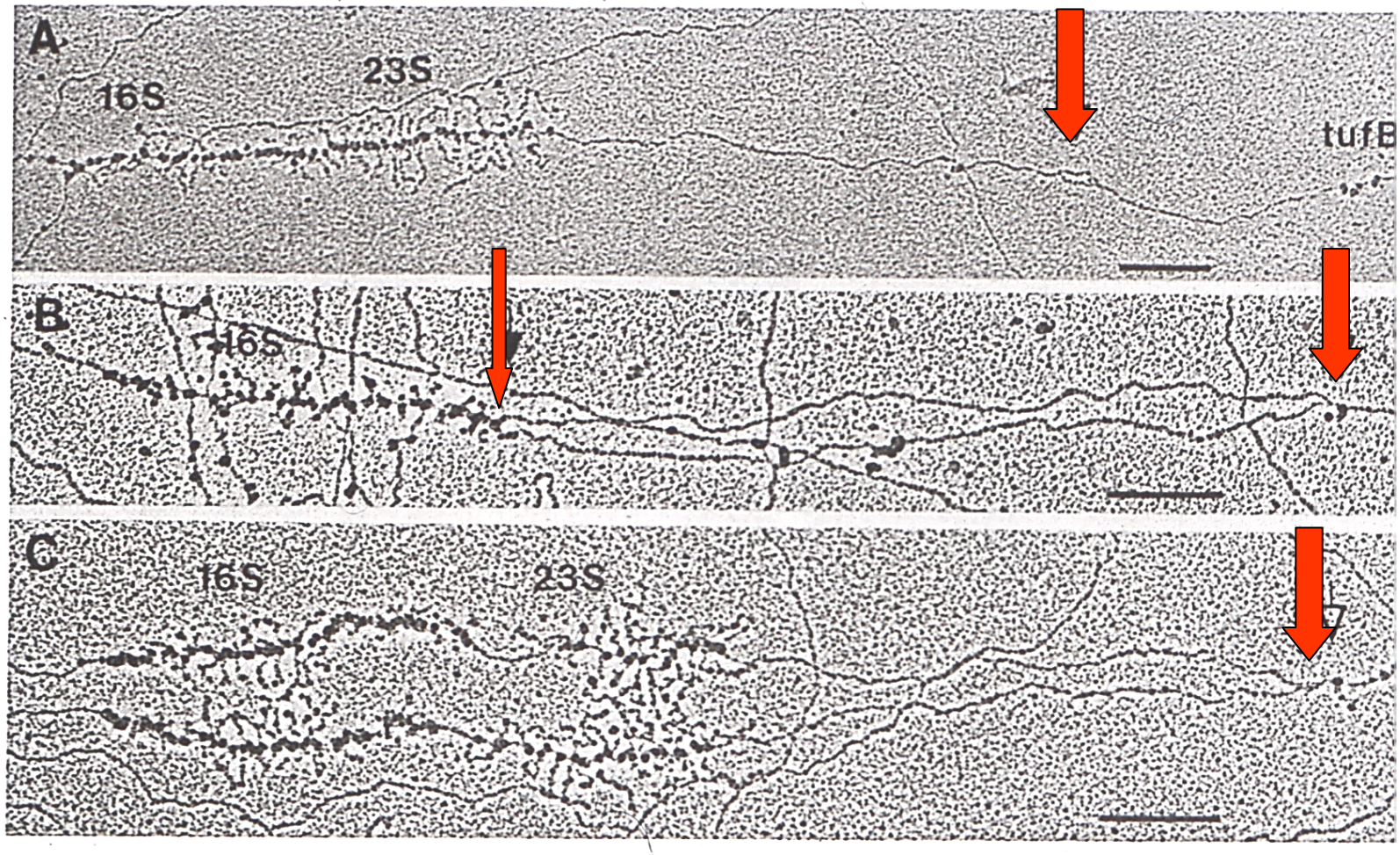


Replikation und Transkription, gleiche Richtung



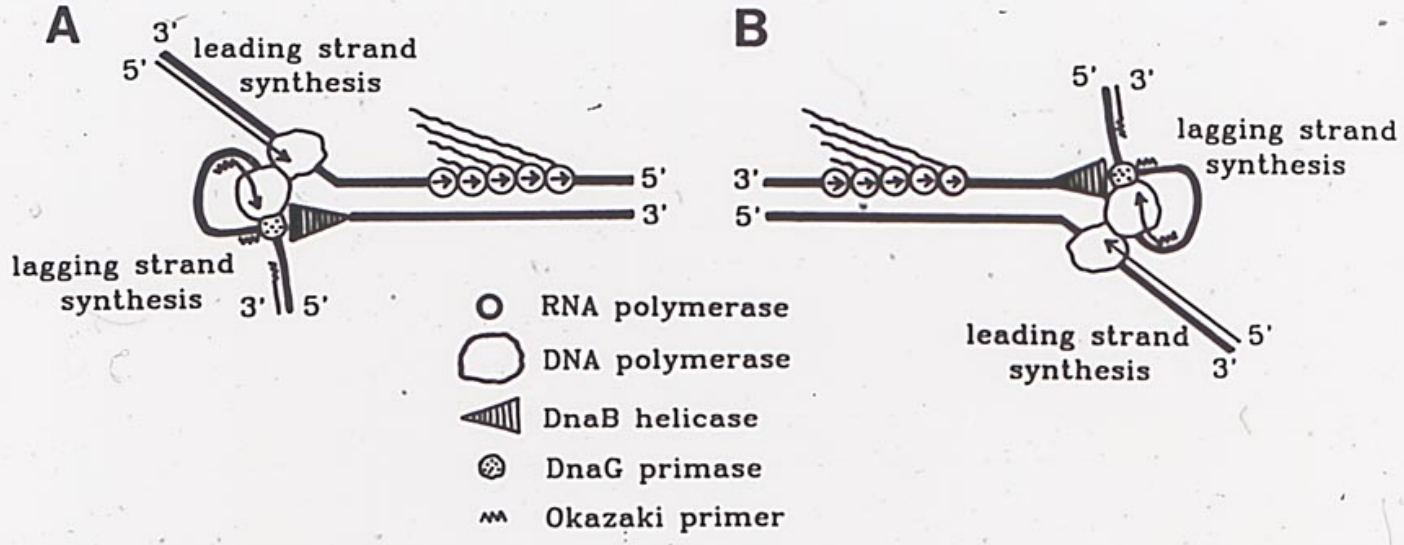
Replikation und Transkription, gegenläufige Richtung, die Transkription wird durch Replikation unterbrochen

Figure 5. CF95: Micrographs of Oppositely Oriented Replication and Transcription



Replikation und Transkription

Figure 1. Relative Orientation of Replication and Transcription Affects the Subunits Impacted When DNA and RNA Polymerases Collide

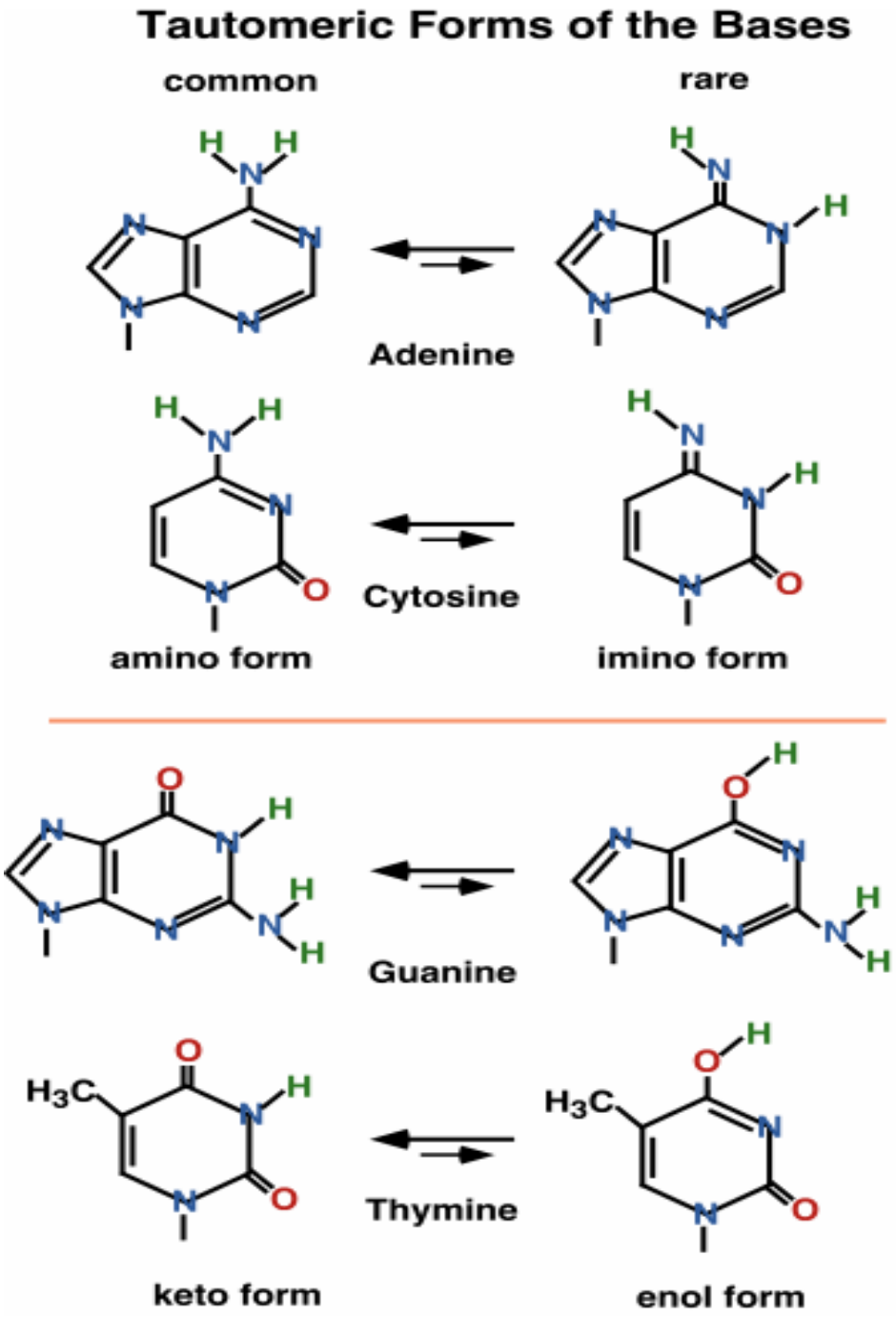


Replication forks are drawn in the asymmetric dimer configuration for coupled leading- and lagging-strand DNA synthesis. Thick lines, parental strands of the DNA duplex; thin lines, newly synthesized DNA. RNA and DNA polymerases move along the particular DNA strand which serves as the template for their activities in a 3′-to-5′ direction. (A) Replication fork and RNA polymerases moving in the same direction. The leading-strand DNA polymerase is on the same DNA strand as the RNA polymerases. (B) Replication fork and RNA polymerases moving in opposite directions. The leading-strand DNA polymerase and the RNA polymerases are on opposite DNA strands. The DnaB helicase, moving in a 5′-to-3′ direction, is on the same strand as the RNA polymerases.

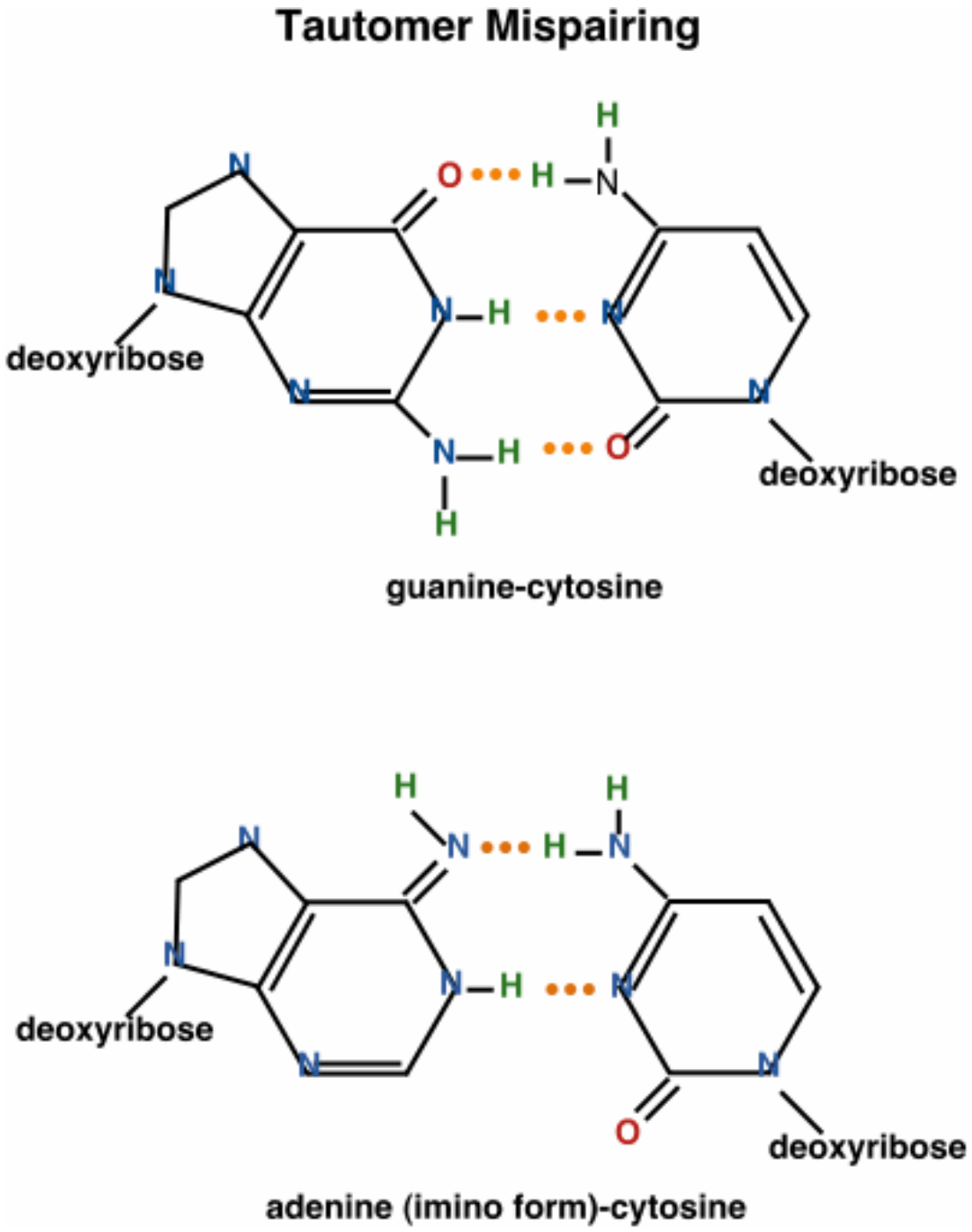
Fehler bei der Replikation

....und ihre Reparatur

Tautomerie der Nukleobasen

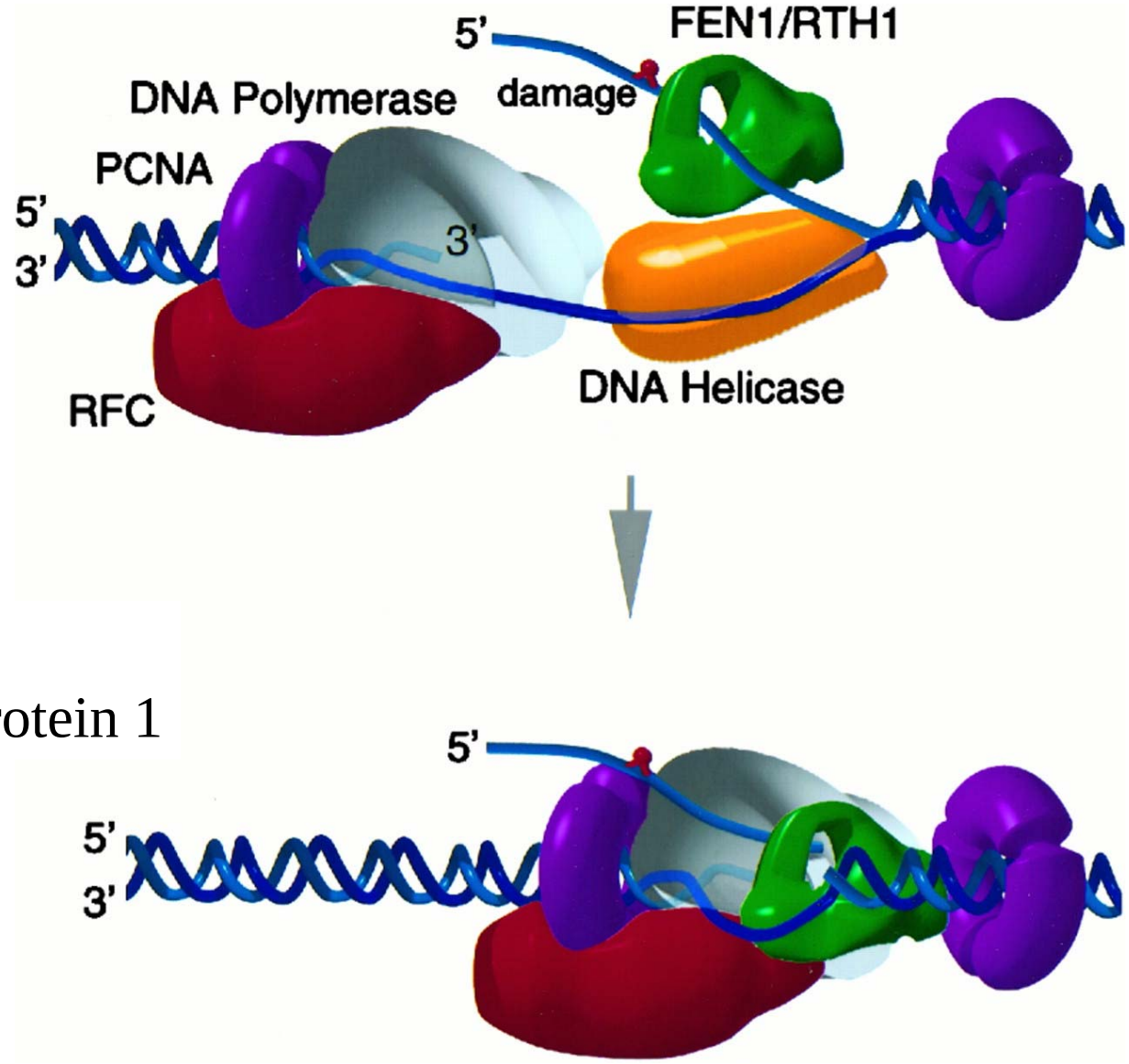


Replikationsfehler durch tautomere Formen der Nukleobasen



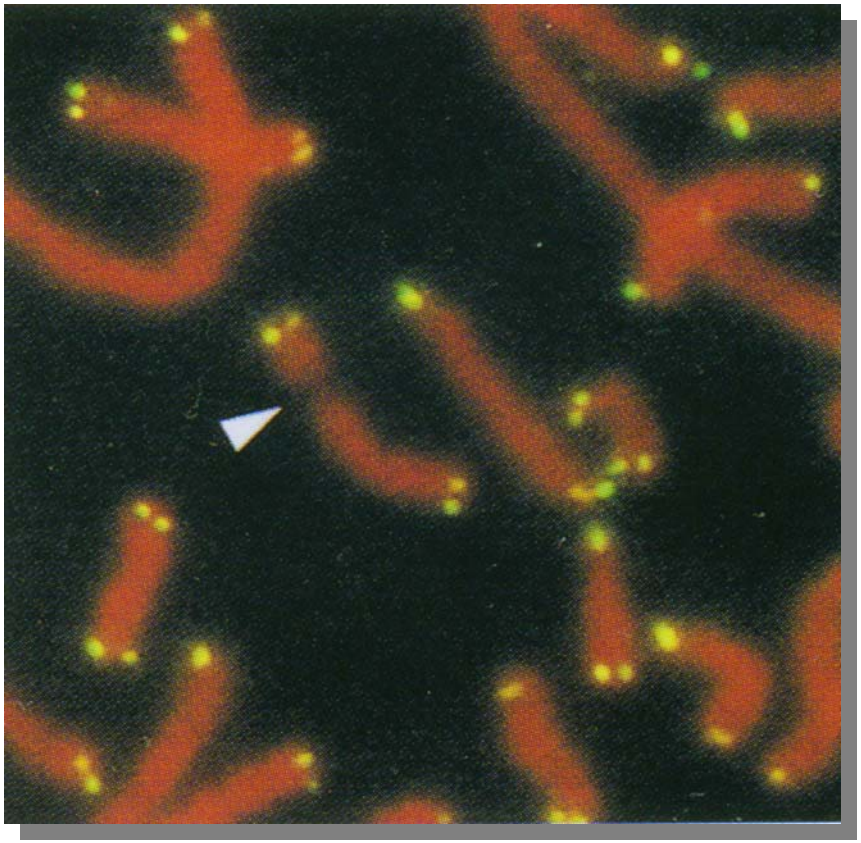
Reparatur von Replikations-fehlern

FEN1=
flap structure specific protein 1



Replikation der Telomere

Speziell gebaute Chromosomen-Enden (Telomere)
sowie eigens dafür vorgesehene Replikationsenzyme
(Telomerase) sorgen dafür, dass die Verluste kompensiert werden

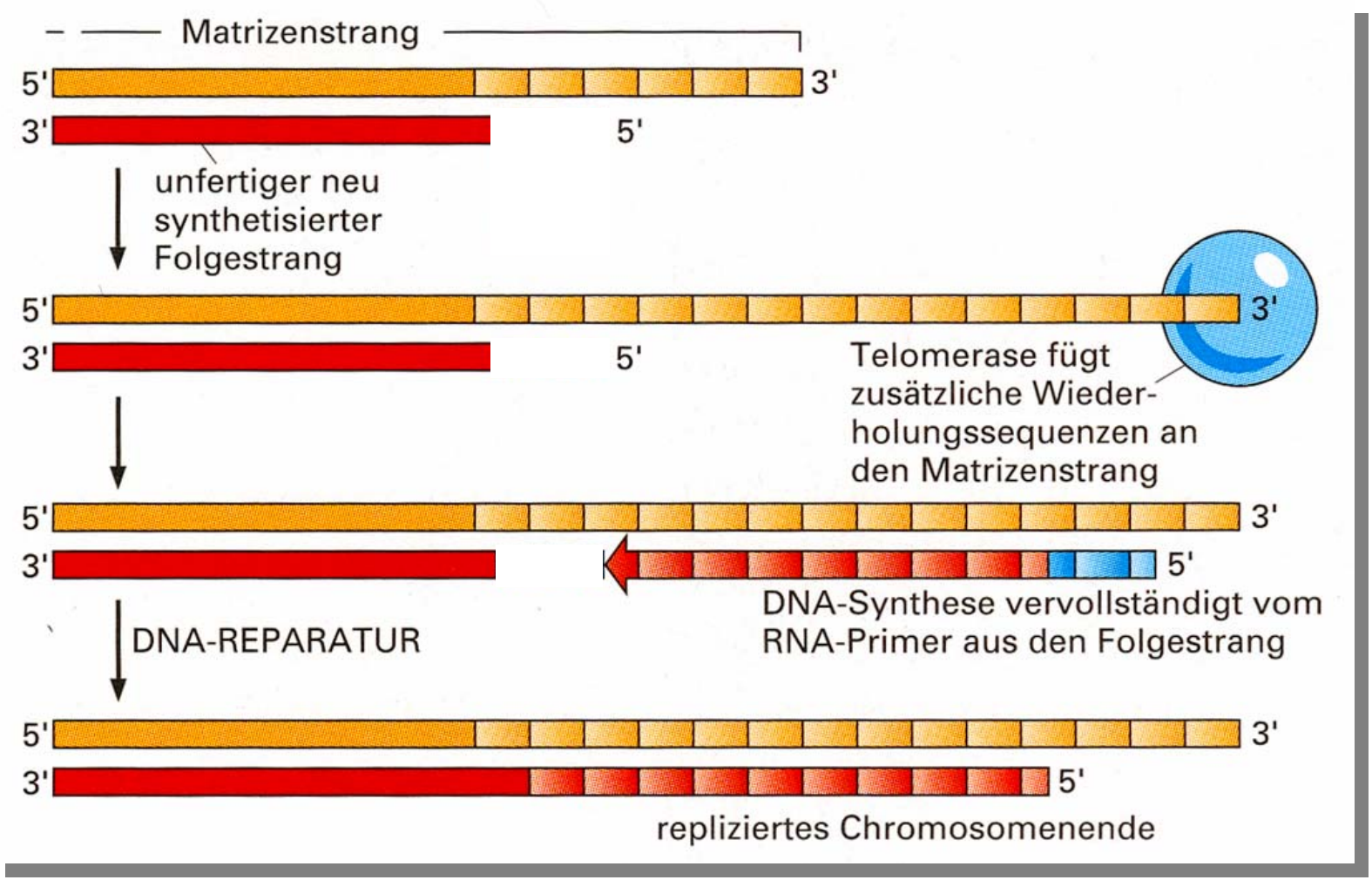


• die Telomer-DNA der meisten Tiere und Pflanzen enthält kurze, tandem-repetitive Sequenzen

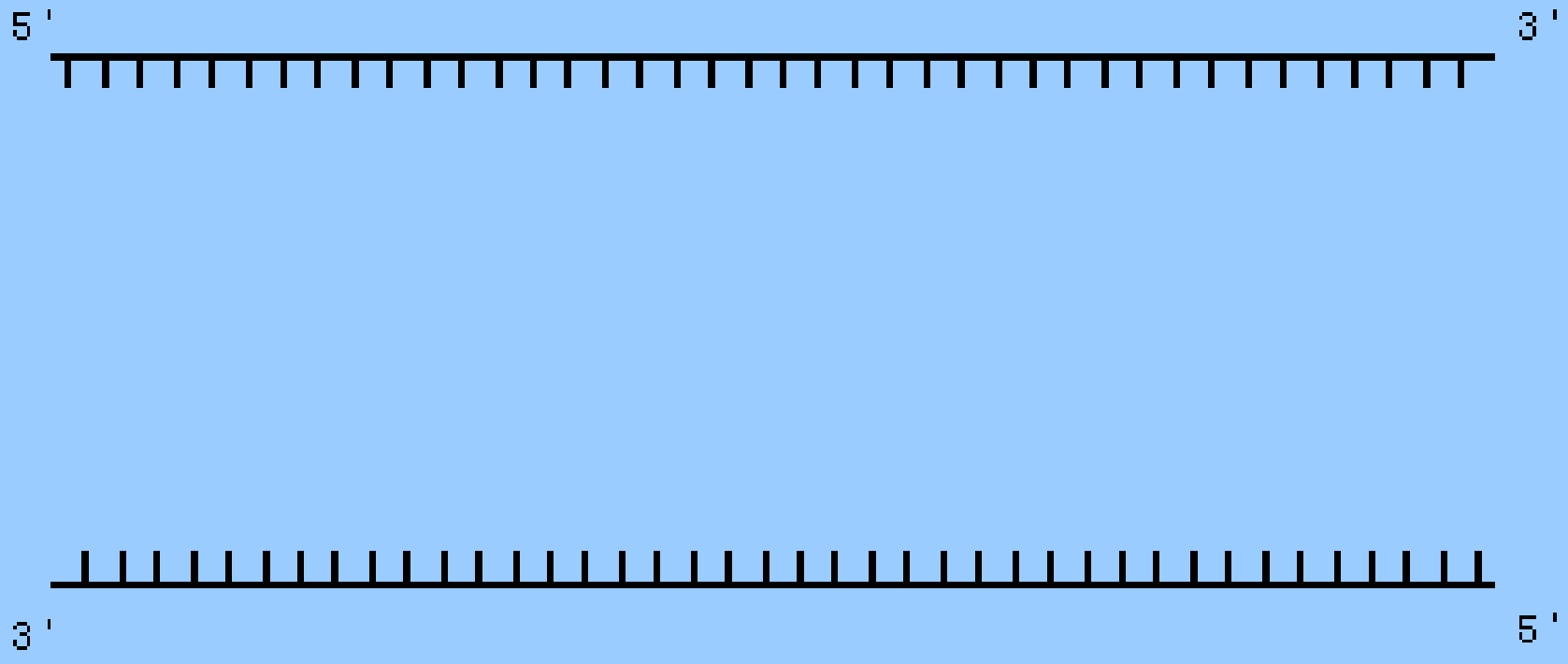
z. B. $(TTAGGG)_n$ beim Menschen

(Ausnahme: Dipteren wie Drosophila,
sowie wenige Pflanzenarten)

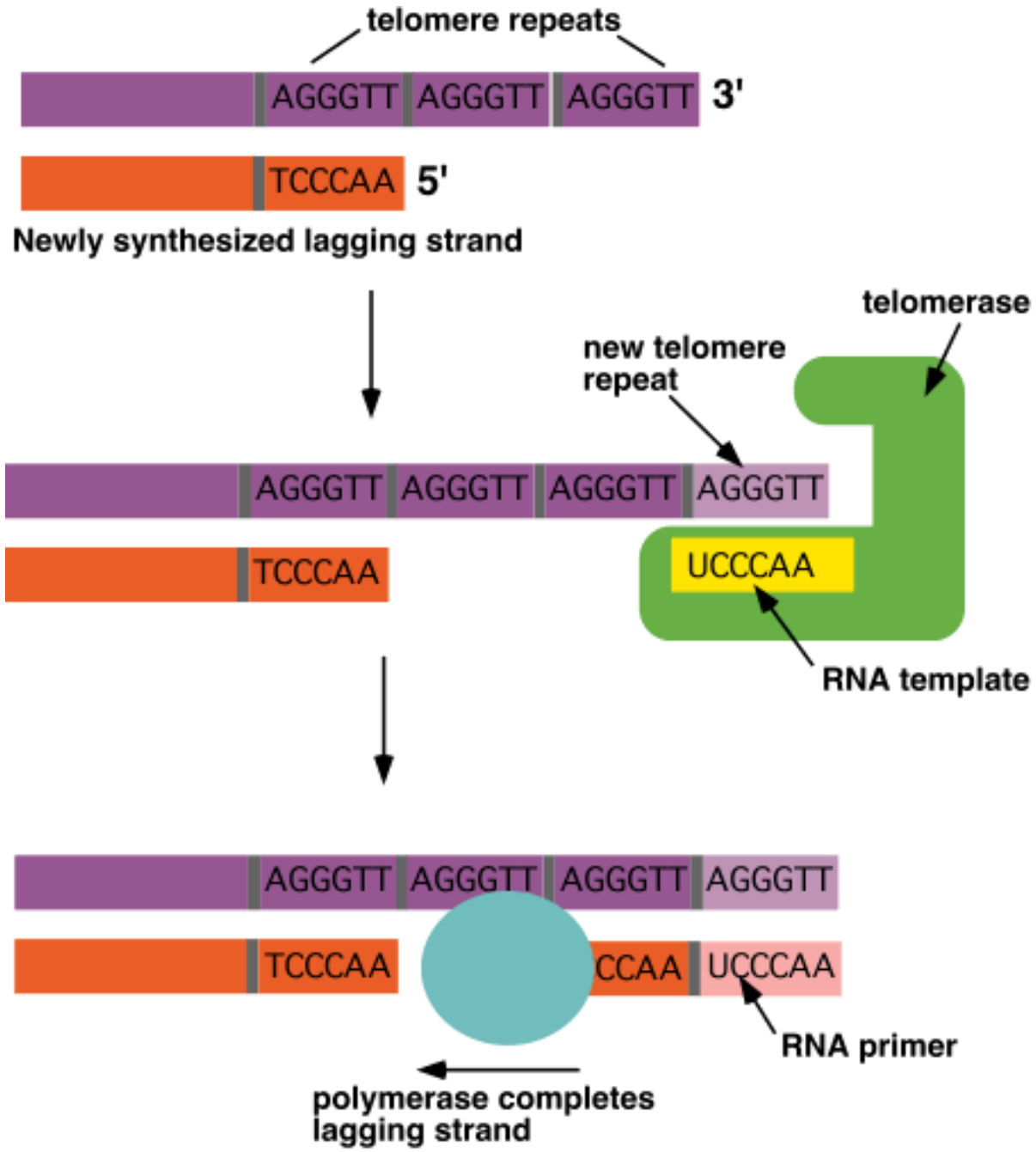
Die Telomerase verlängert den überhängenden 3‘-Strang



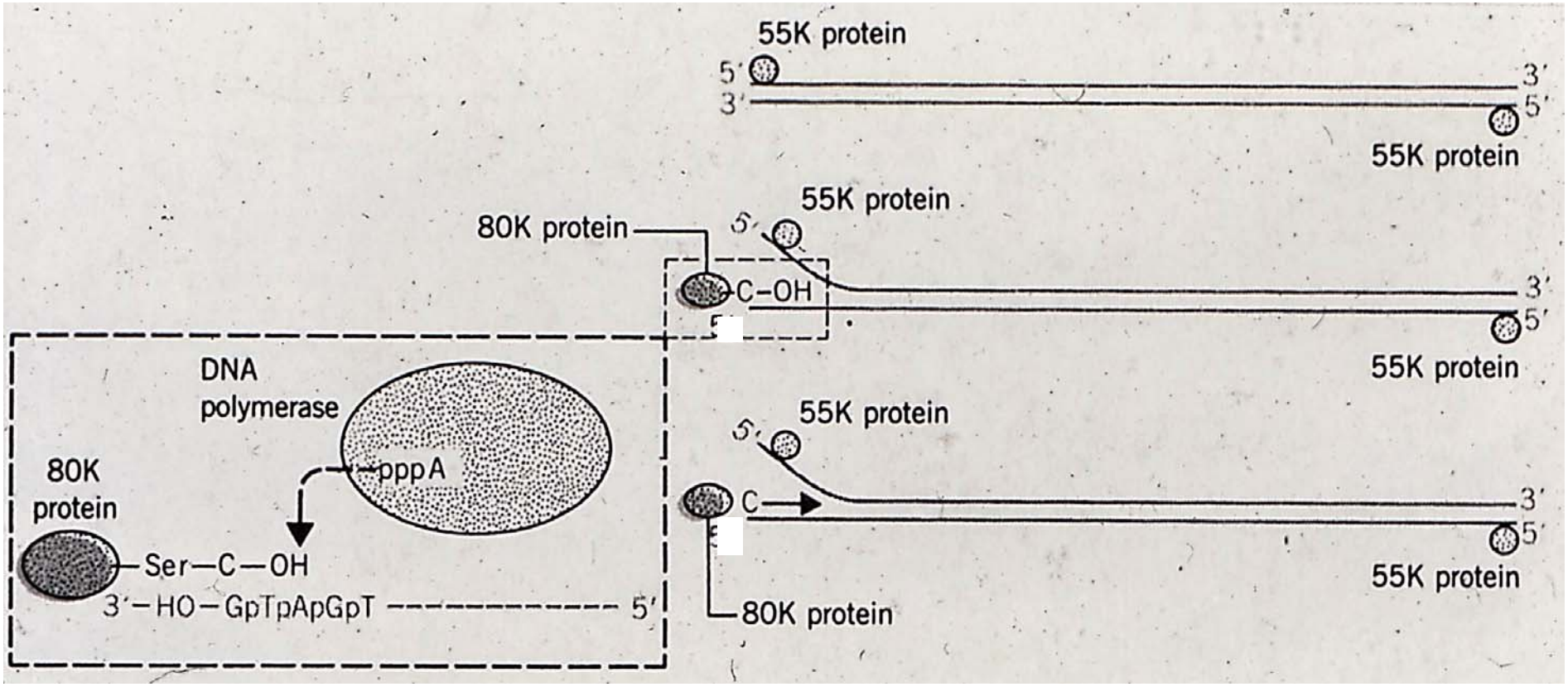
Replication of the lagging strand of a linear chromosome encounters a problem at the 3' end

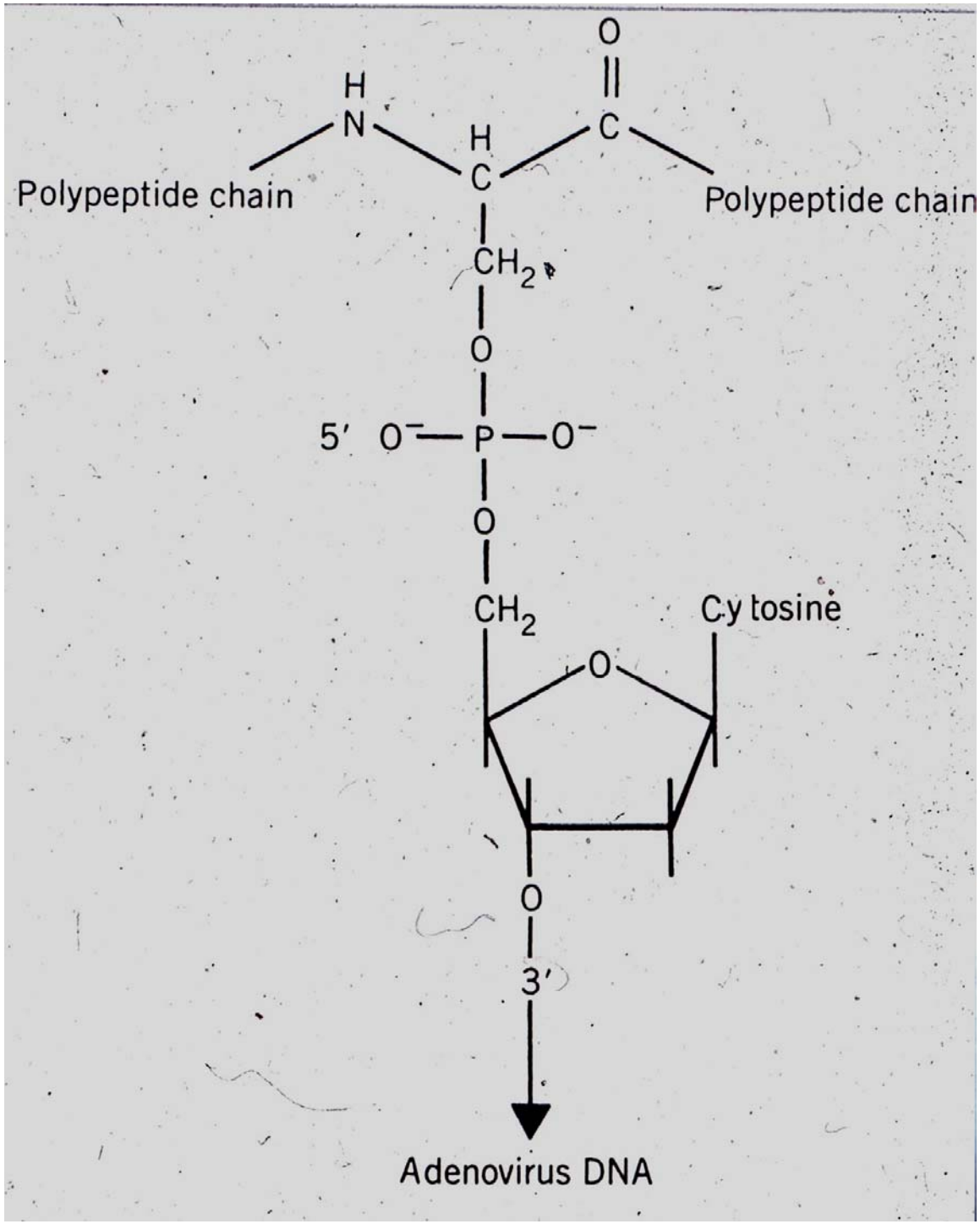


Telomer-replikation



Replikation linearer Virus-Genome
Beispiel Adenovirus

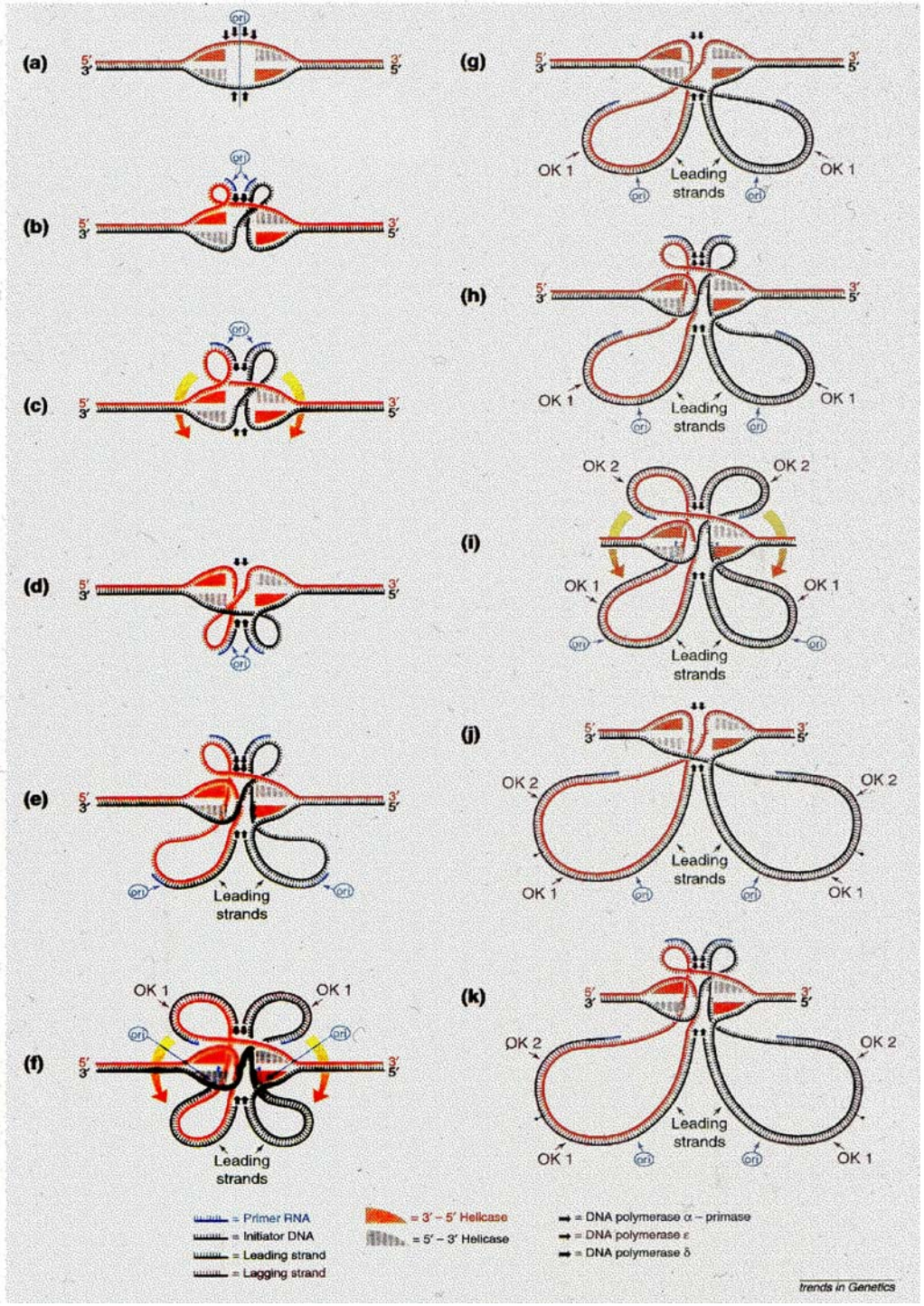


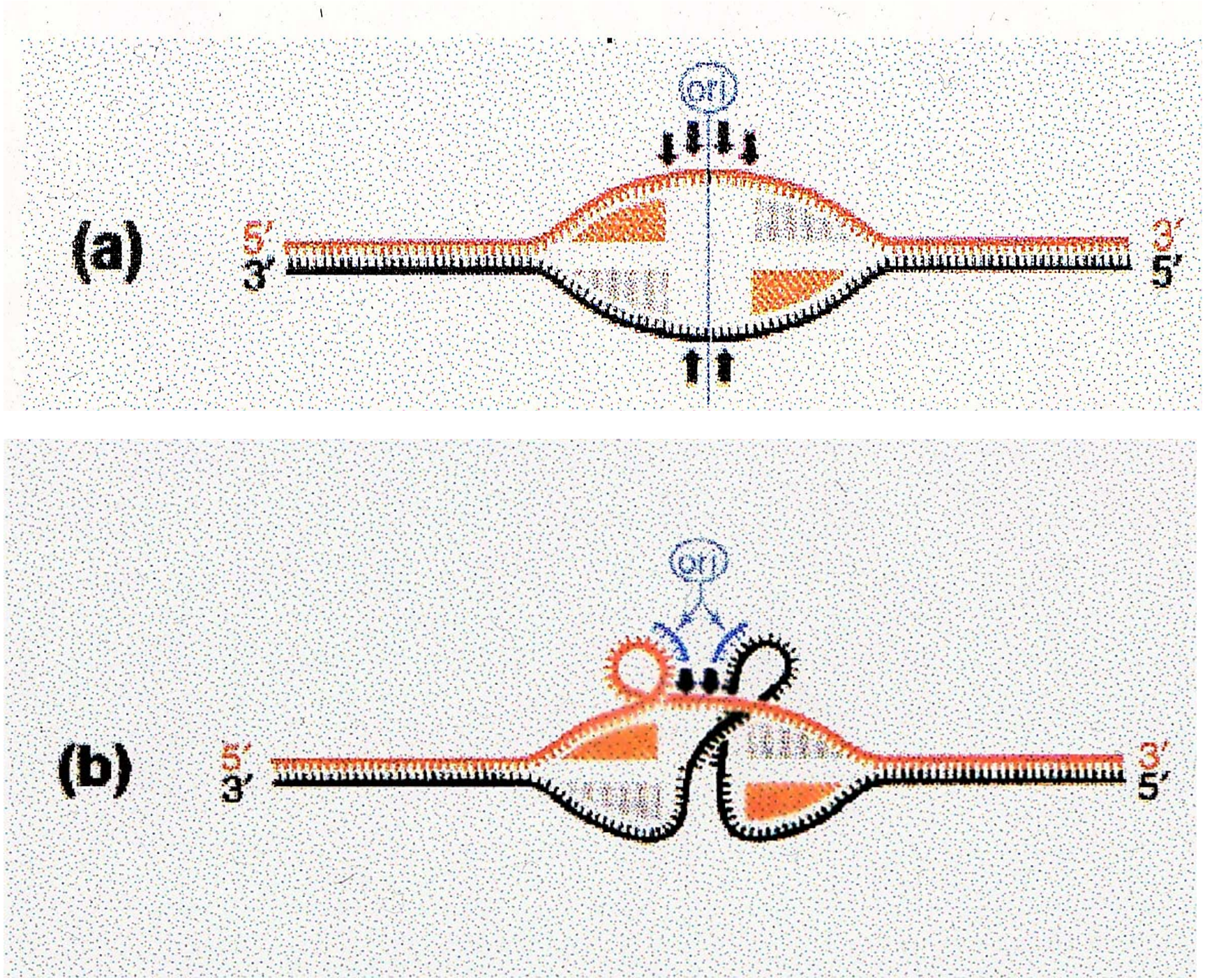

O
H
N
H
C
C
Polypeptide chain
Polypeptide chain
CH2
O
5′ O⁻ P O⁻
O
CH2
Cytosine
O
O
3′
Adenovirus DNA

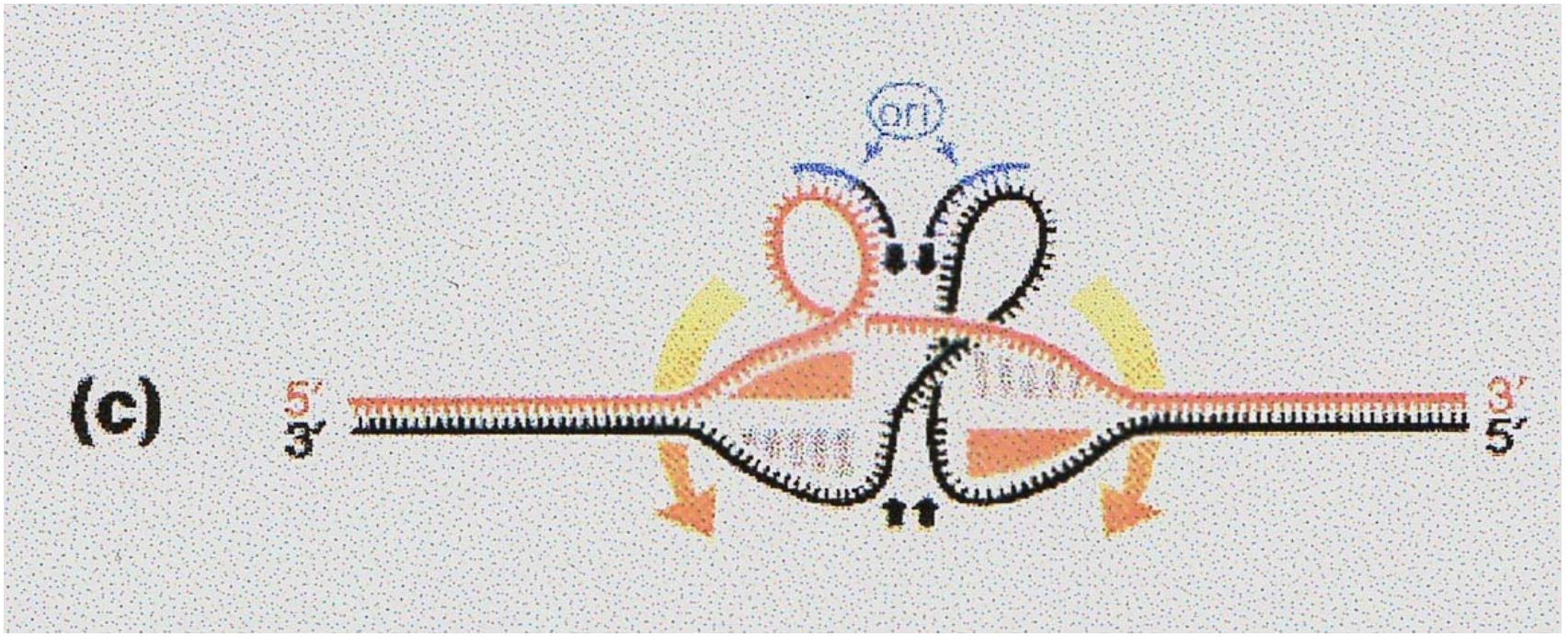
„Stationäre“ Replikation

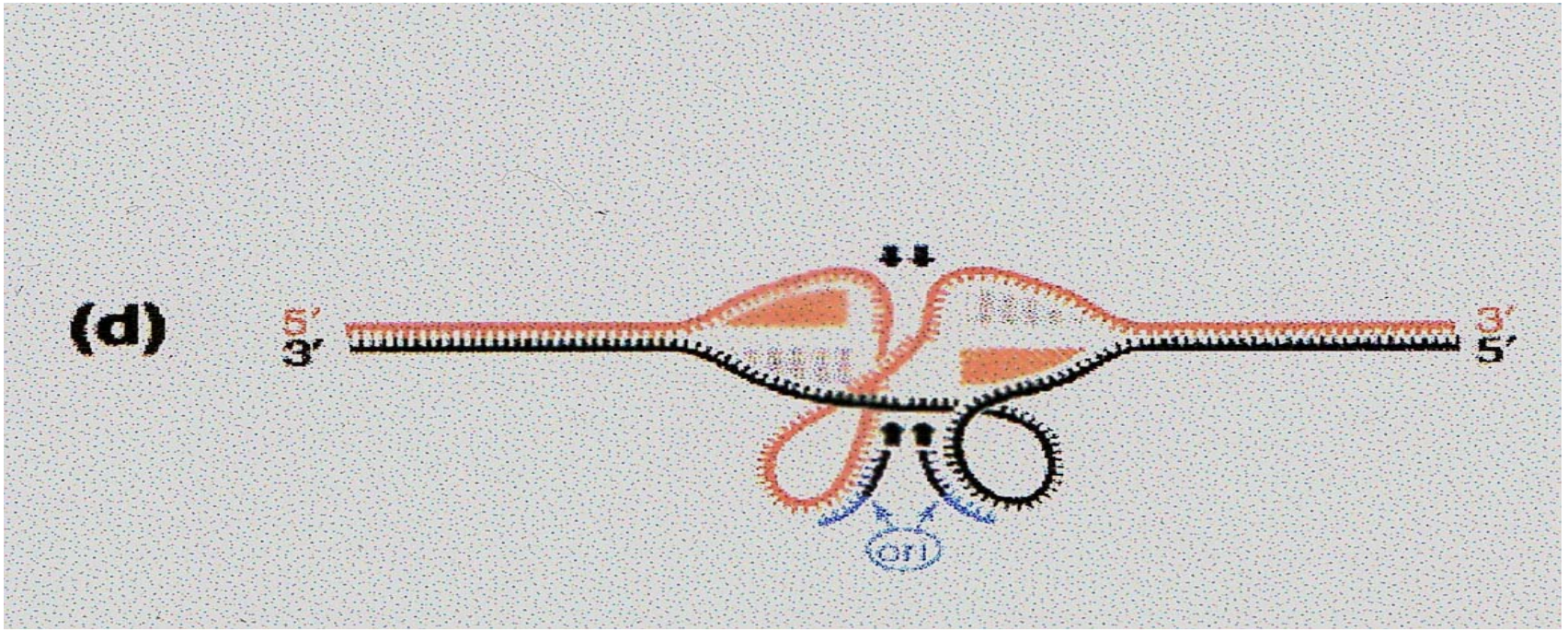
- Es gibt Hinweise darauf, dass nicht das Replisom, sondern die DNA sich bei der Replikation bewegt:

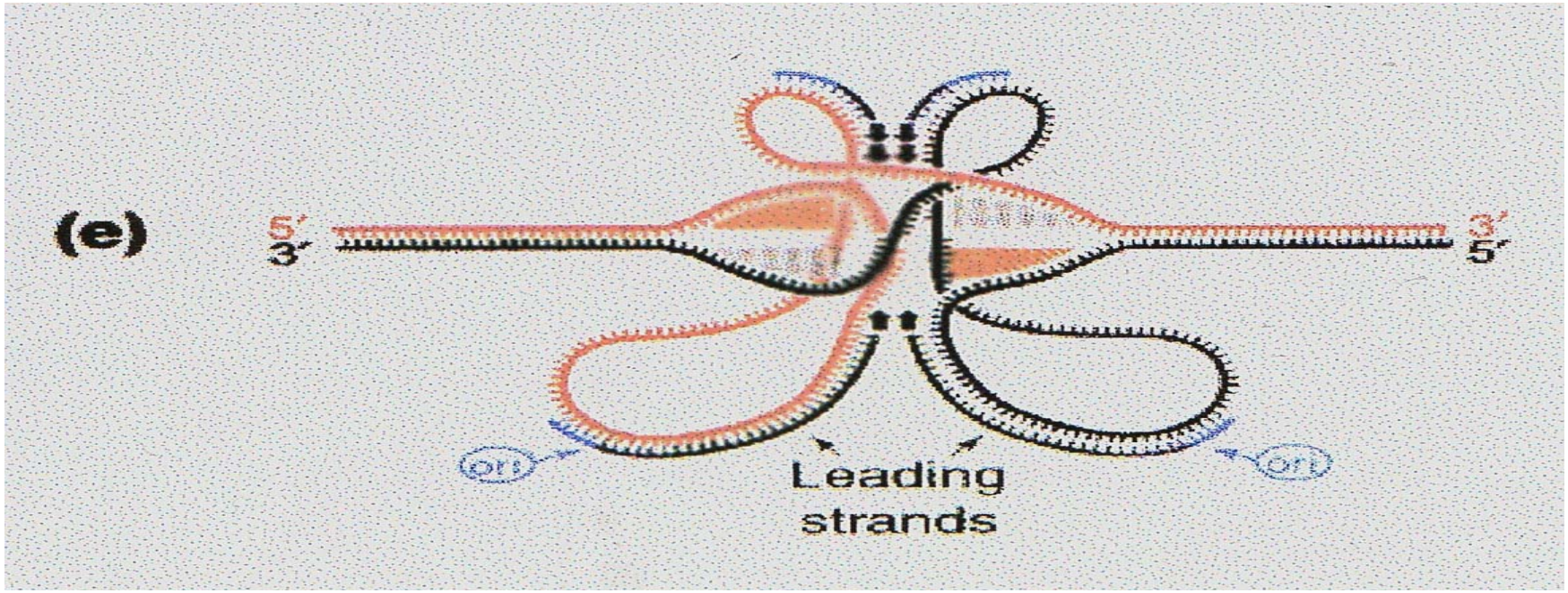
  „die Fabrik steht fest und das Fließband bewegt sich“

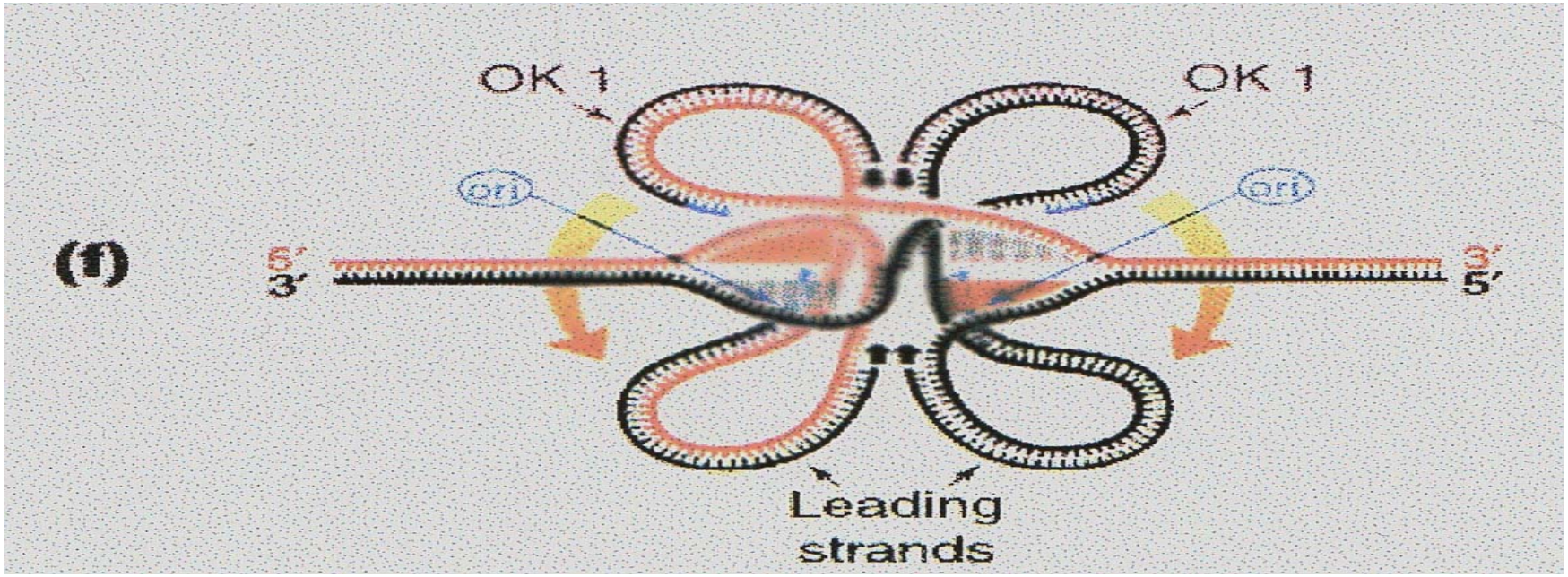

(a)
(b)
(c)
(d)
(e)
(f)
(g)
(h)
(i)
(j)
(k)
ori
5′
3′
OK 1
OK 2
Leading strands
= Primer RNA
= Initiator DNA
= Leading strand
= Lagging strand
= 3′ – 5′ Helicase
= 5′ – 3′ Helicase
= DNA polymerase α – primase
= DNA polymerase ε
= DNA polymerase δ
trends in Genetics

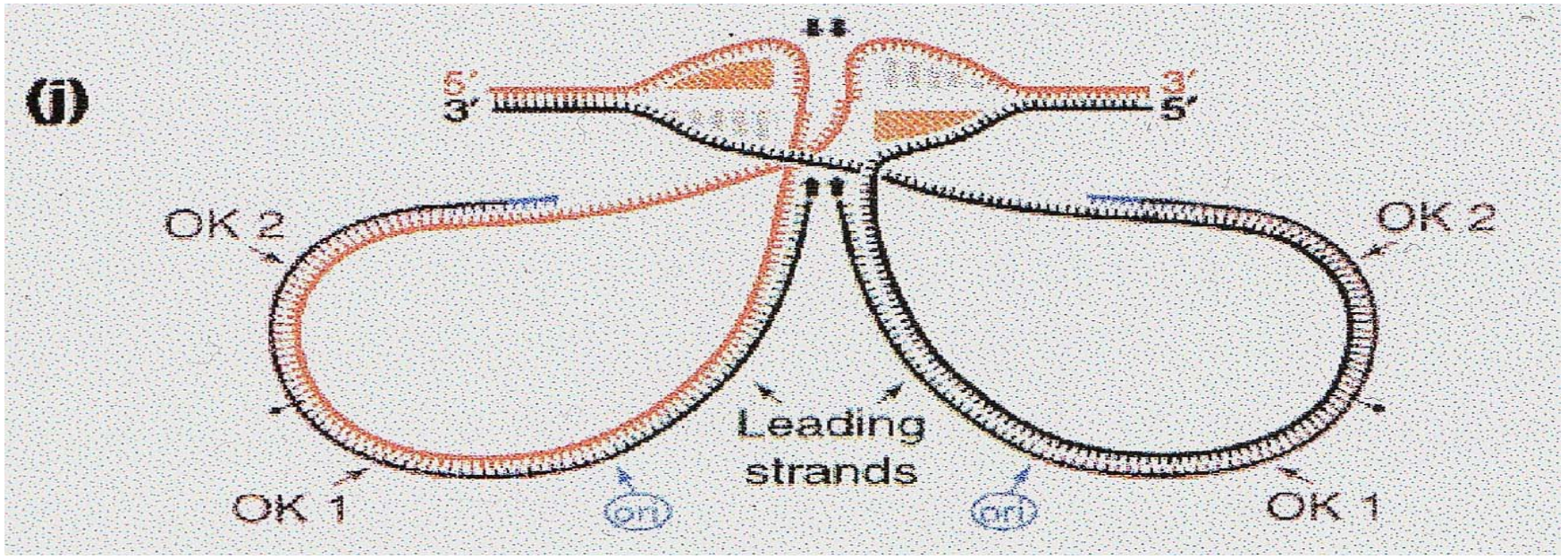

(a)
ori
5′
3′
3′
5′
(b)
ori
5′
3′
3′
5′

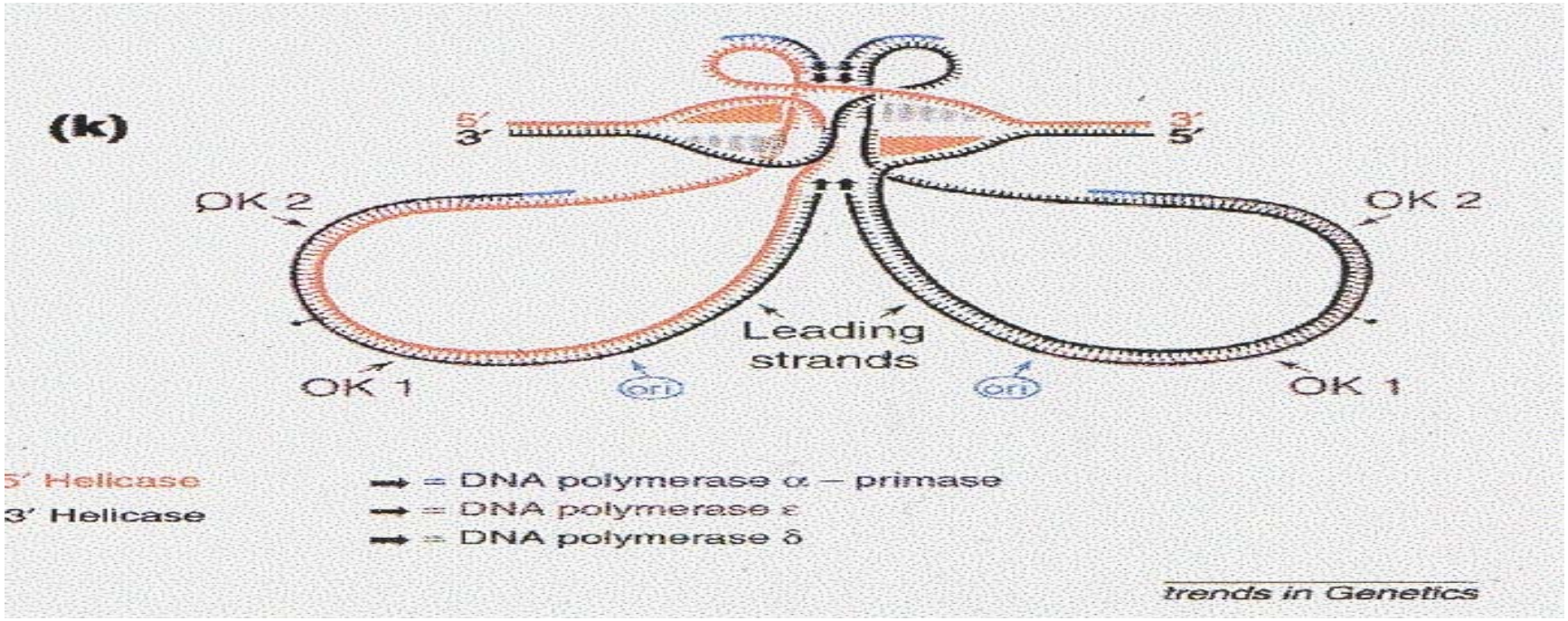

(c)
ori
5'
3'
3'
5'


(d)
5'
3'
3'
5'
ori


(e)
5′
3′
3′
5′
ori
ori
Leading
strands


OK 1
OK 1
(f)
ori
ori
5′
3′
3′
5′
Leading
strands


(j)
5′
3′
3′
5′
OK 2
OK 2
Leading
strands
OK 1
ori
ori
OK 1


(k)
5′
3′
3′
5′
OK 2
OK 2
Leading
strands
OK 1
ori
ori
OK 1
5′ Helicase
3′ Helicase
= DNA polymerase α – primase
= DNA polymerase ε
= DNA polymerase δ
trends in Genetics

Genstruktur der Eukaryoten

Abhängig von der Genklasse:

1. RNA Pol I – Gene: 18S, 5,8S, 28S rRNA
2. RNA Pol II – Gene: alle mRNAs
3. RNA Pol III – Gene: tRNAs, 5S rRNA, einige snRNAs

Eukaryotische RNA-Polymerasen

Eigenschaften der kerncodierten und im Kern aktiven DNS-abhängigen RNS Polymerasen			
	I	II	III
Transkriptions-produkt	Vorstufen der rRNS	Vorstufen der mRNS	5S rRNS, und Vorstufen der tRNS
Lokalisierung im Kern	Nukleolus	Nukleoplasma	Nukleoplasma
nach R. WOLLGIEN, 1982			

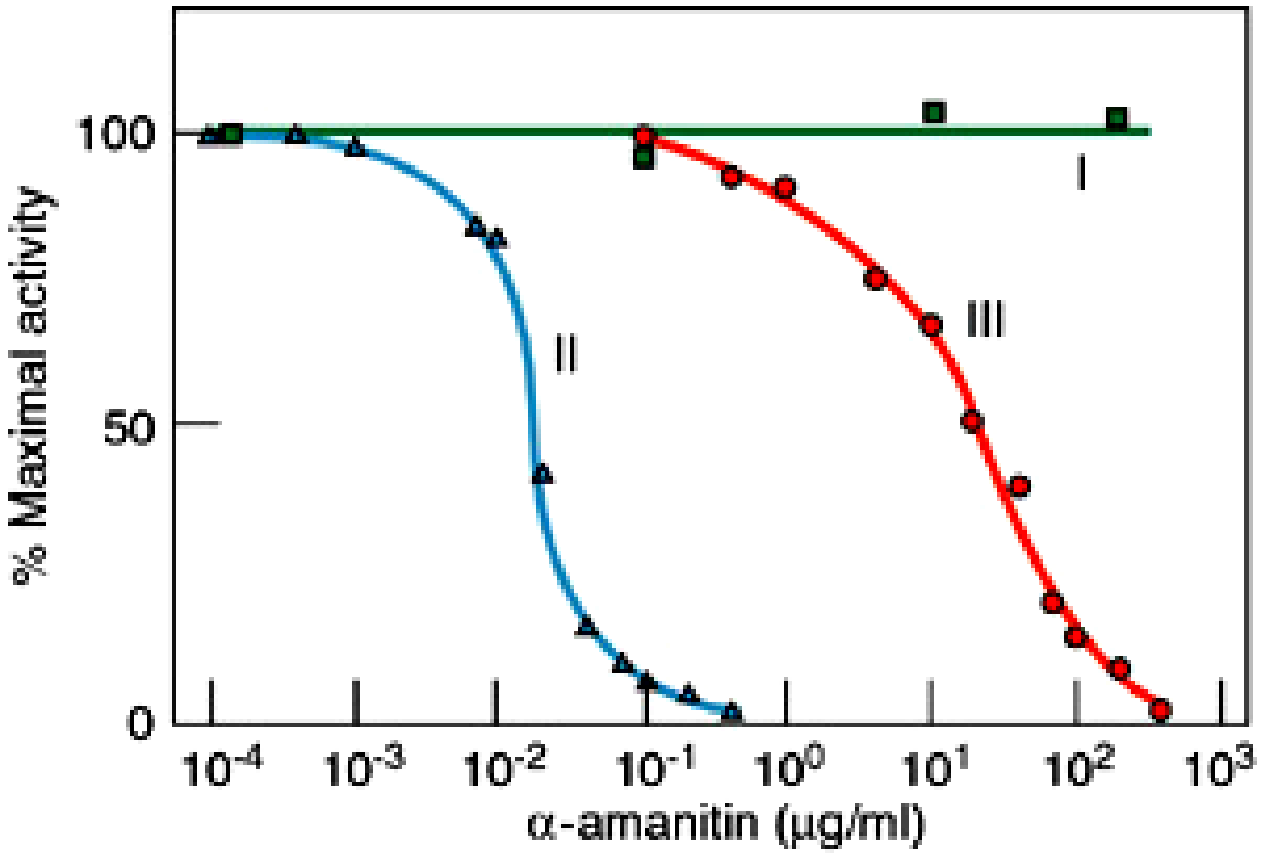
Die verschiedenen RNA-Polymerasen unterscheiden sich:

- Hemmbarkeit durch Actinomycin

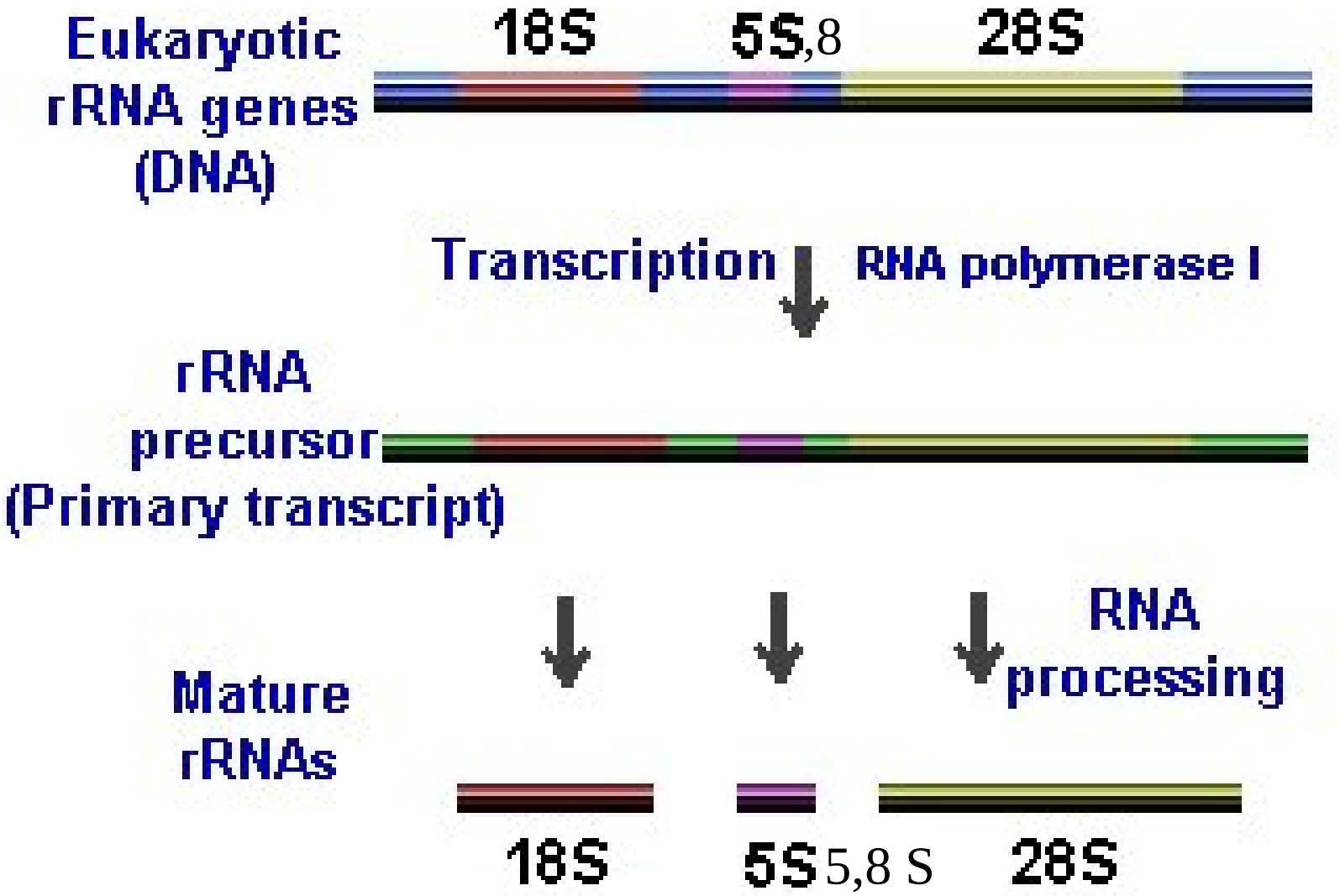
  Pol I sehr stark; Pol II/III schwächer
- Hemmbarkeit durch alpha-Amanitin

  Pol I nicht; Pol II sehr stark; Pol III schwach

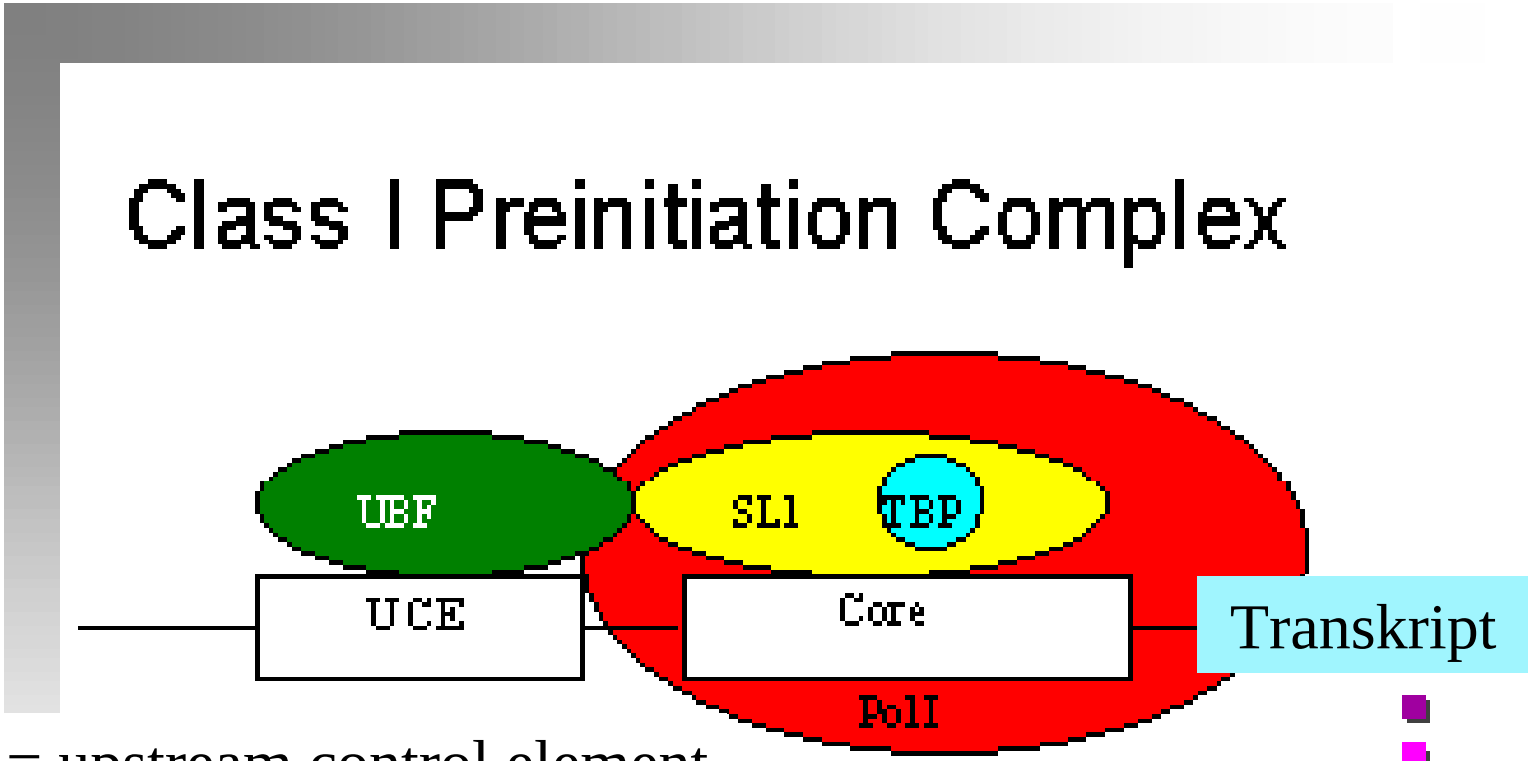
Hemmung der Polymerasen durch alpha-Amanitin



RNA-Polymerase I Gene:



RNA Polymerase I Promotor und Initiationskomplex



= upstream control element

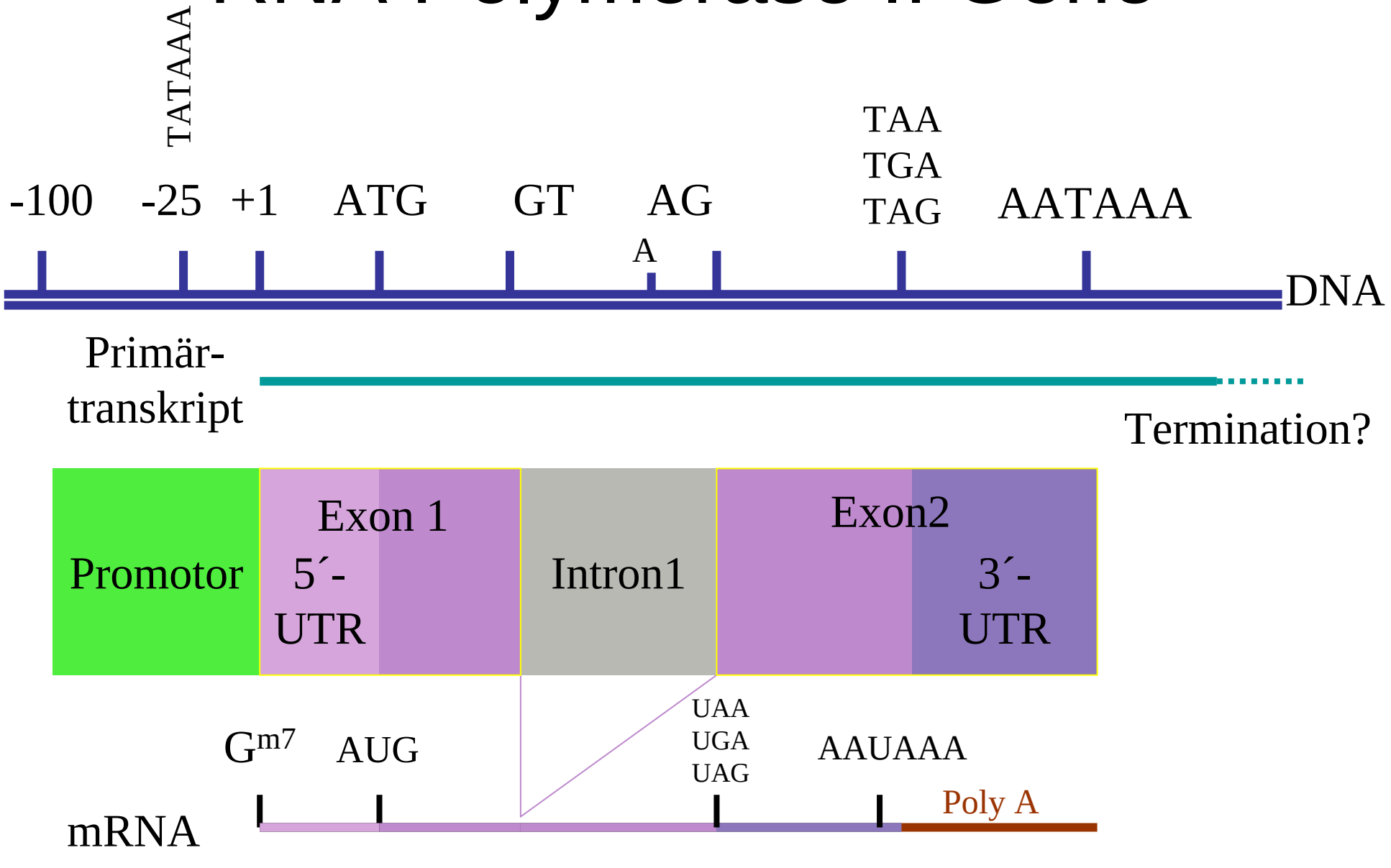
RNA Polymerase I Gene
haben einen 5´ aufwärts Promotor

RNA-Polymerase I Gene

- Promotor 5´stromaufwärts
- Primärtranskript enthält 18S, 5,8S und 28S rRNA sowie „spacer“ RNA
- Gene immer repetitiv vorhanden
- Gene in Gruppen („cluster“) tandemartig angeordnet (i. d. Regel „head to tail“)
- nur bei manchen Organismen: „Amplifikation“

  (= zusätzliche, extrachromosomale Genkopien in speziellen Geweben)

RNA Polymerase II Gene



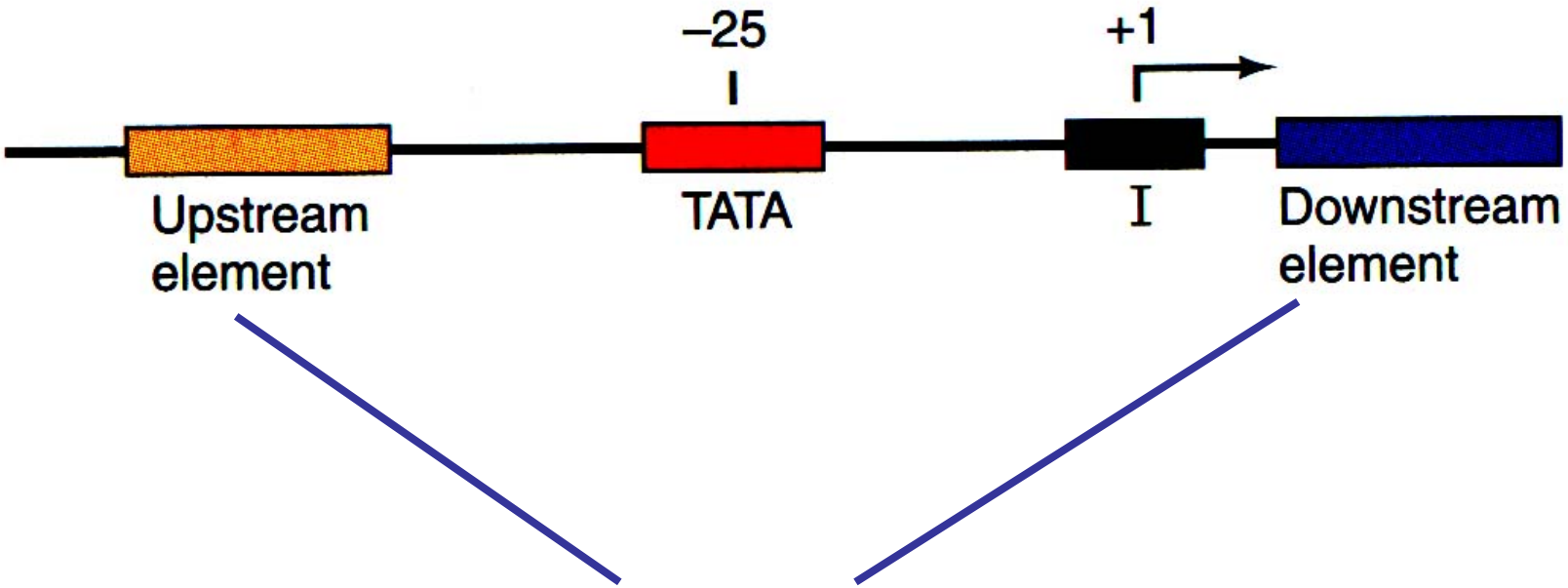
RNA – Polymerase II Gene

- 5-´stromaufwärts Promotor (oft TATA-Box bei -25)
- Intron – Exon – Struktur (i. d. Regel)
- Primärtranskript („hnRNA“) enthält 5´UTR, alle Exons und Introns, 3´- UTR
- Transkriptionsstart +1, = „Cap-Site“
- An Intron/Exongrenzen „consensus splice sites“ (Exon/ GT..Intron..AG/Exon)
- Polyadenylierungssignal
- Termination oft ungenau definiert
- Regulation durch „Enhancer“

RNA Pol II Promotorelemente Sequenz:Position:Funktion

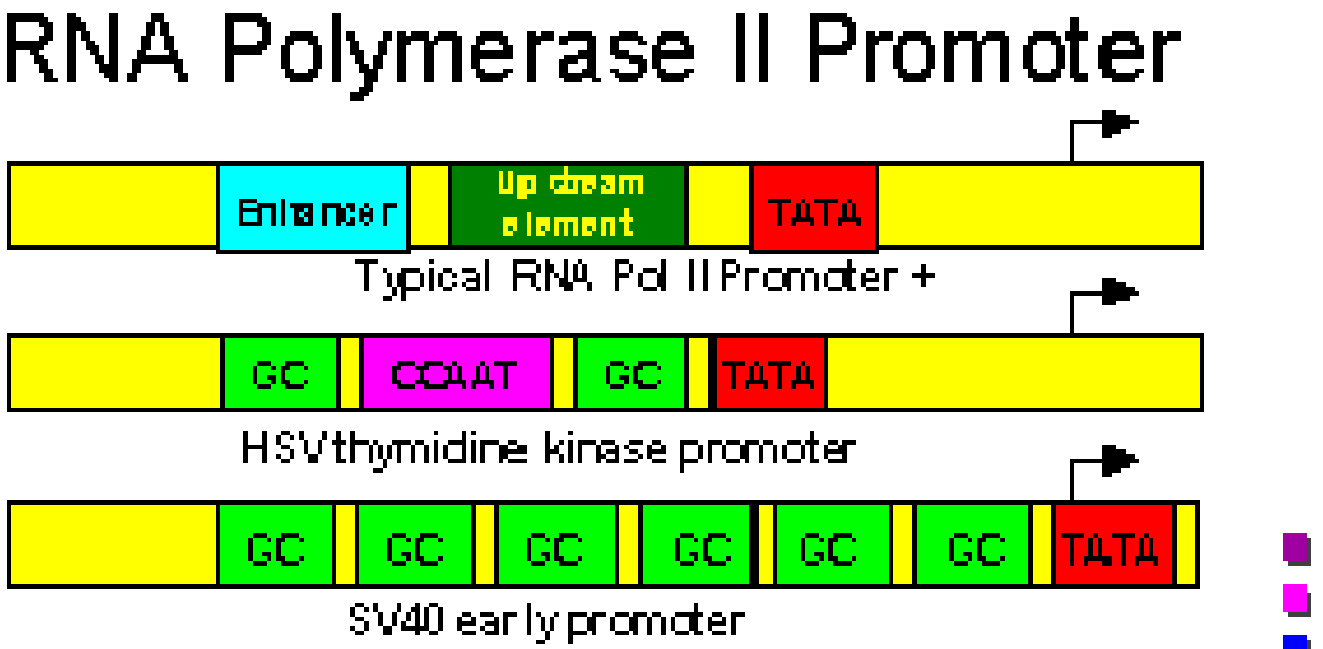
Name	Sequenz	Position	Funktion
TATA-Box Hogness-Box	TATAAA	-25 bis -30	Definiert Transkript-startpunkt
CAT-Box	GGCCAATC	-60 bis -80	Polymerase-Bindung via CBP
GC-Box	GGGCG	Variabel und mehrfach 1 bis 107	Definiert RNA-Pol Bindungstelle

Promotor Elemente RNA-Pol II

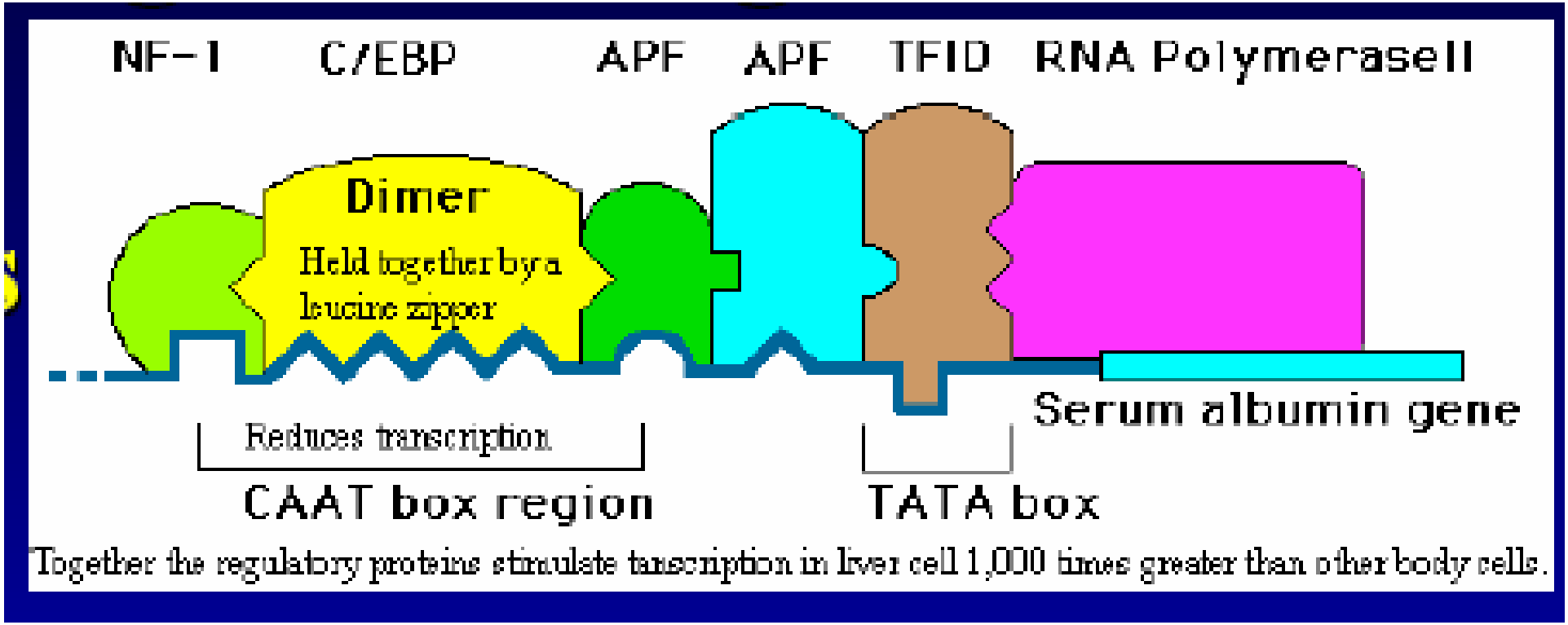


Enhancer oder Silencer

RNA Polymerase II Promotor



RNA Polymerase II Promotor
Beispiel Serumalbumin Gens



Enhancer

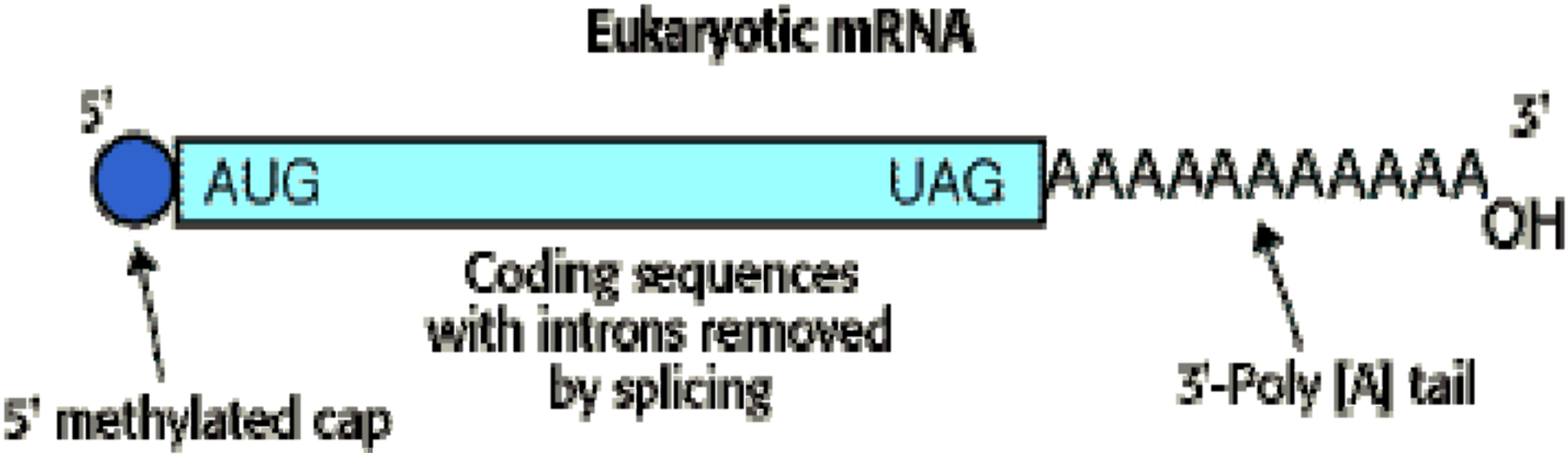
sind cis-regulatorische Kontrollelemente,
die unabhängig von ihrer Orientierung,
Lage oder Distanz zum Gen
die Aktivität eines Gens steigern können

Silencer

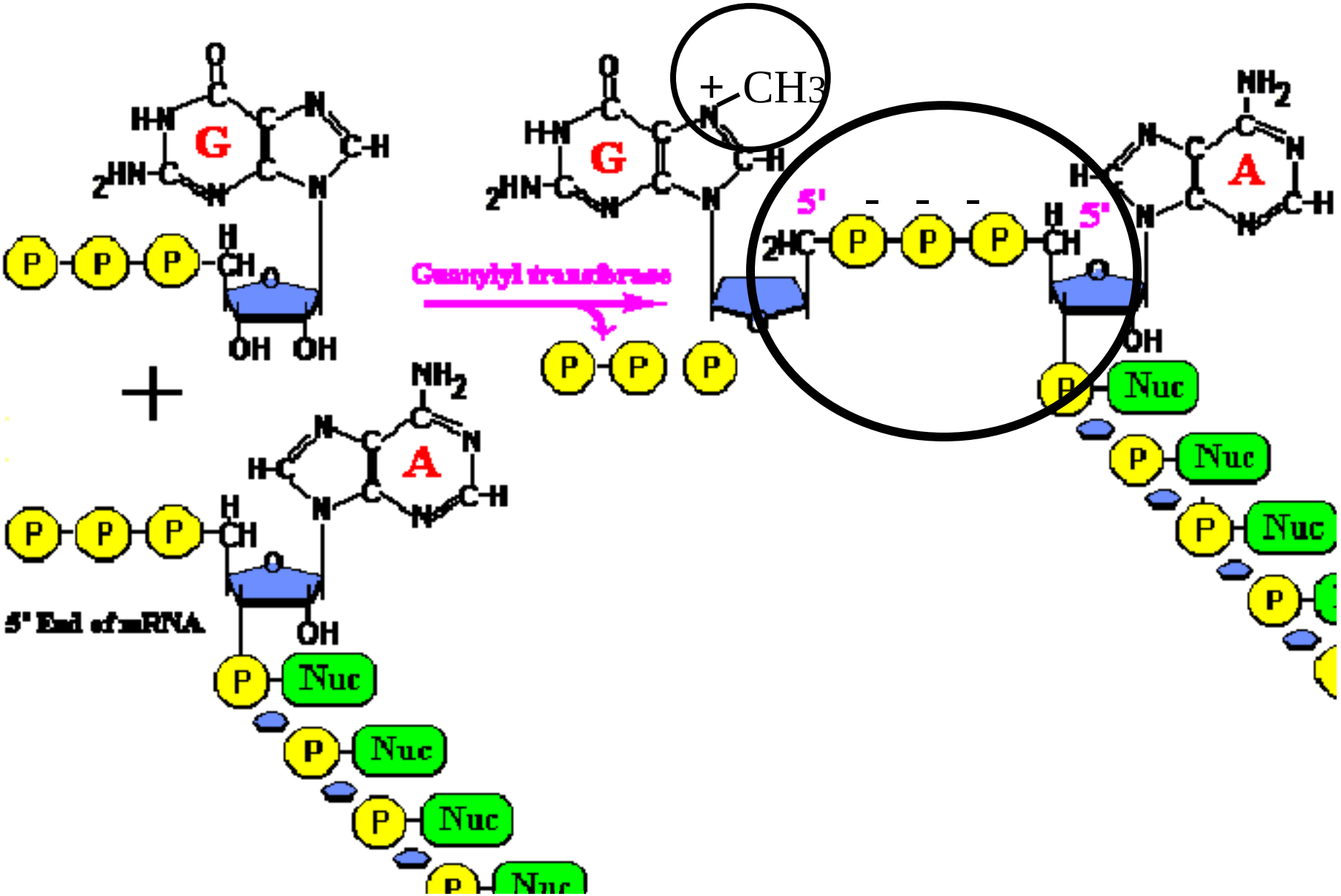
Wirkungsweise wie negativer Enhancer,
setzt Genaktivität herab

„posttranskriptionelle“ Modifikationen

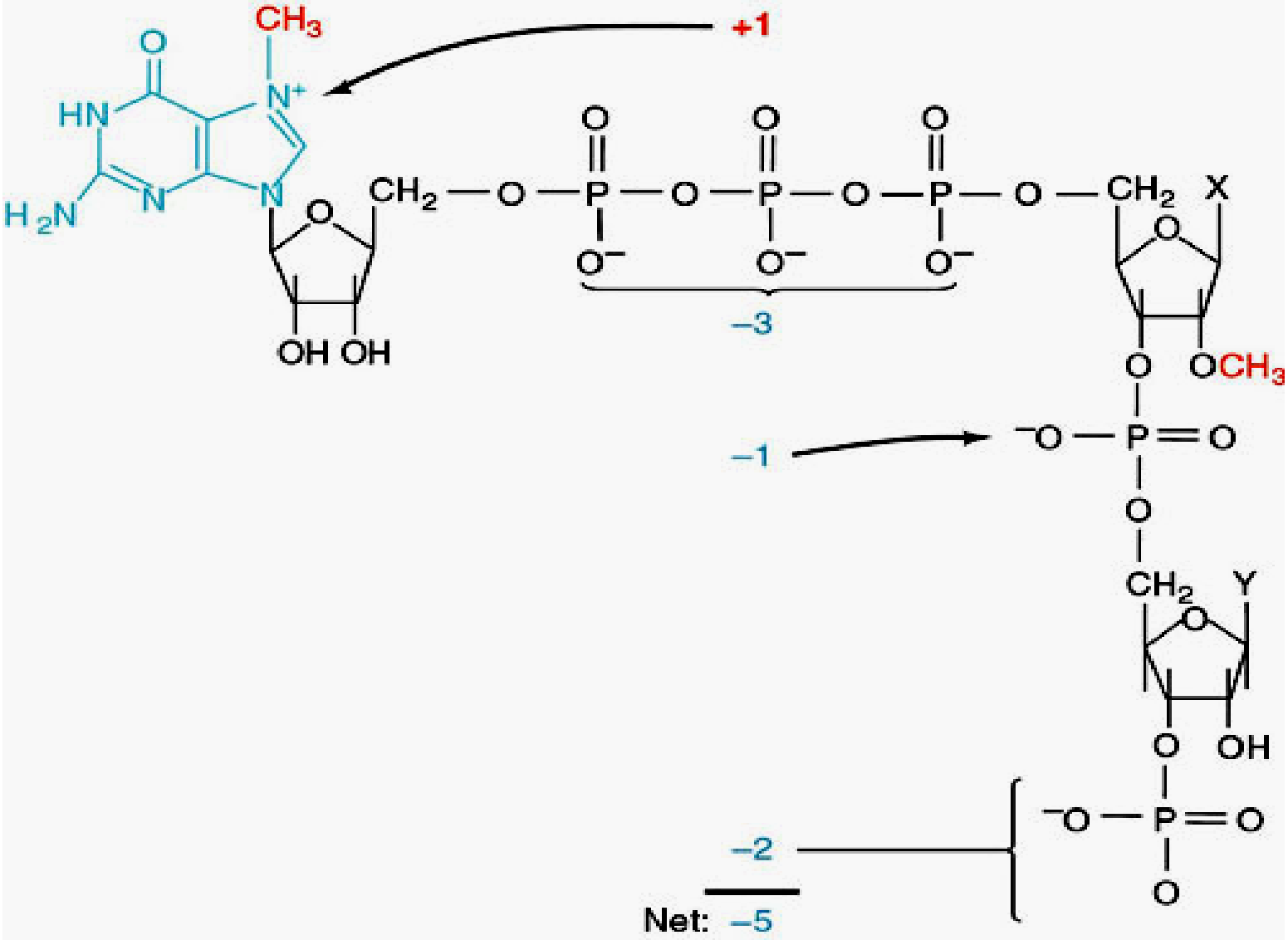
typisch für eukaryotische mRNA:



„Capping“ der mRNA



Cap-Struktur am 5´-Ende der mRNA



Funktion der Cap-Struktur

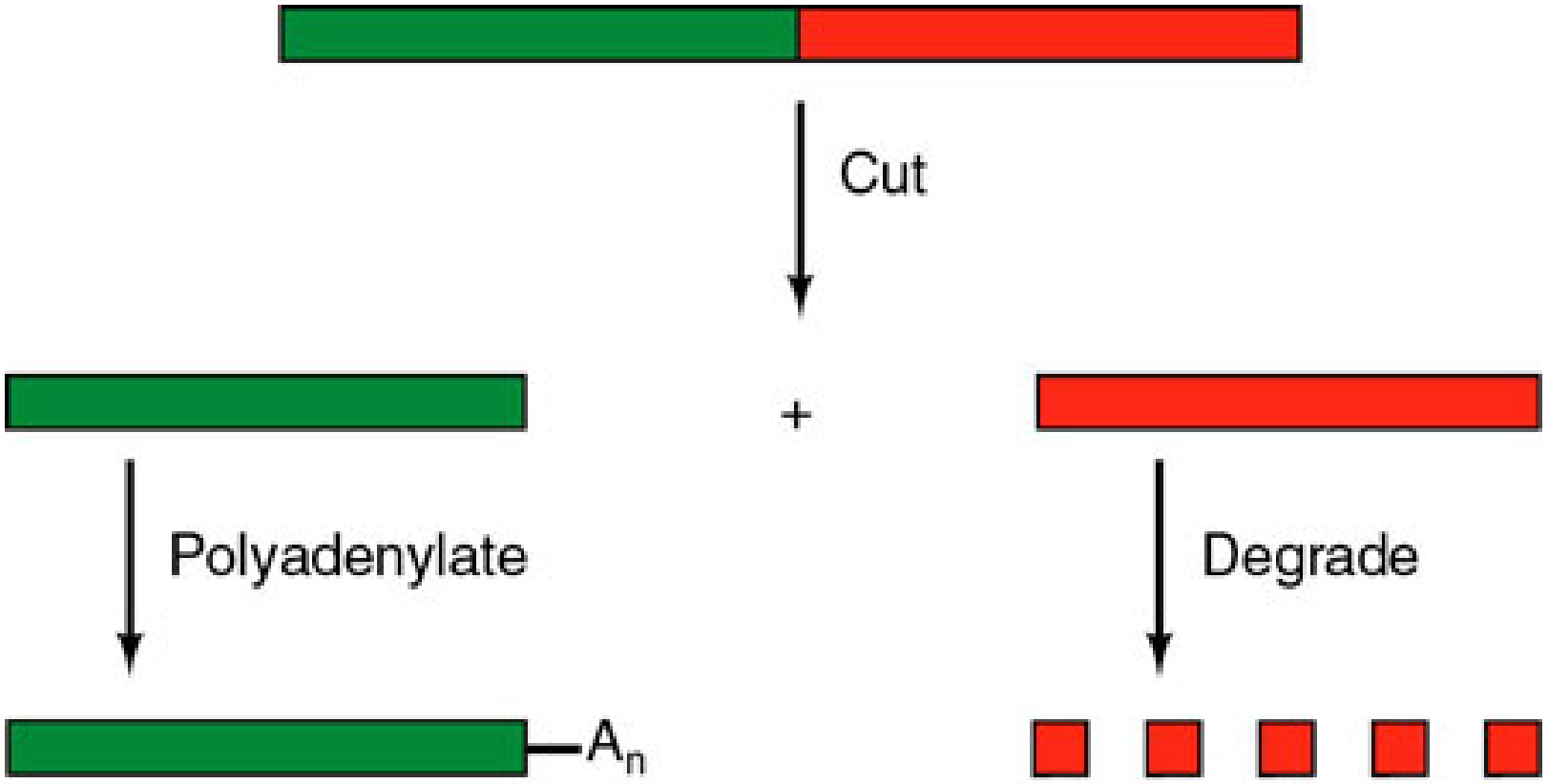
- erhöht Stabilität der mRNA
- induziert Splicing
- fördert Export aus dem Nukleus
- vermittelt Bindung der Ribosomen an mRNA und macht Translation möglich

Poly-Adenylierung am 3´Ende der mRNA

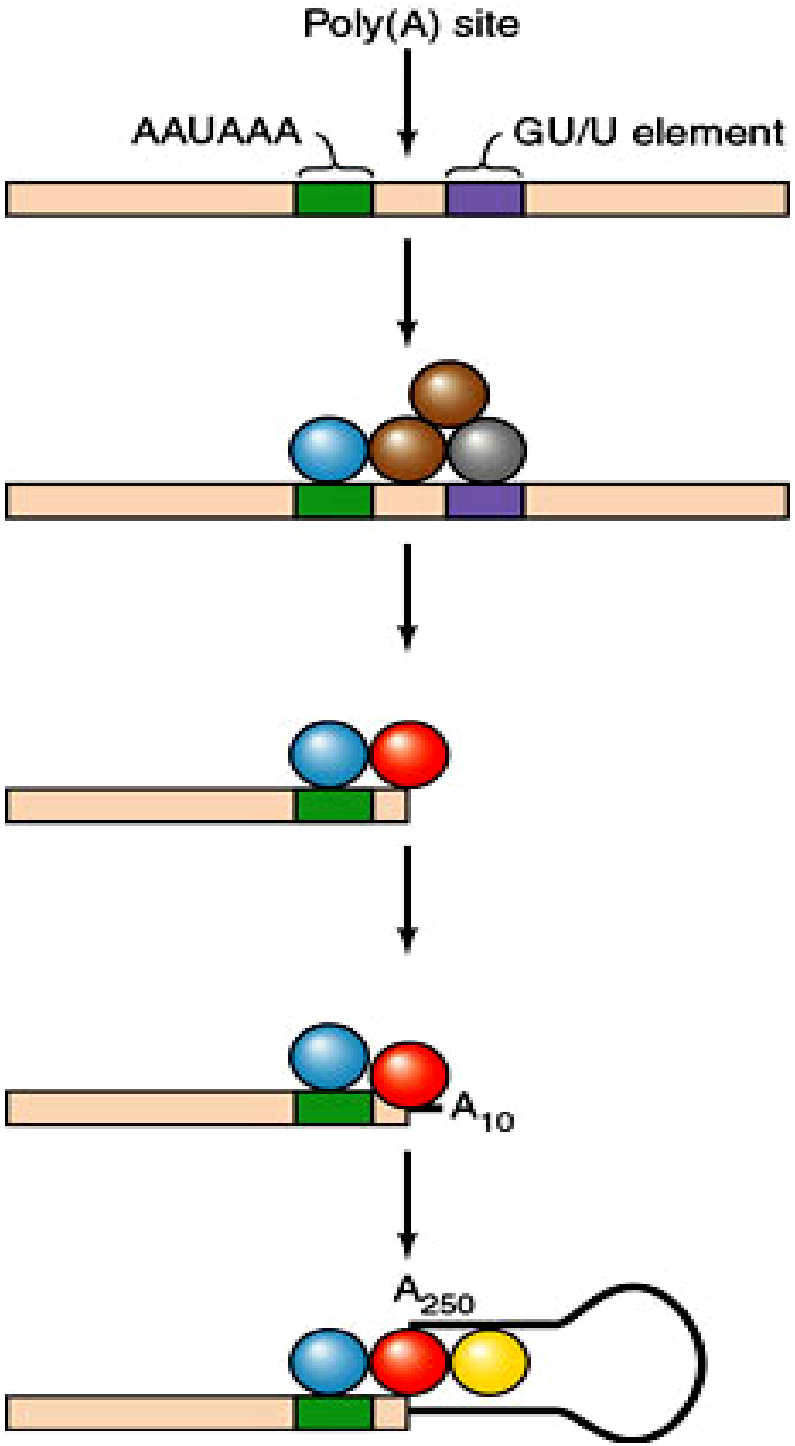
1. Trimmen des Primärtranskripts an definierter Stelle (23-24 Basen stromabwärts des Poly A-Signals AAUAA)

2. Anfügen von Adenin-Nukleotiden

„Polyadenylierung“: Trimmen der mRNA und Anhängen von mehreren Adeninnukleotiden



Die Polyadenylierung wird von einem Enzymkomplex aus mehreren Enzymen erledigt



Die Polyadenylierung ist ähnlich komplex wie die Initiation!

Aus: Strange bedfellows: polyadenylation factors at the promoter
Olga Calvo1 and James L. Manley, GENES & DEVELOPMENT 17:1321–1327 © 2003

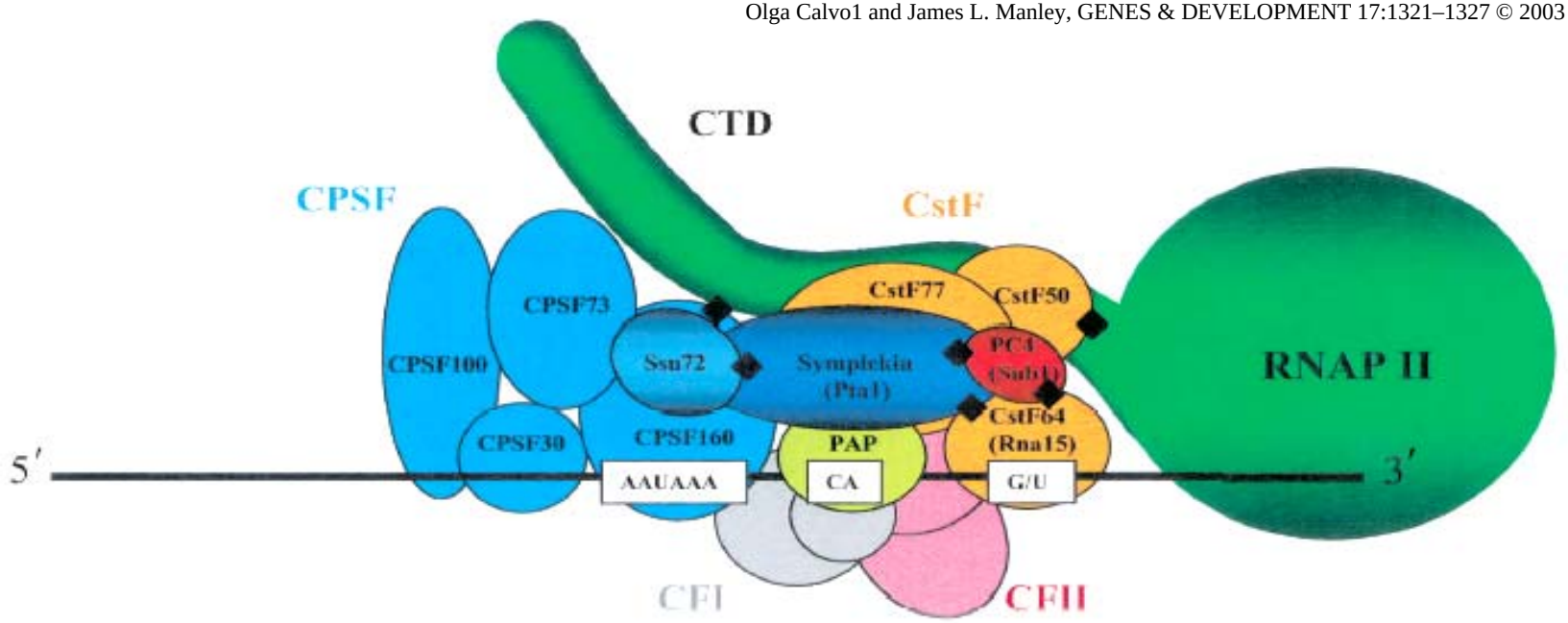


Figure 2. Schematic representation of the polyadenylation machinery. The majority of the components of the mammalian and yeast polyadenylation complexes are conserved, including all currently known factors that function in the transcription connection. For simplicity, only the mammalian nomenclature is depicted; the yeast names of factors that have important roles in the events described here are also indicated. (Note that although an apparent human homolog of Ssu72 exists, it has not yet been characterized functionally). ◆, documented protein–protein interactions that help link transcription and 3′ processing (see text). Polyadenylation signal sequences (upstream AAUAAA, CA cleavage site consensus, and downstream G/U-rich region) are boxed. CPSF, cleavage-polyadenylation specificity factor; CstF, cleavage stimulation factor; CFI and CFII, cleavage factors I and II, respectively; PAP, poly(A) polymerase.

Poly-Adenylierung am 3´Ende der mRNA

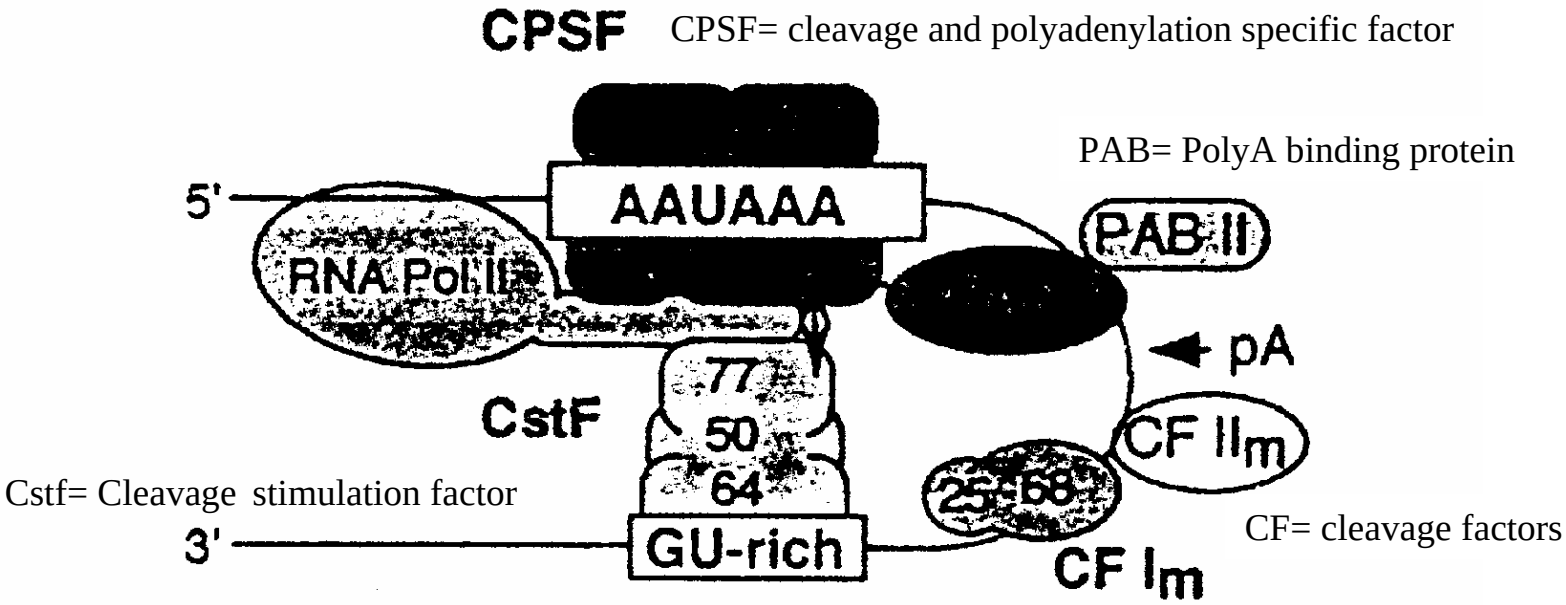


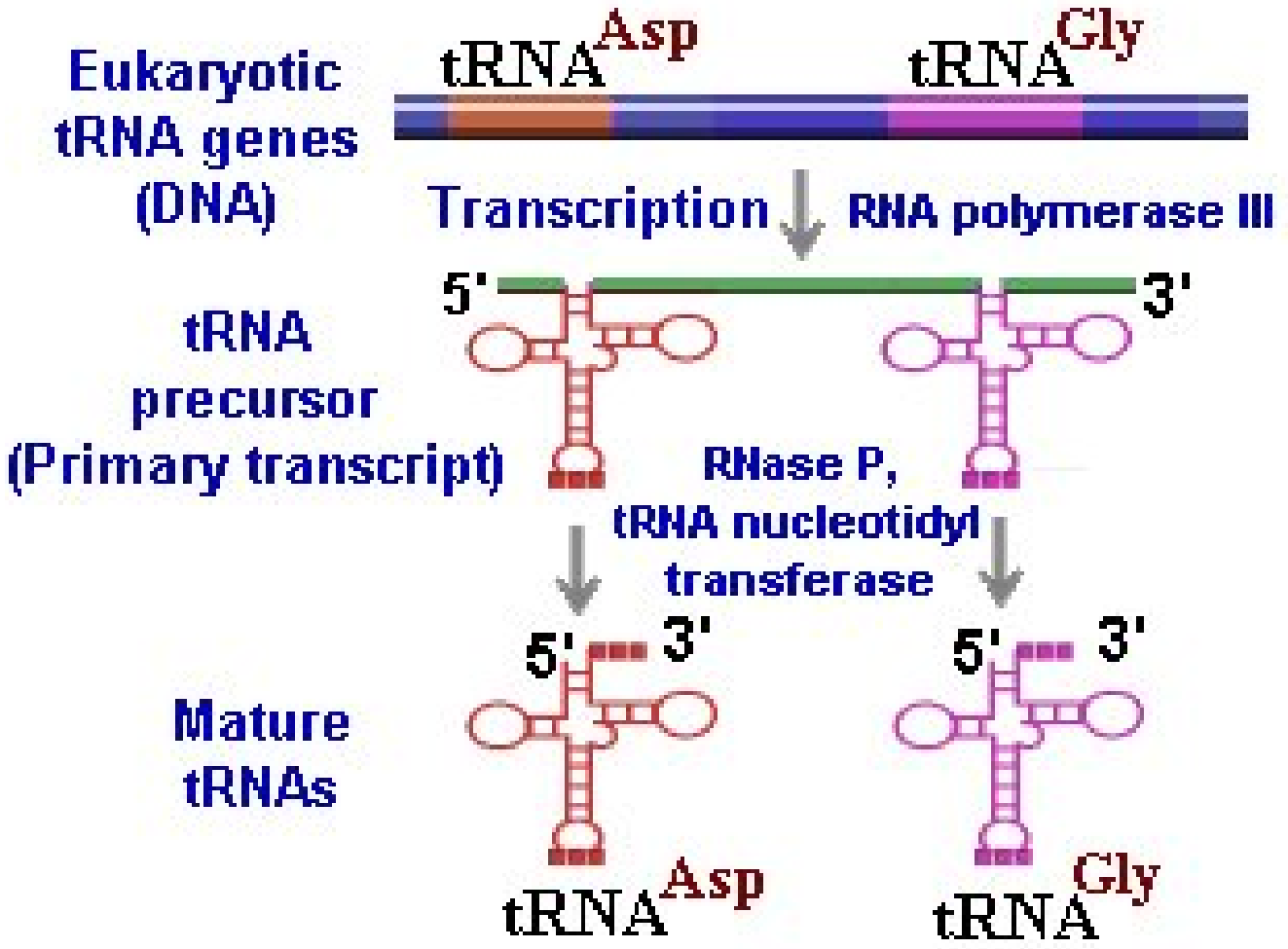
Figure 1. The Mammalian pre-mRNA 3′ End Processing Complex

Experimentally demonstrated protein:protein interactions are indicated by double-headed arrows. pA indicates the poly(A) addition site.

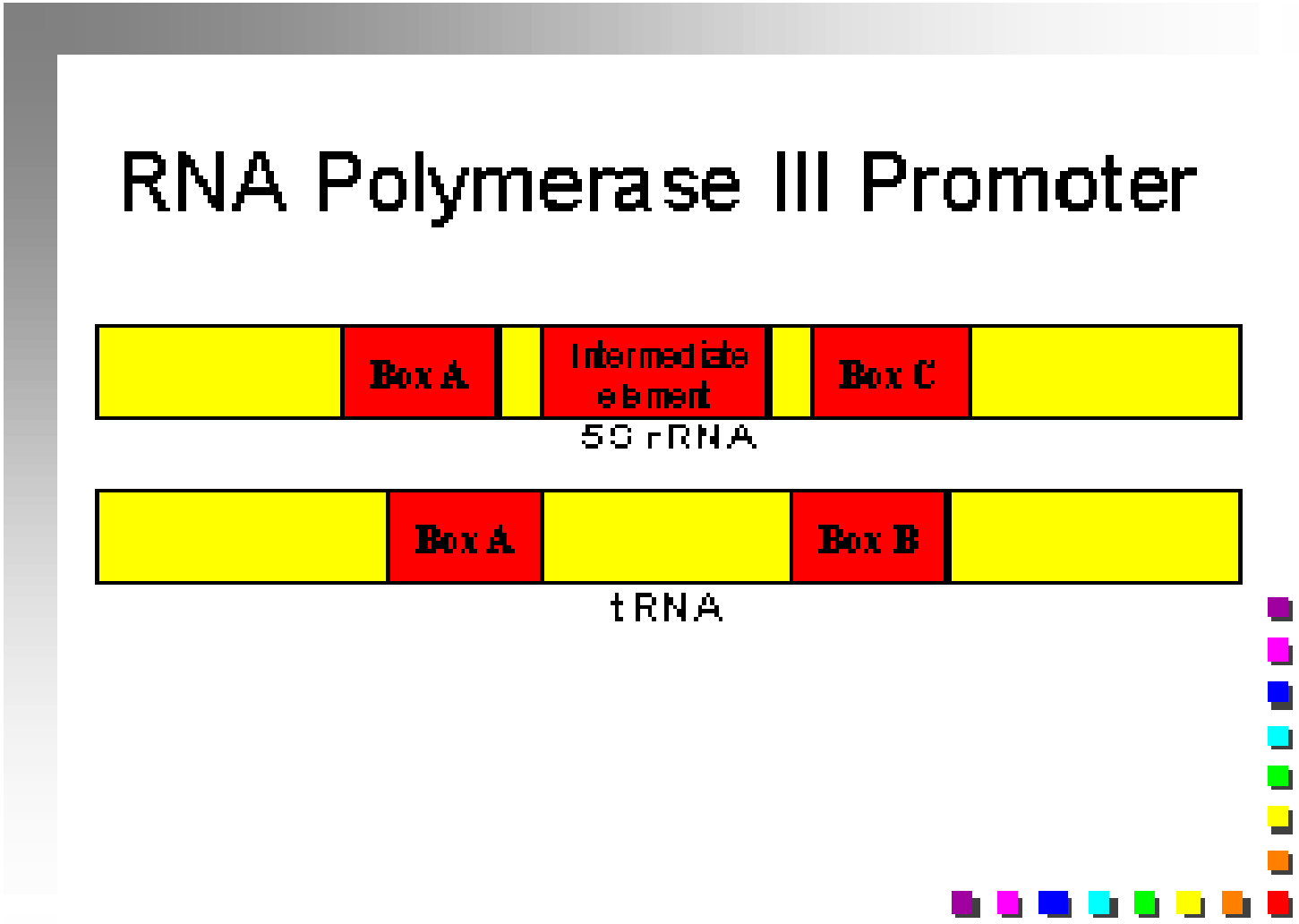
Funktion der Poly-Adenylierung?

- Stabilität der mRNA
- Translatierbarkeit

Polymerase III – Gene: tRNAs, 5S rRNA, snRNAs

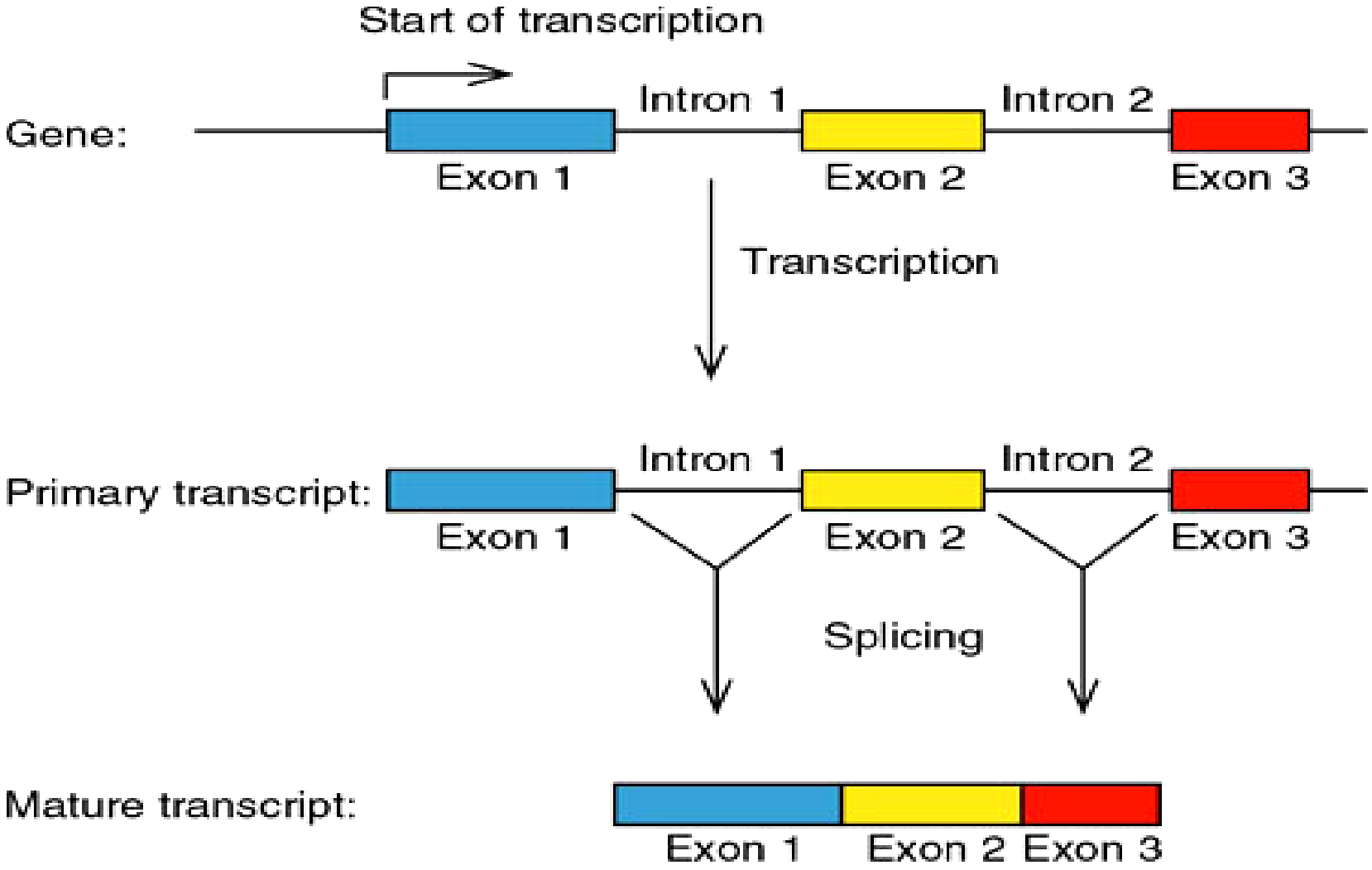


RNA Polymerase III -Gene
haben **interne!** Promotoren
d. h. die Promotoren liegen im transkribierten Bereich



Splicing (Spleißen)

wichtiger Teil der RNA-Prozessierung; die Intronabschschnitte werden aus dem Primärtranskript entfernt



Je nach Spleißmechanismus werden vier verschiedene Gruppen von Introns unterschieden:

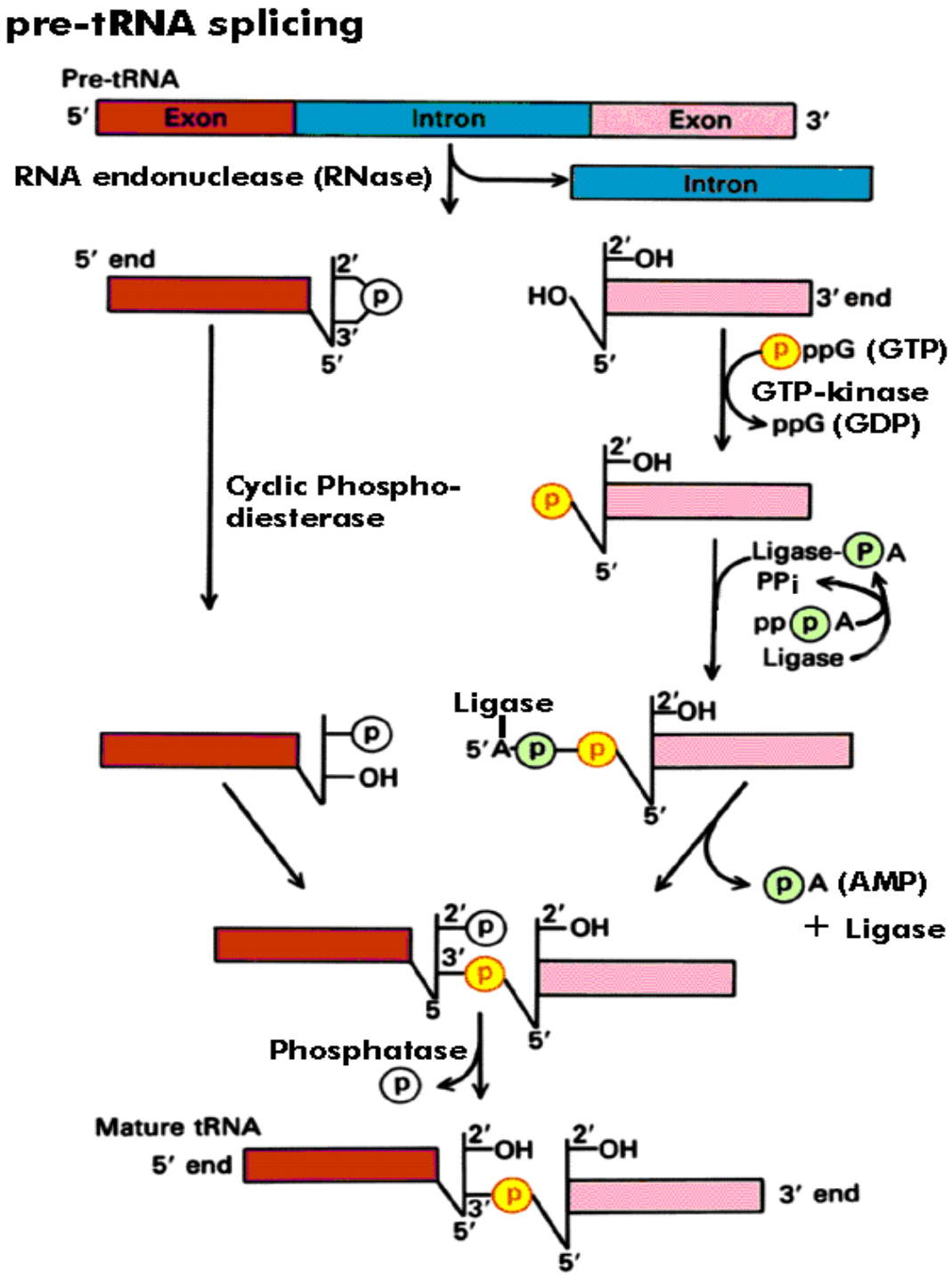
- tRNA Introns
- Autokatalytische Introns Gruppe I
- Autokatalytische Introns Gruppe II
- hn-/mRNA Introns

tRNA Splicing:

= zwei Stufenprozess

1. Herausschneiden des Introns durch Endonuklease

2. Verknüpfen der Exons durch RNA-Ligase

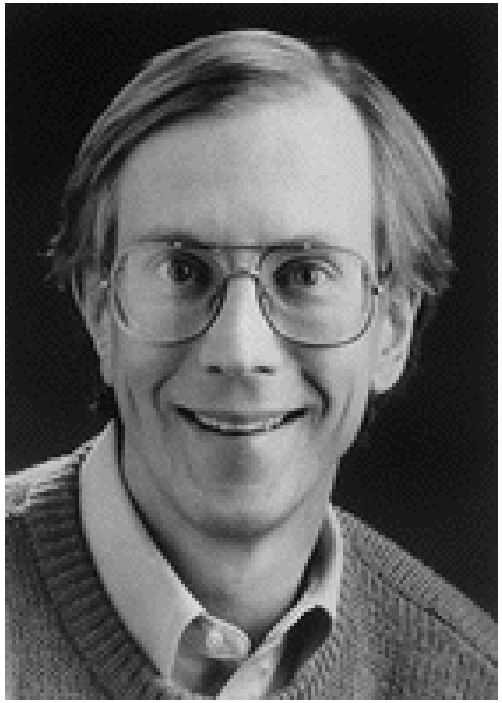


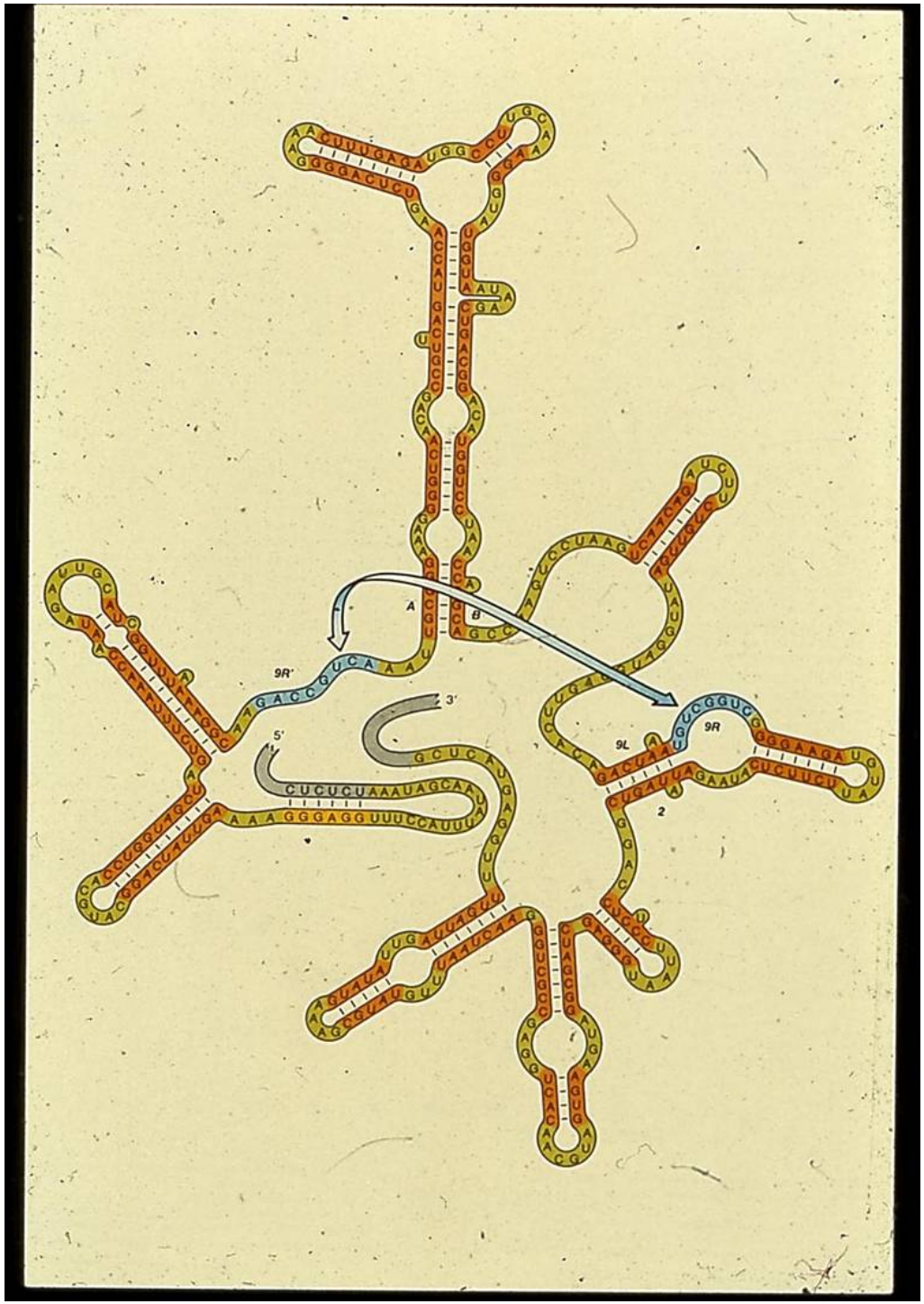
Autokatalytisches Splicing
Ribozyme

Beim **autokatalytischen Spleißen** sorgt die Intron-RNA selbst (autokatalytisch) dafür, dass die RNA an den Intron-Exon-Grenzen geschnitten und die beiden Exon-Enden (3´-Ende von Exon n mit dem 5´-Ende von Exon m) über eine Phosphodiesterbindung verknüpft werden. Weil die RNA bei diesem Prozess wie ein Enzym katalytisch aktiv ist, werden diese RNAs auch als **Ribozyme** bezeichnet

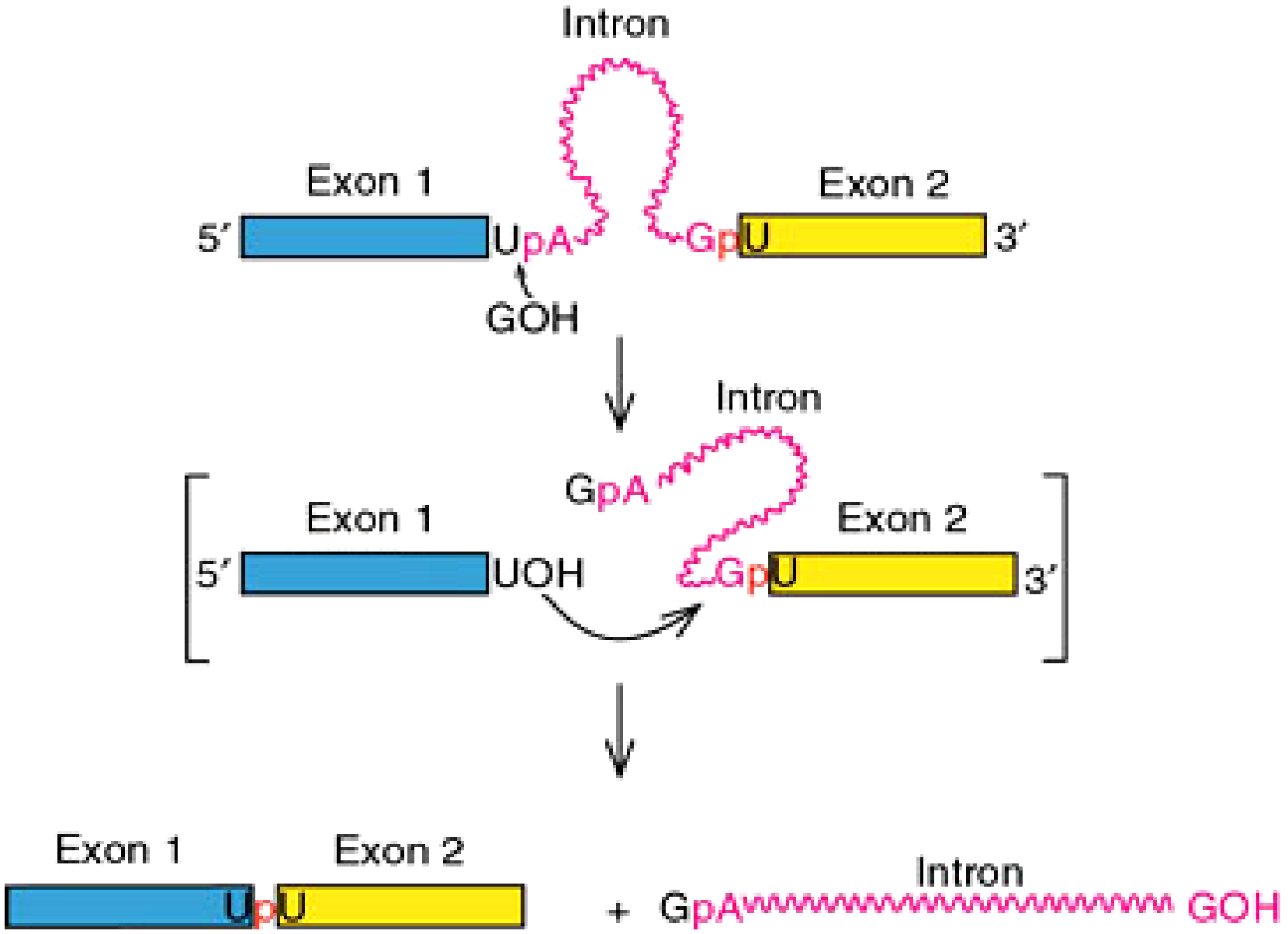
Ribozym-Struktur

Entdecker der Ribozyme
Th. R. Cech
Nobelpreis 1980

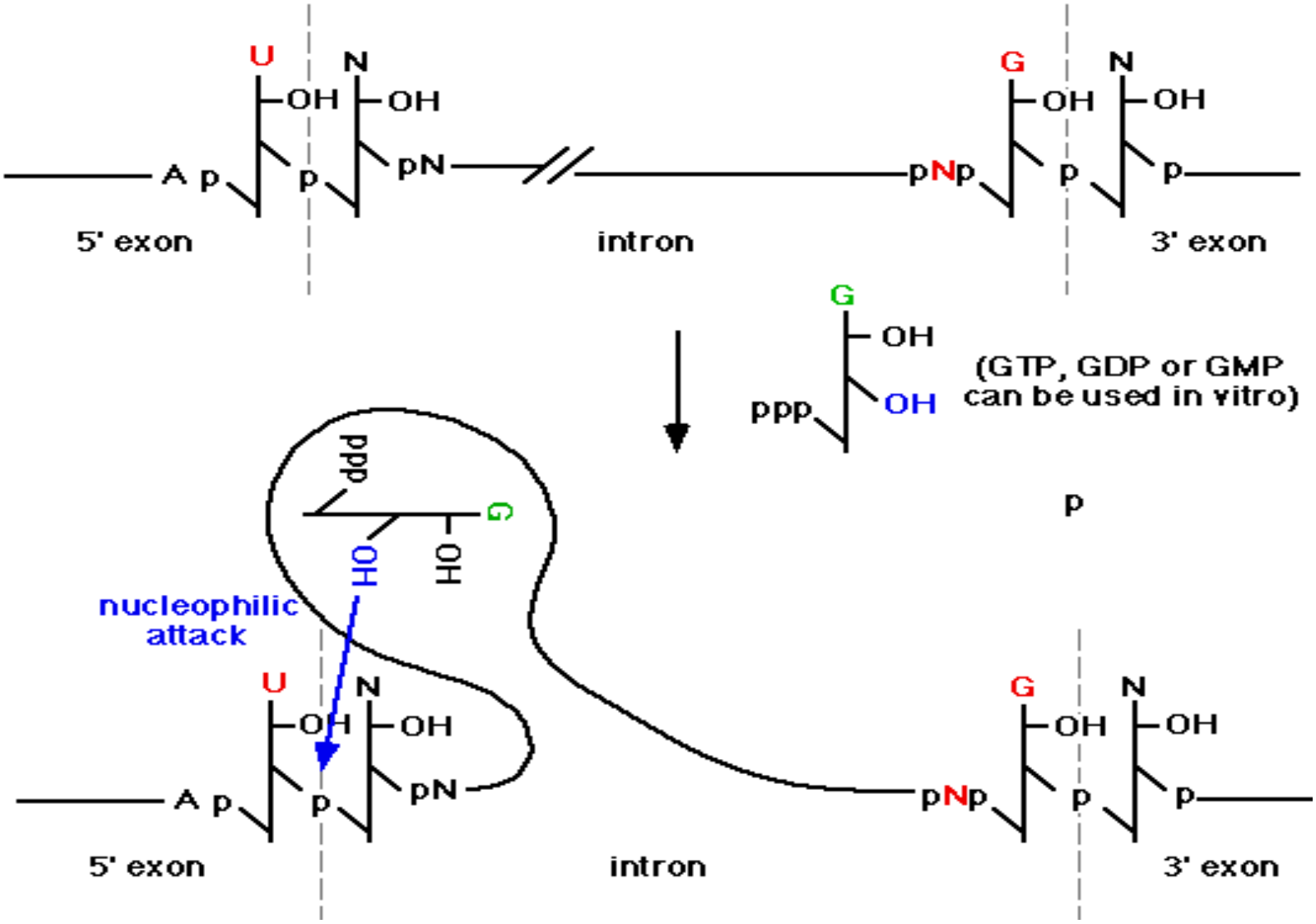




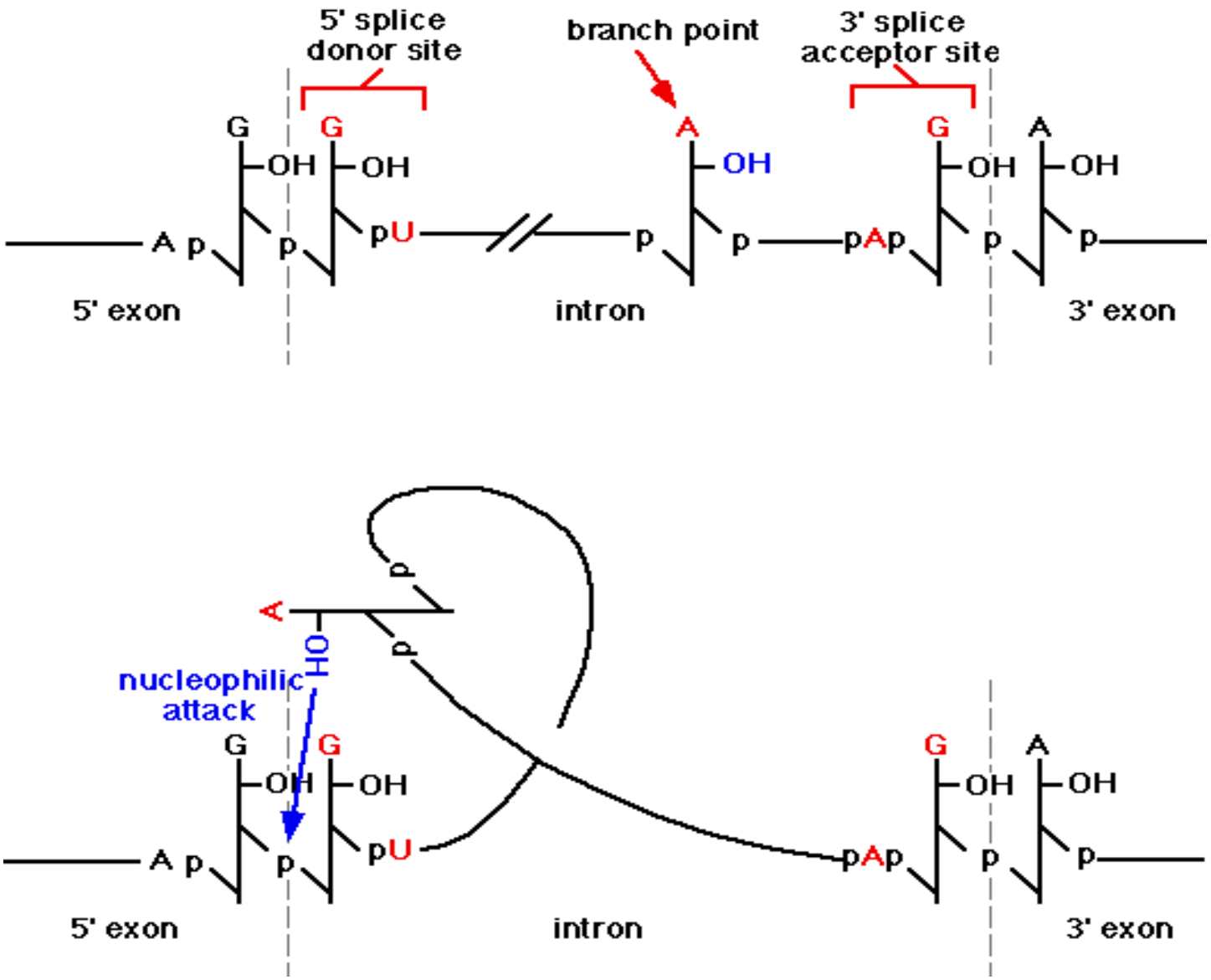
Autokatalytisches Spleißen der Gruppe I Introns bei Prä-rRNA von Tetrahymena



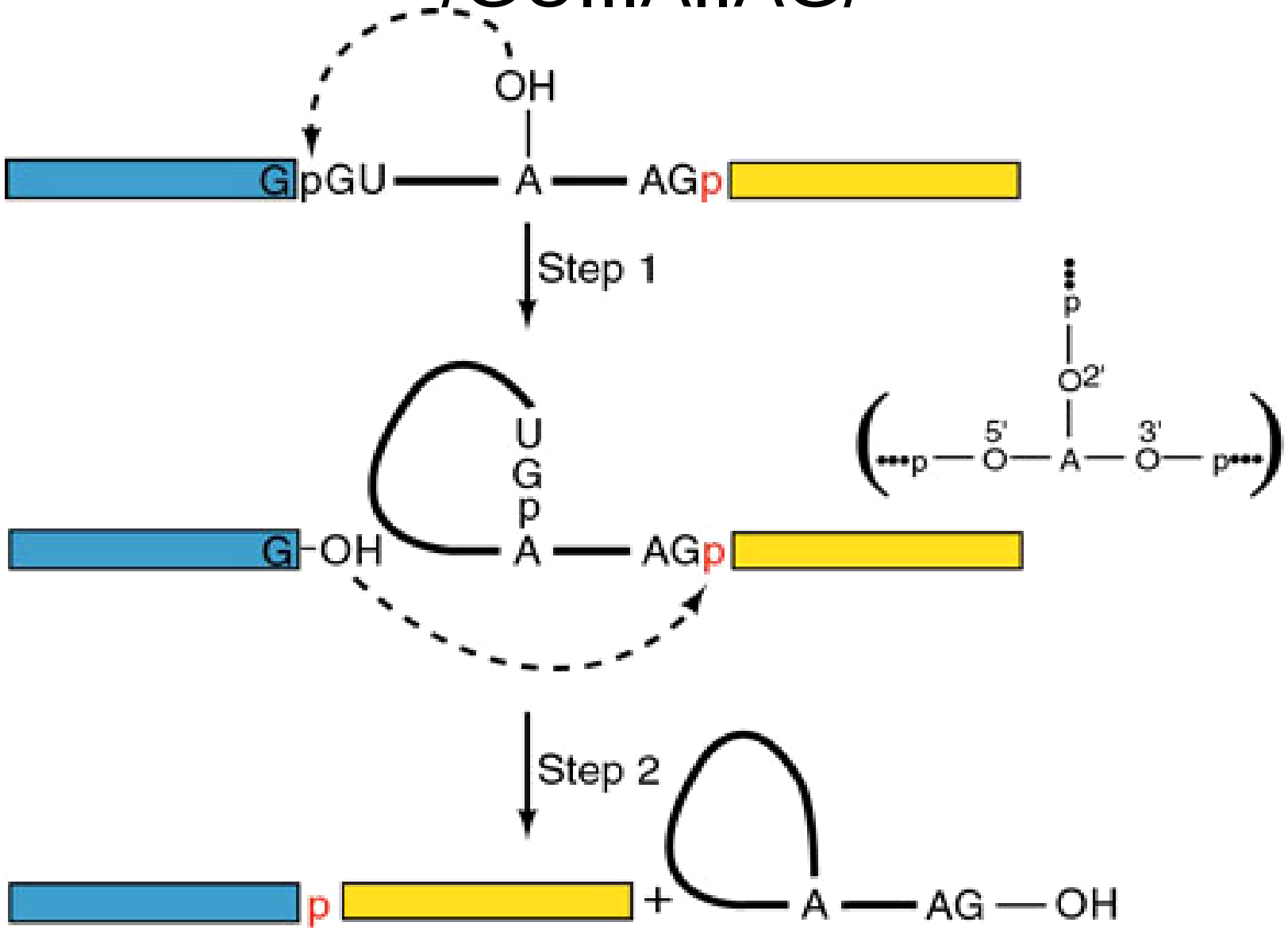
Mechanismus der autokatalytisch spleißenden Gruppe I Introns



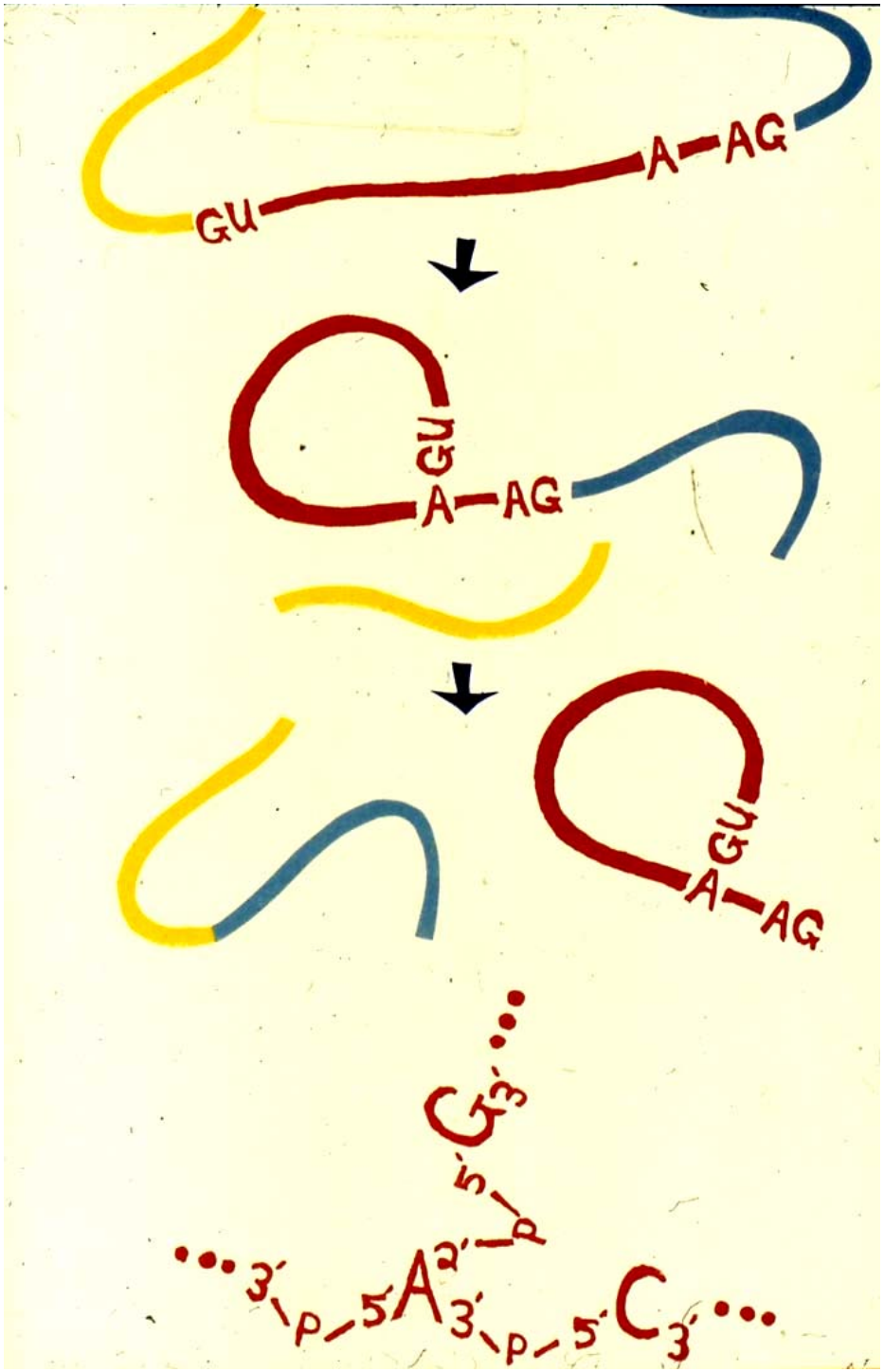
Autokatalytisches Splicing Gruppe II Introns



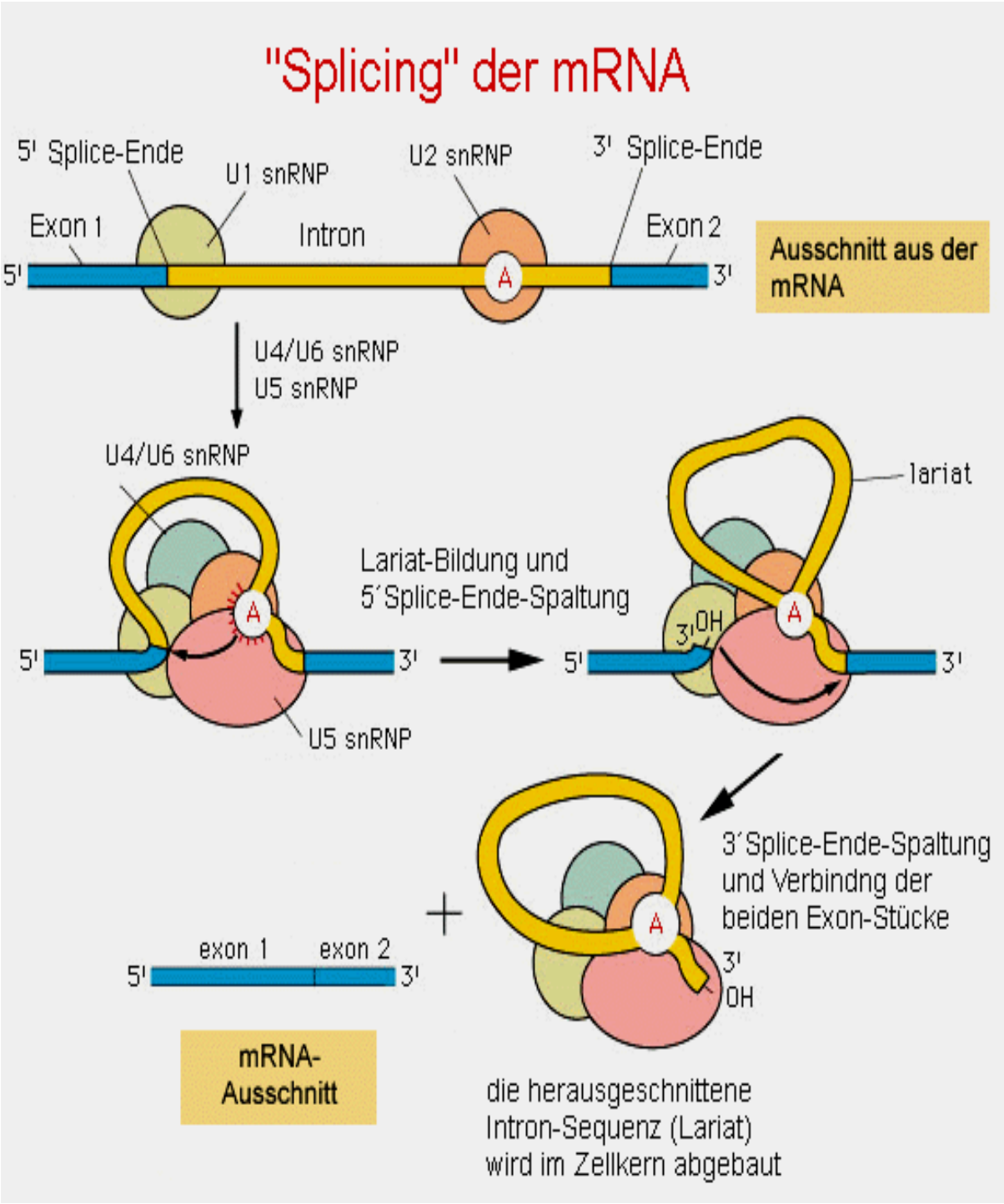
mRNA-Spleißen an der „Consensus Splice Site“ /GU...A..AG/



Beim Spleißen bildet sich ein „Lariat“ im heraus gelösten Intron über eine 2´-5´ Phospho-diesterbindung

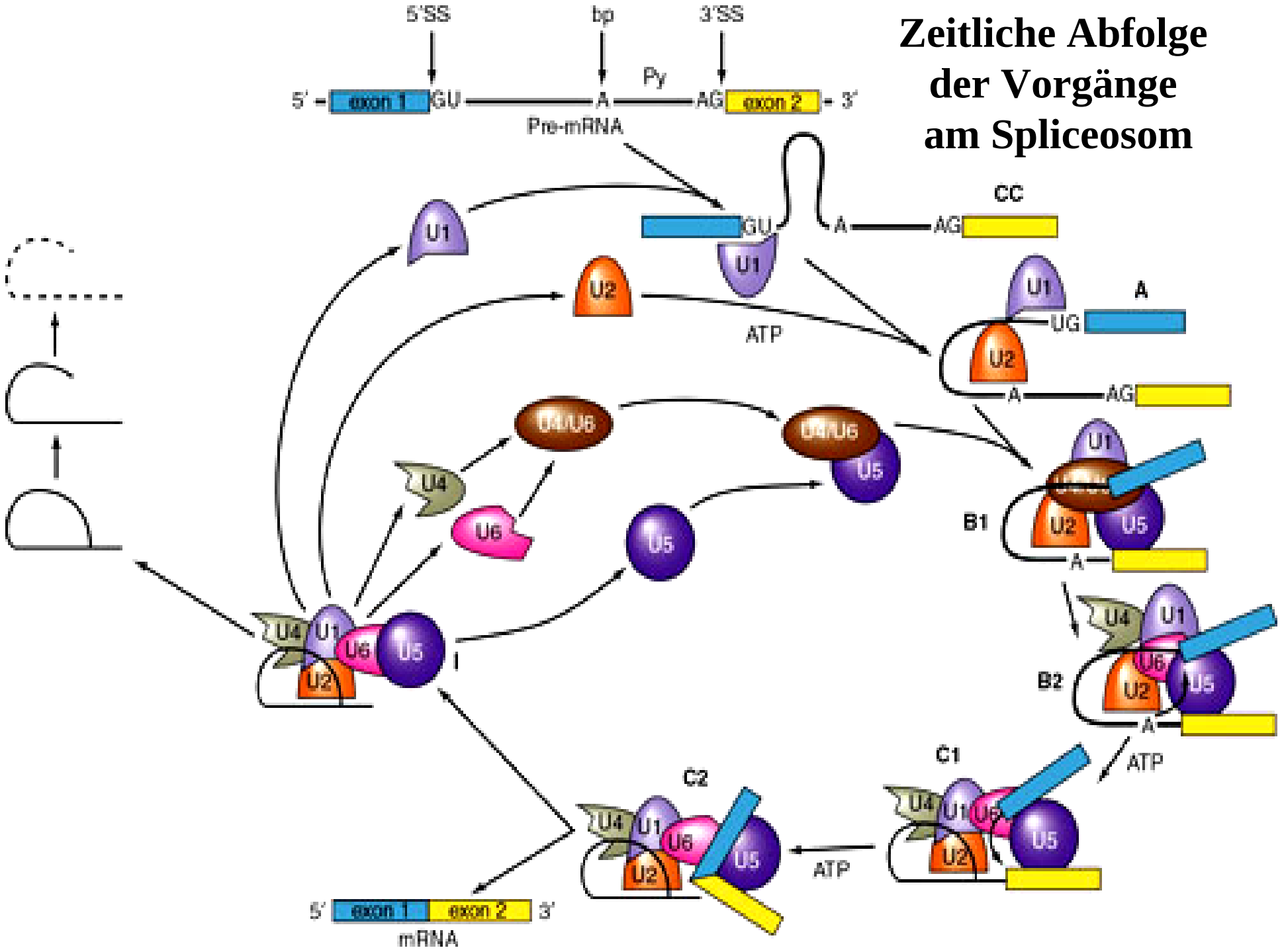


Am Spleißen von mRNAs sind Spliceosomen mit „SN(U)RPS“ beteiligt



snRNPs (SNURPS) enthalten die
snRNAs (sn= „small nuclear“)
U1, U2, U4/6 und U5
(snRNPs= small nuclear ribonucleoprotein)

Zeitliche Abfolge der Vorgänge am Spliceosom



Zusammenfassung

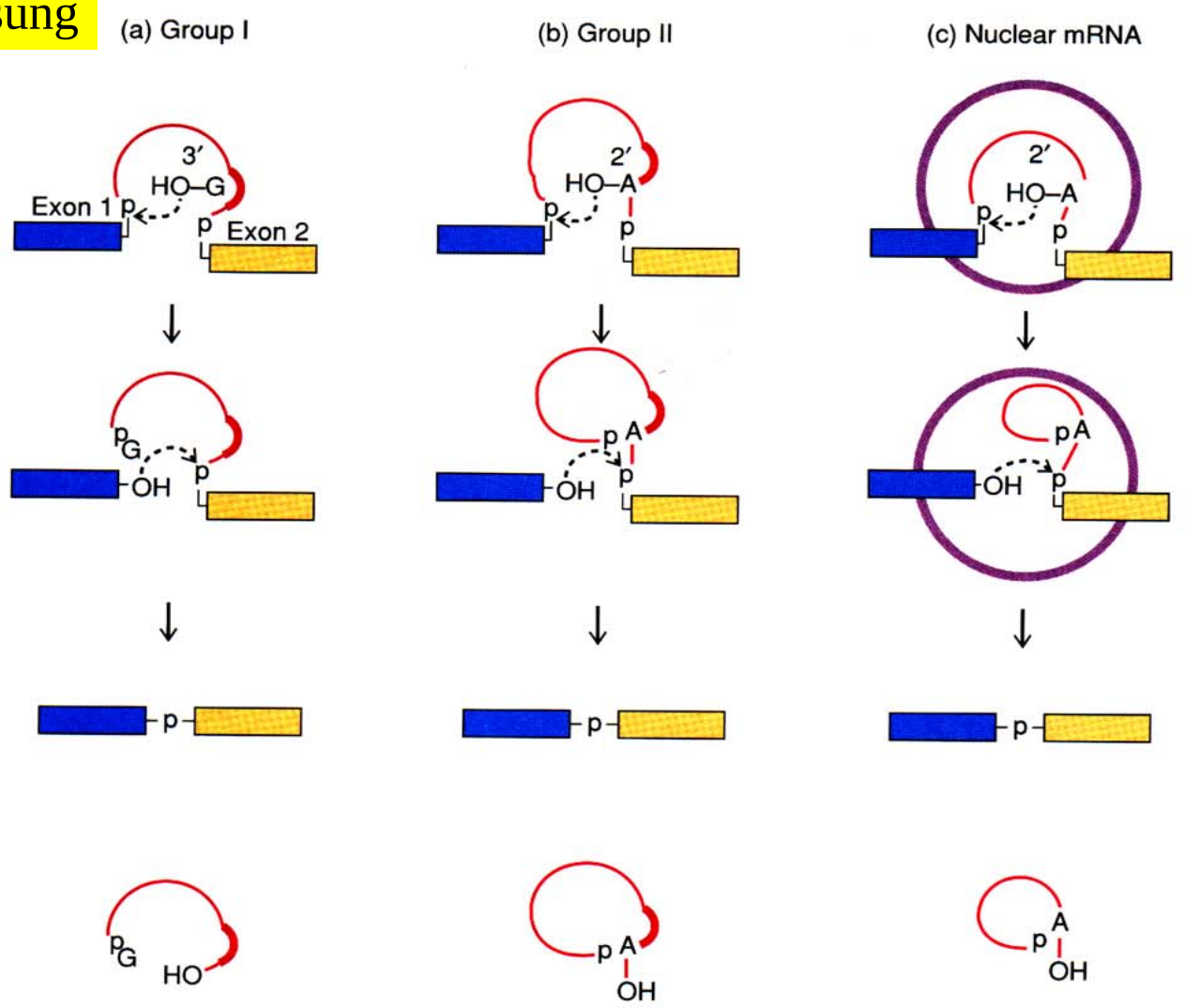
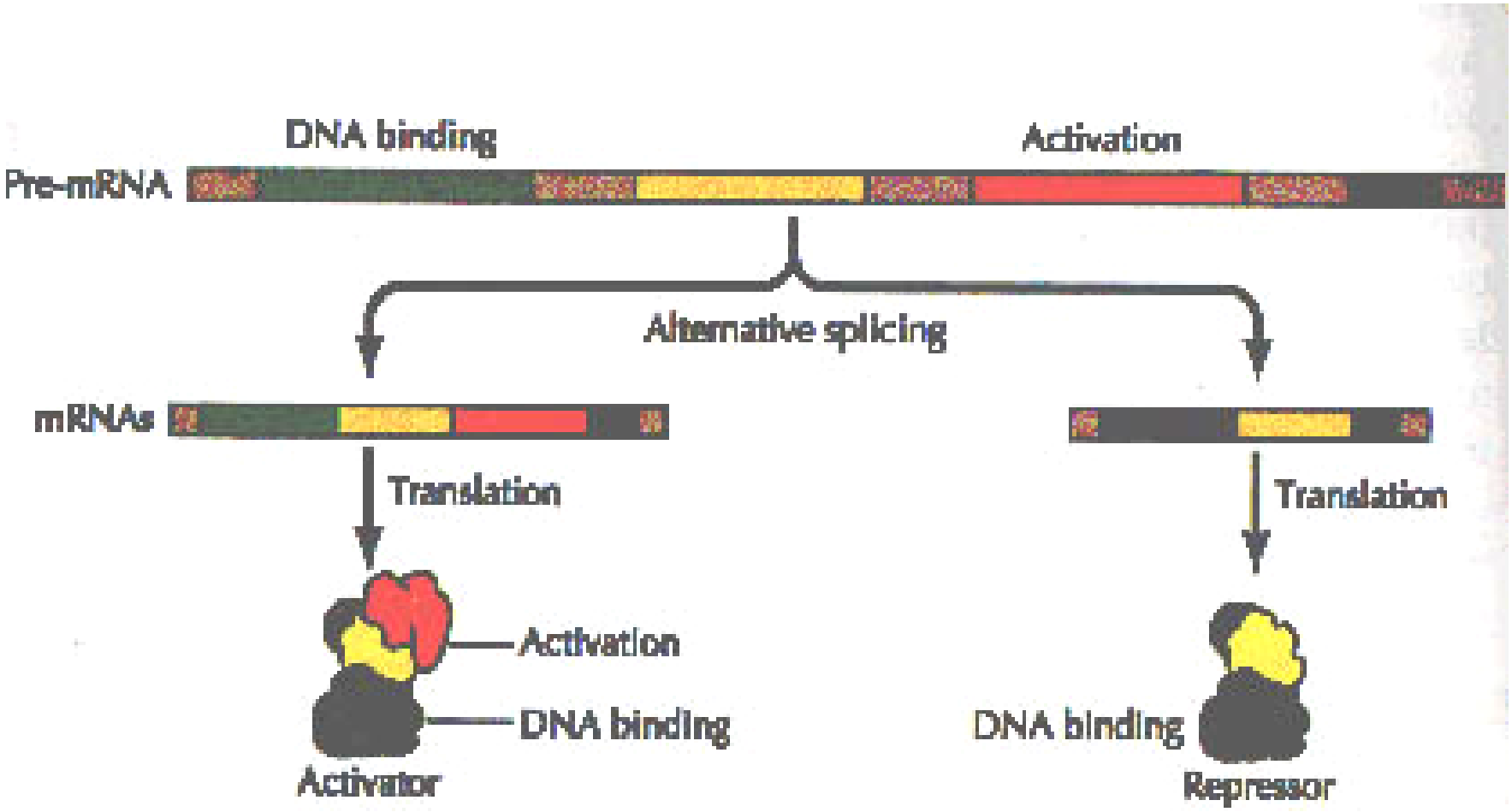
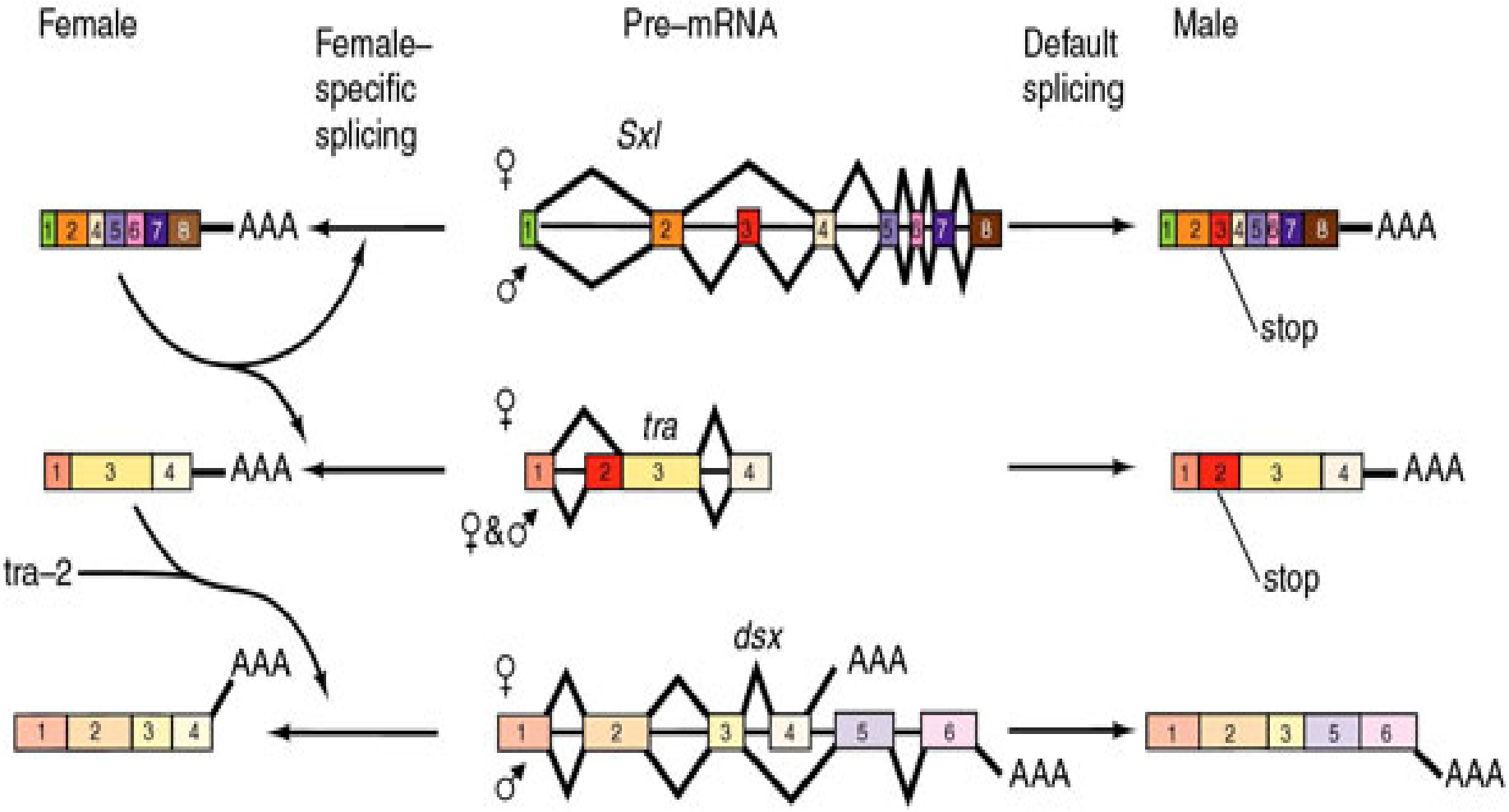


FIGURE 9.42 Summary of three splicing schemes. The major differences in these mechanisms lie in the first step. The self-splicing of group I introns (*a*) is initiated by a guanosine nucleotide that presumably resides in a pocket in the intron (represented by a thickened semicircle). This guanosine attacks the phosphate linking exon 1 (blue) and the intron (red). In group II (*b*), an adenosine nucleotide that is part of the intron itself plays this initiation role, resulting in a lariat-shaped intermediate. This adenosine is represented as adjacent to a pocket similar to the one in group I introns that harbors the initiating guanosine. Nuclear mRNA precursors (*c*) follow a splicing scheme remarkably similar to that used by group II introns. The major difference is that nuclear mRNA splicing requires a spliceosome (purple).

Alternatives oder differenzielles Splicing erhöht die Zahl der Proteine



Alternatives Spleißen bestimmt bei *Drosophila melanogaster* das Geschlecht



Neues Thema:
Steuerung der Transkription

- Die Aktivierung von Genen erfolgt durch **Transkriptionsfaktoren** (TFs)
- Es gibt **basale TFs** (immer vorhanden und für jede Transkription notwendig) und **spezifische TFs** (gewebs-/zellspezifisch; hormoninduziert; entwicklungsspezifisch etc.)
- Jede Genklasse hat eigene TFs

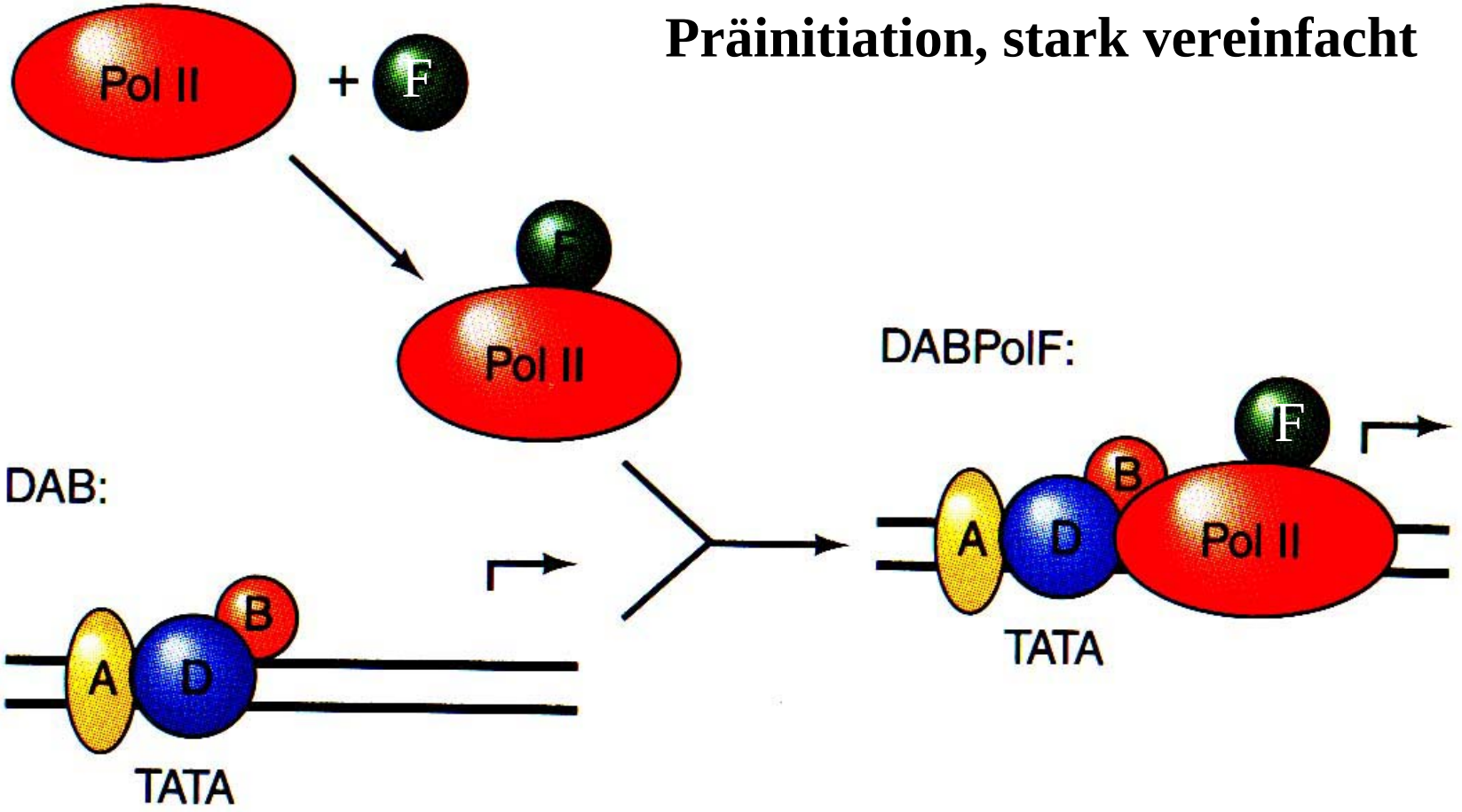
Die basalen Transkriptionsfaktoren:

- Je nach Genklasse werden die Transkriptionsfaktoren TF IA, TF IB ...;

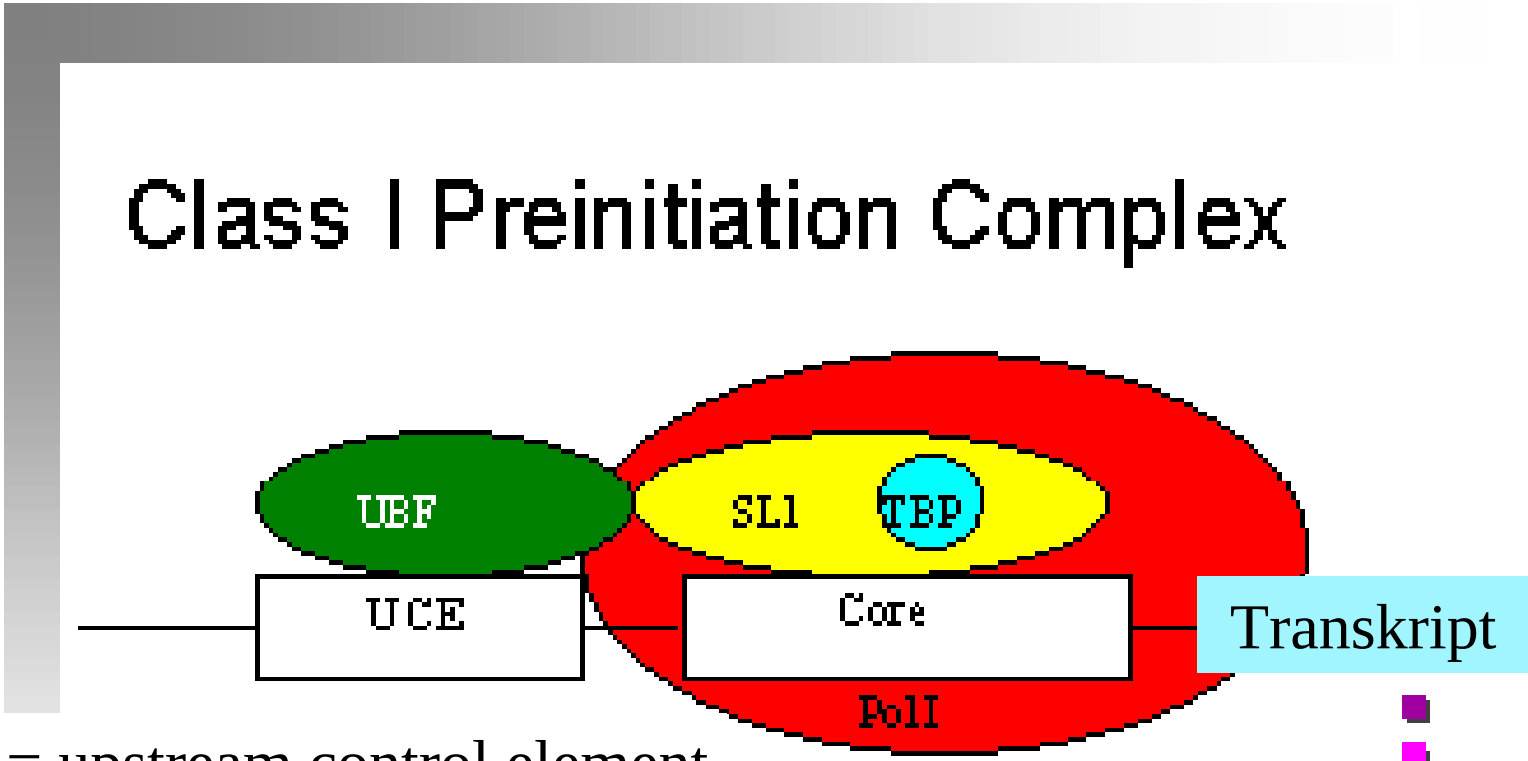
  TF IIA, TF IIB ..; TF IIIA etc. bezeichnet

Daneben gibt es eine Reihe anders benannter Proteine, die die Genaktivität steuern und nicht immer Teil des basalen Transkriptionskomplexes sind (z. B. SP1).

Der Basale Transkriptionskomplex (Präinitiationskomplex) der Pol II – Gene:
TF IID. TF IIA. TF IIB + Pol II-TFIIF



RNA Polymerase I Promotor und Initiationskomplex



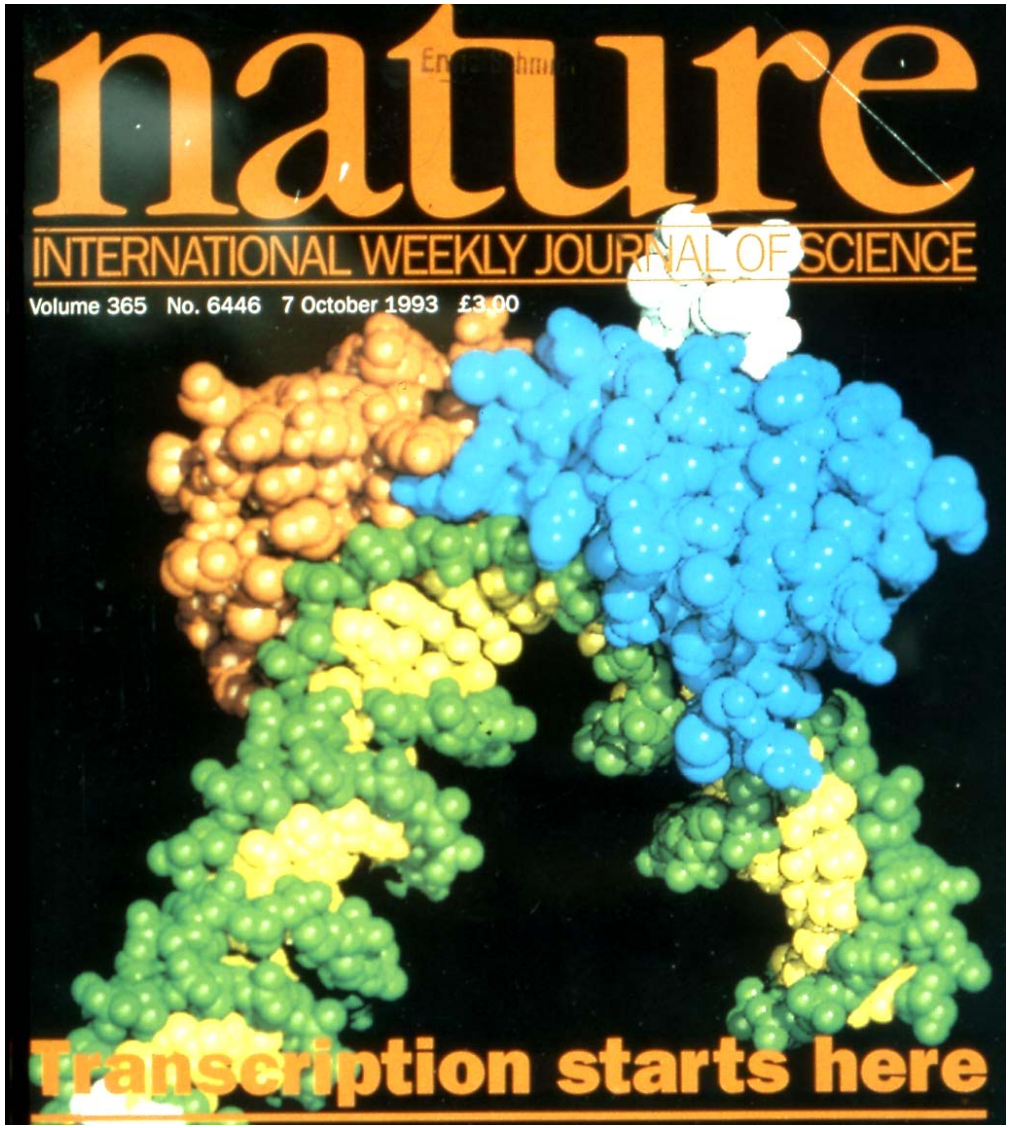
= upstream control element

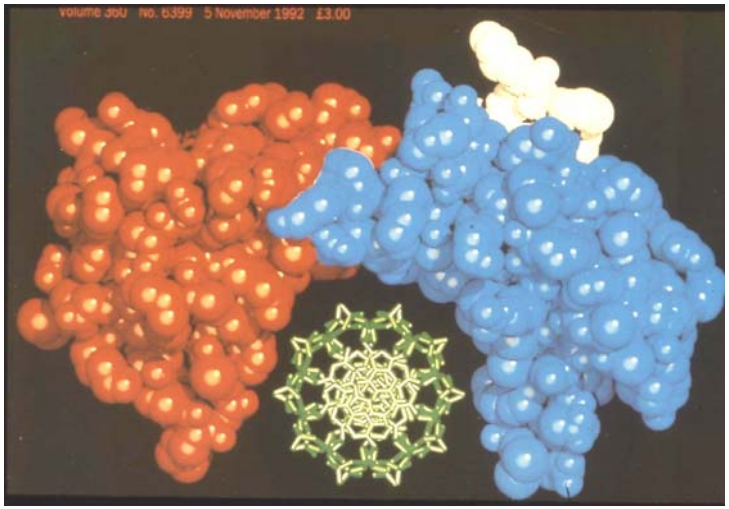
RNA Polymerase I Gene
haben einen 5´ aufwärts Promotor

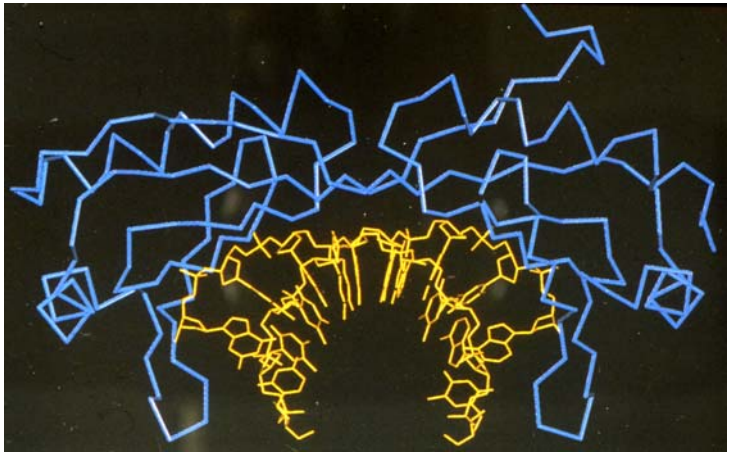
Besondere Rolle von TF IID:

- **TF IID** enthält als Untereinheit das **TBP** („TATA-Box binding Protein). Das TBP erkennt die TATA-Box und bindet als erstes Protein an den Gen-Promotor. Erst danach erfolgt die Bindung der anderen TFs und schließlich zuletzt die der RNA-Polymerase II in Verbindung mit TF IIF.

TATA-Box binding protein (TBP)



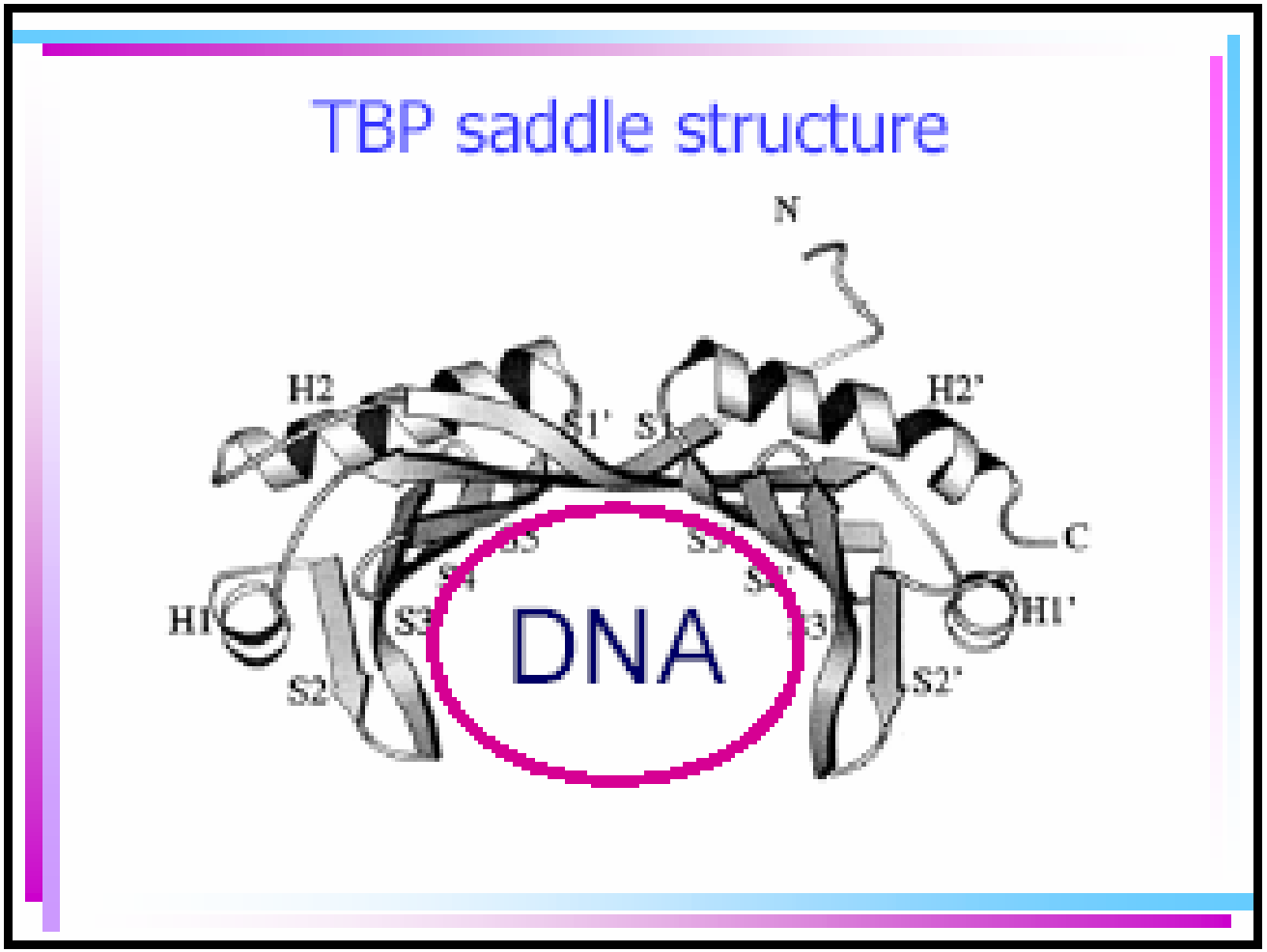




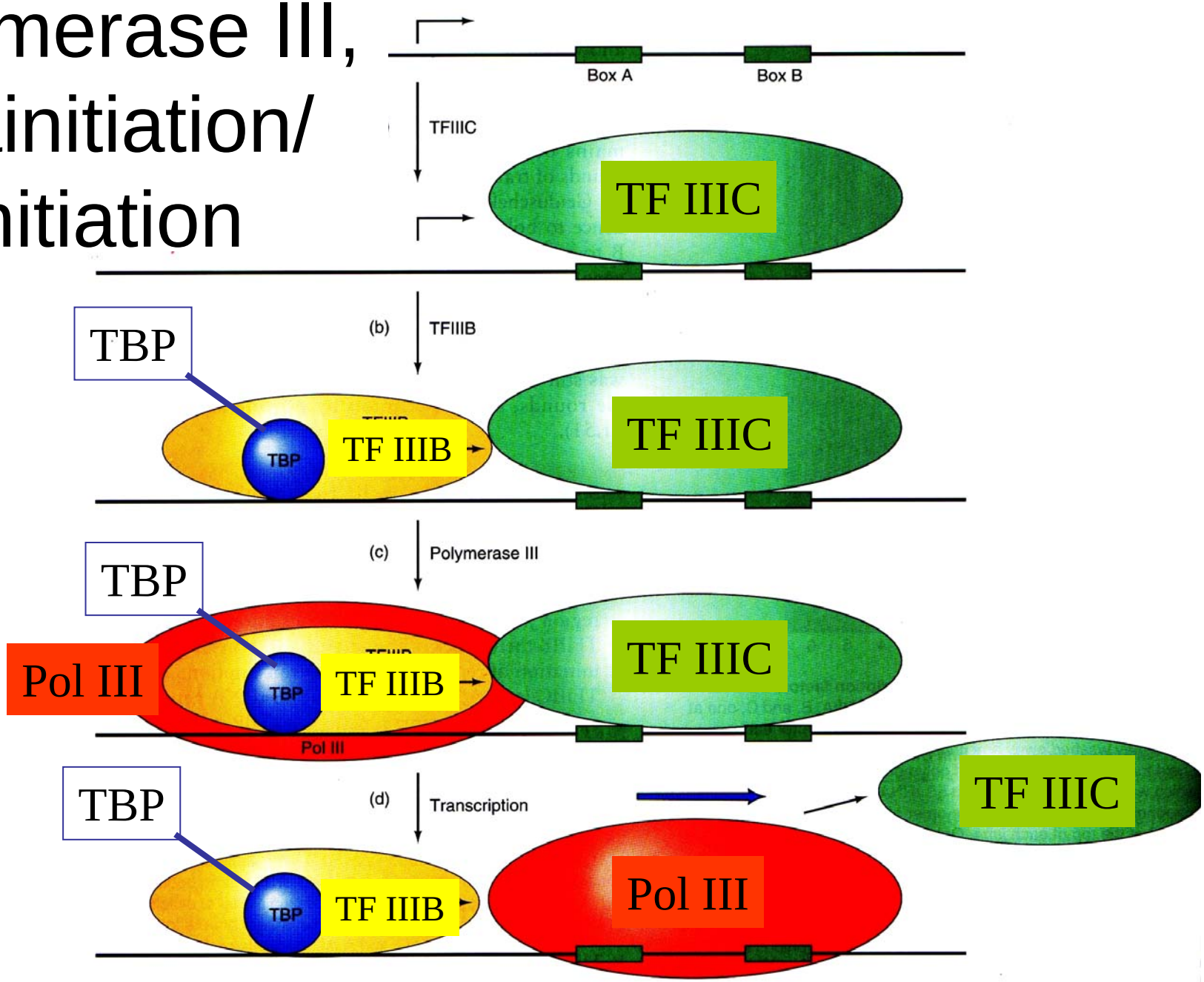
TBP (TATA-box binding protein)

- TBP bindet im Gegensatz zu den meisten DNA-bindenden Proteinen in der „kleinen Grube“ der DNA
- TBP krümmt die DNA durch die Bindung und verursacht so einen scharfen „Knick“
- TBP vermittelt die Bindung weiterer TFs an den Promotor
- TBP ist auch bei Genen ohne TATA-Box am Präinitiationskomplex beteiligt, und zwar auch bei Pol I- und Pol III-Genen

Sattel-Struktur des TBP auf der DNA



Polymerase III, Präinitiation/ Initiation

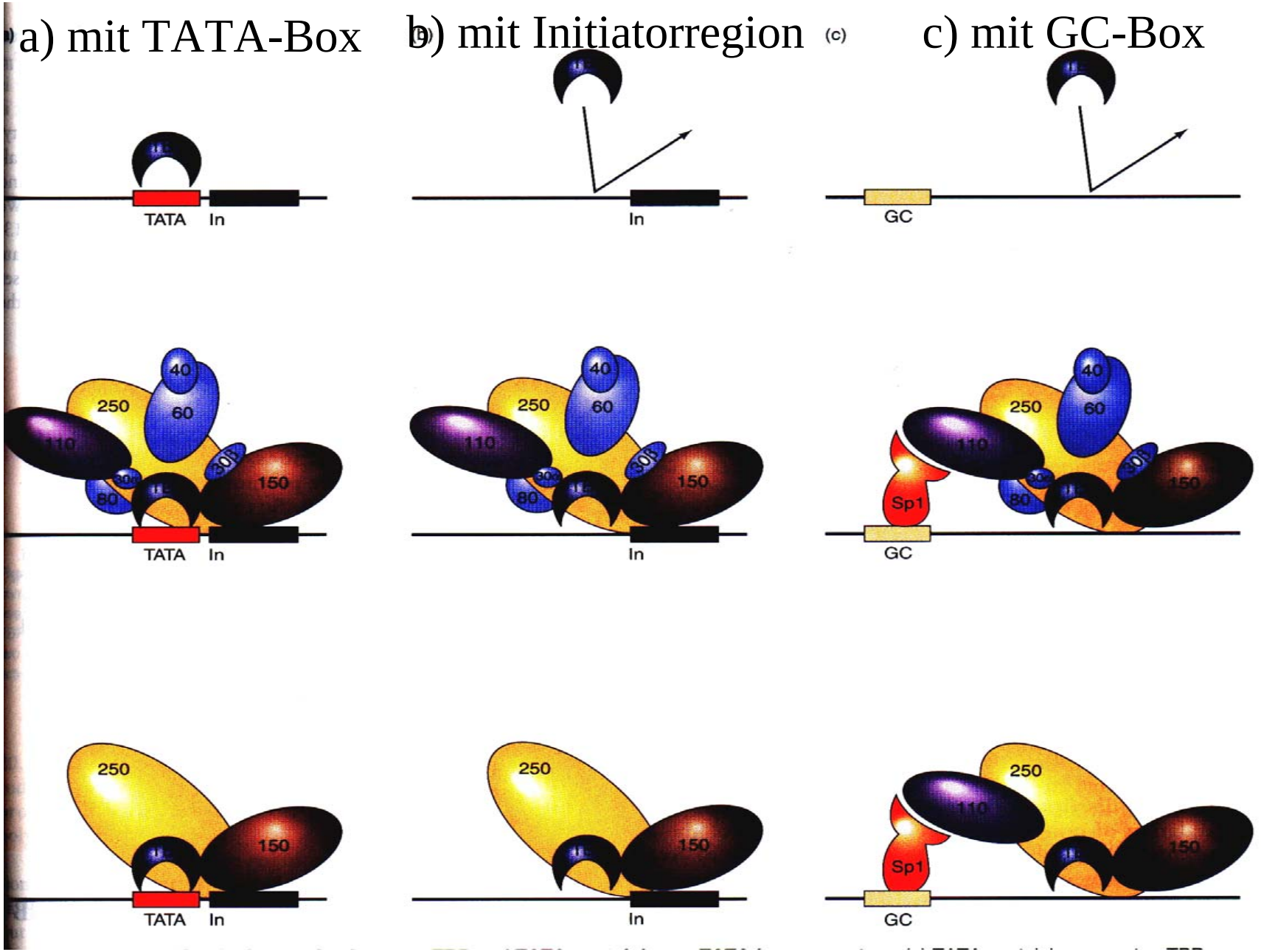


Präinitiation bei Genen

a) mit TATA-Box

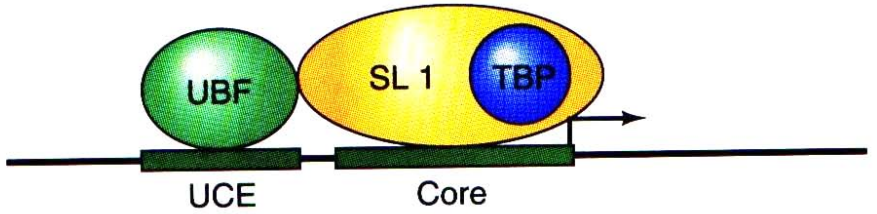
b) mit Initiatorregion

c) mit GC-Box

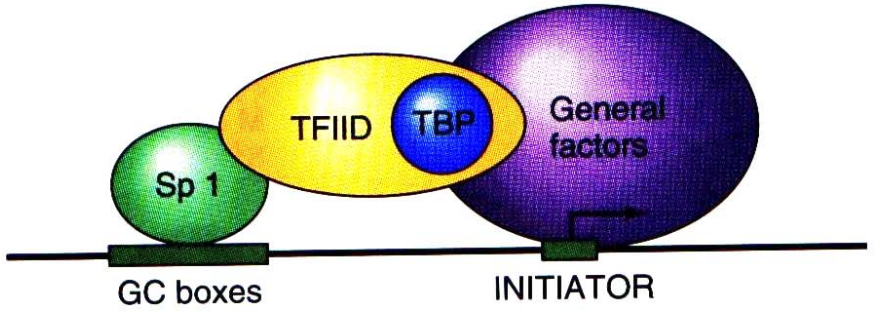


Zusammen-fassung: Präinitiations-komplexe der verschiedenen Genklassen

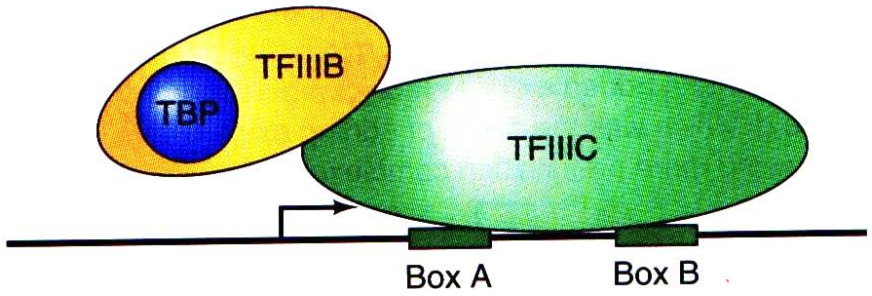
CLASS I
rRNA

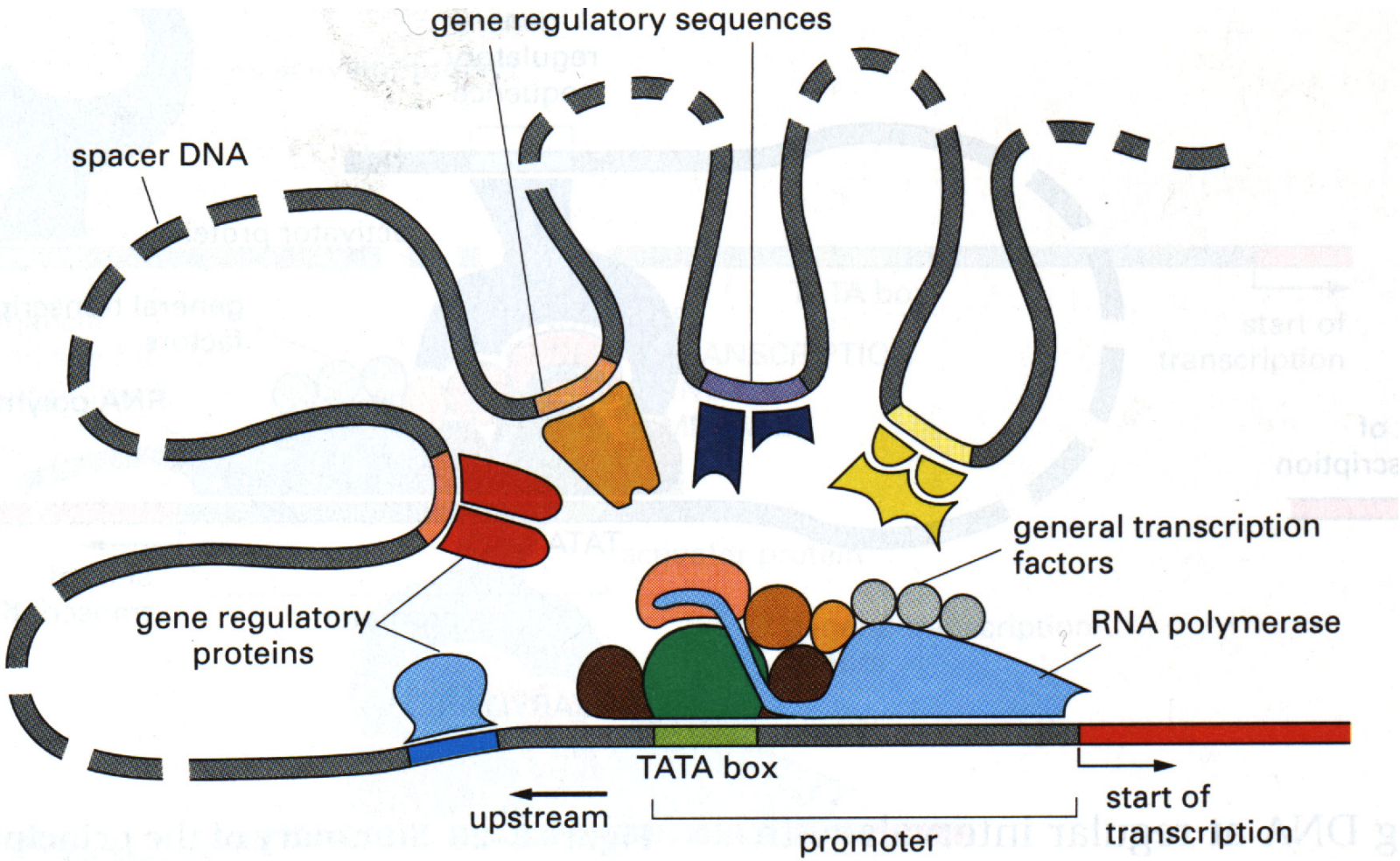


CLASS II
G_6I

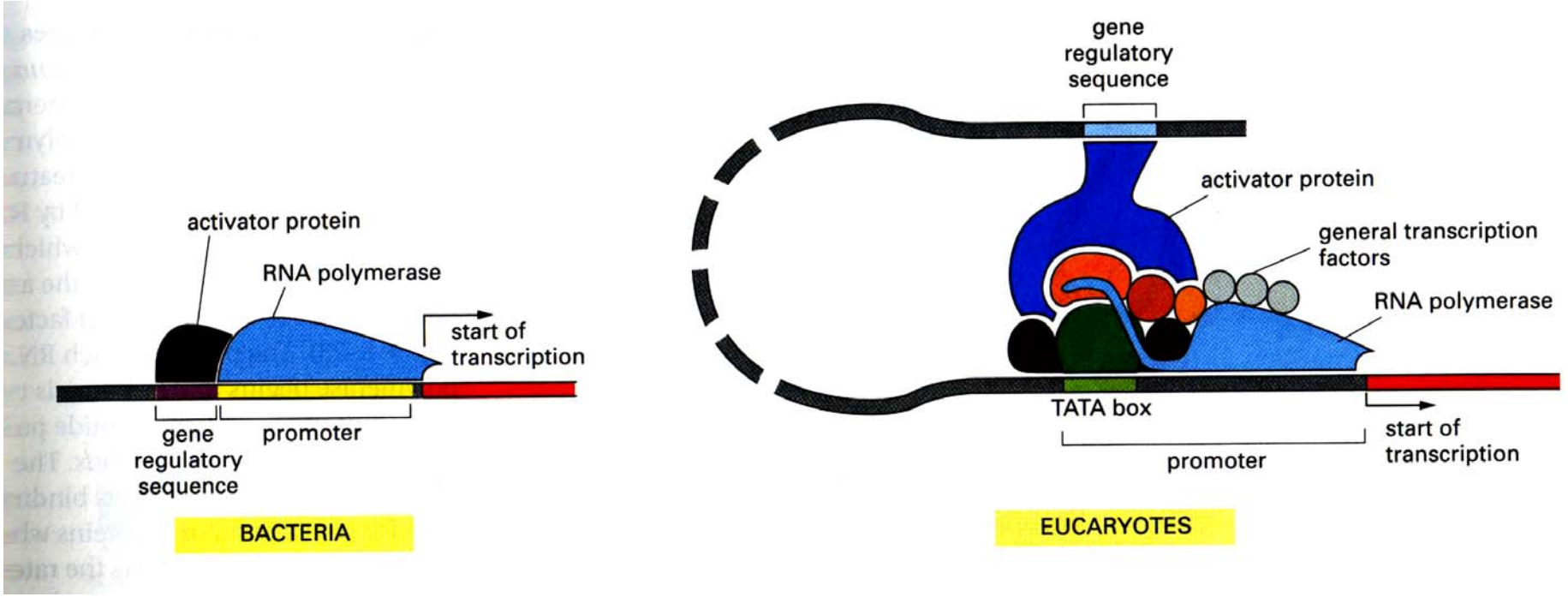


CLASS III
VA_I




gene regulatory sequences
spacer DNA
general transcription factors
RNA polymerase
gene regulatory proteins
TATA box
upstream
start of transcription
promoter

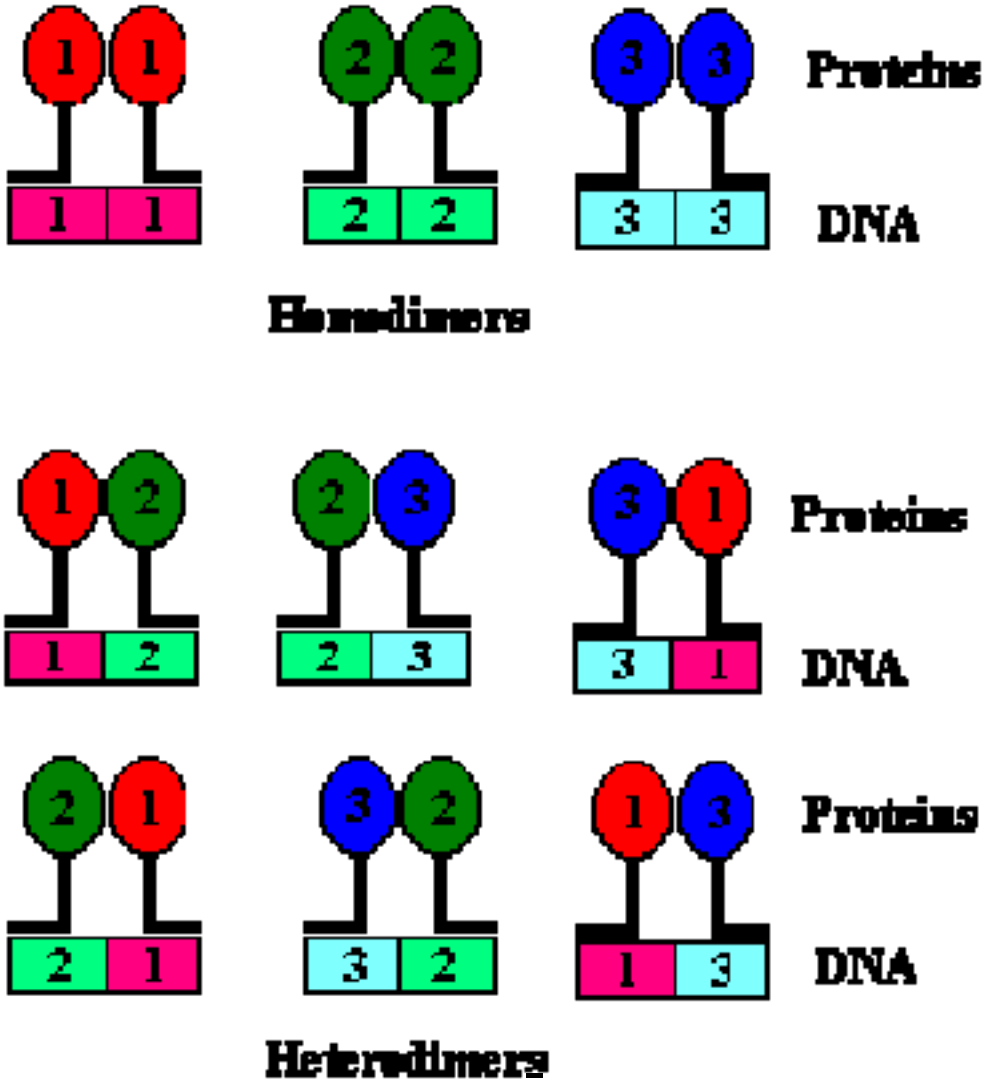
Vergleich Prokaryoten - Eukaryoten



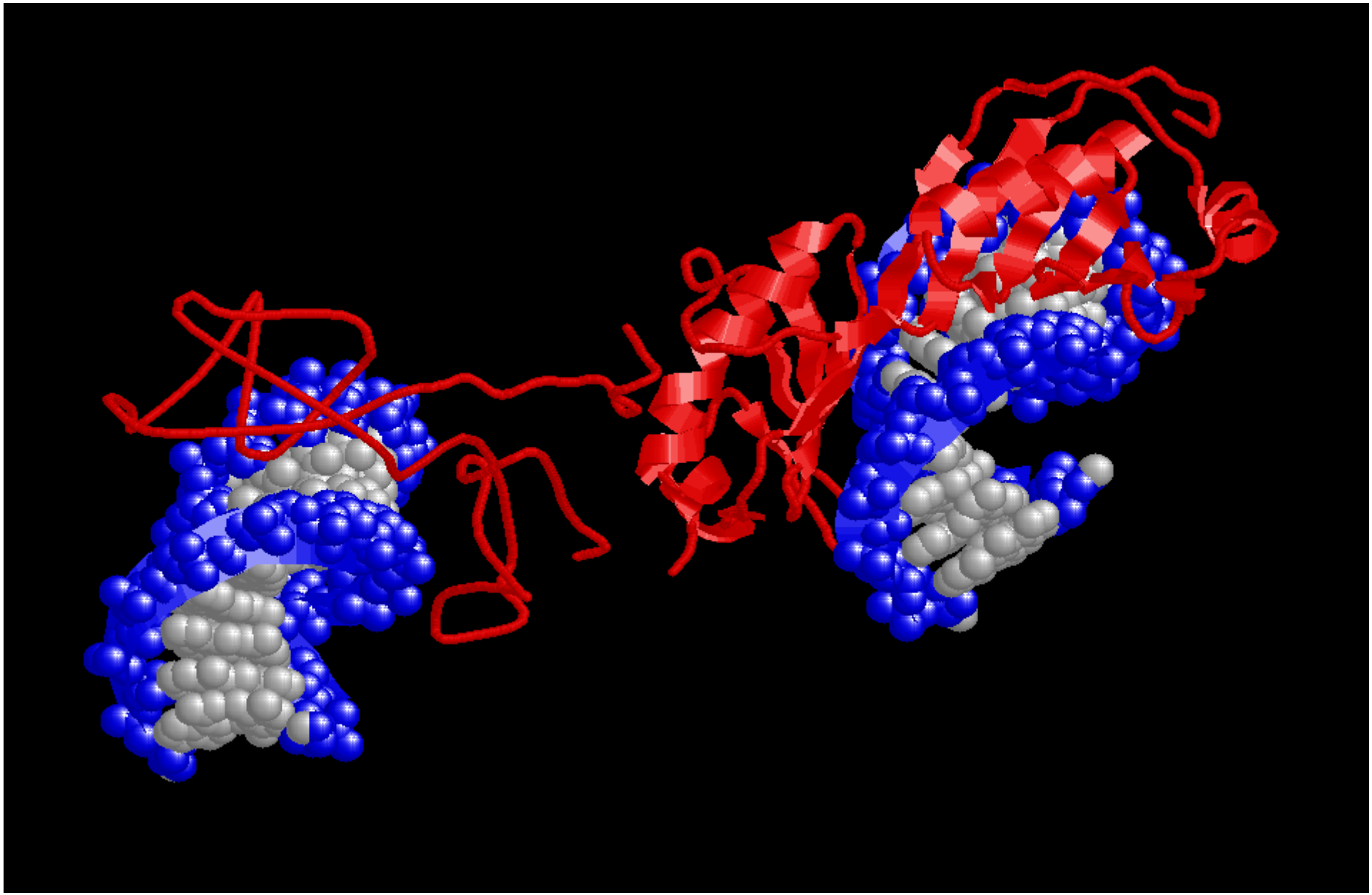
DNA-bindende Proteine:

- Die wichtigsten „Interpreter“ des DNA-Kommando-Codes
- Die Vermittler zwischen ankommenden Signalen und Umsetzung durch die Gene
- „transaktive“ Steuerungselemente von Genen oder ganzen Gengruppen
- Globale oder lokale Modifikatoren der Chromatinstruktur und damit der Genaktivität

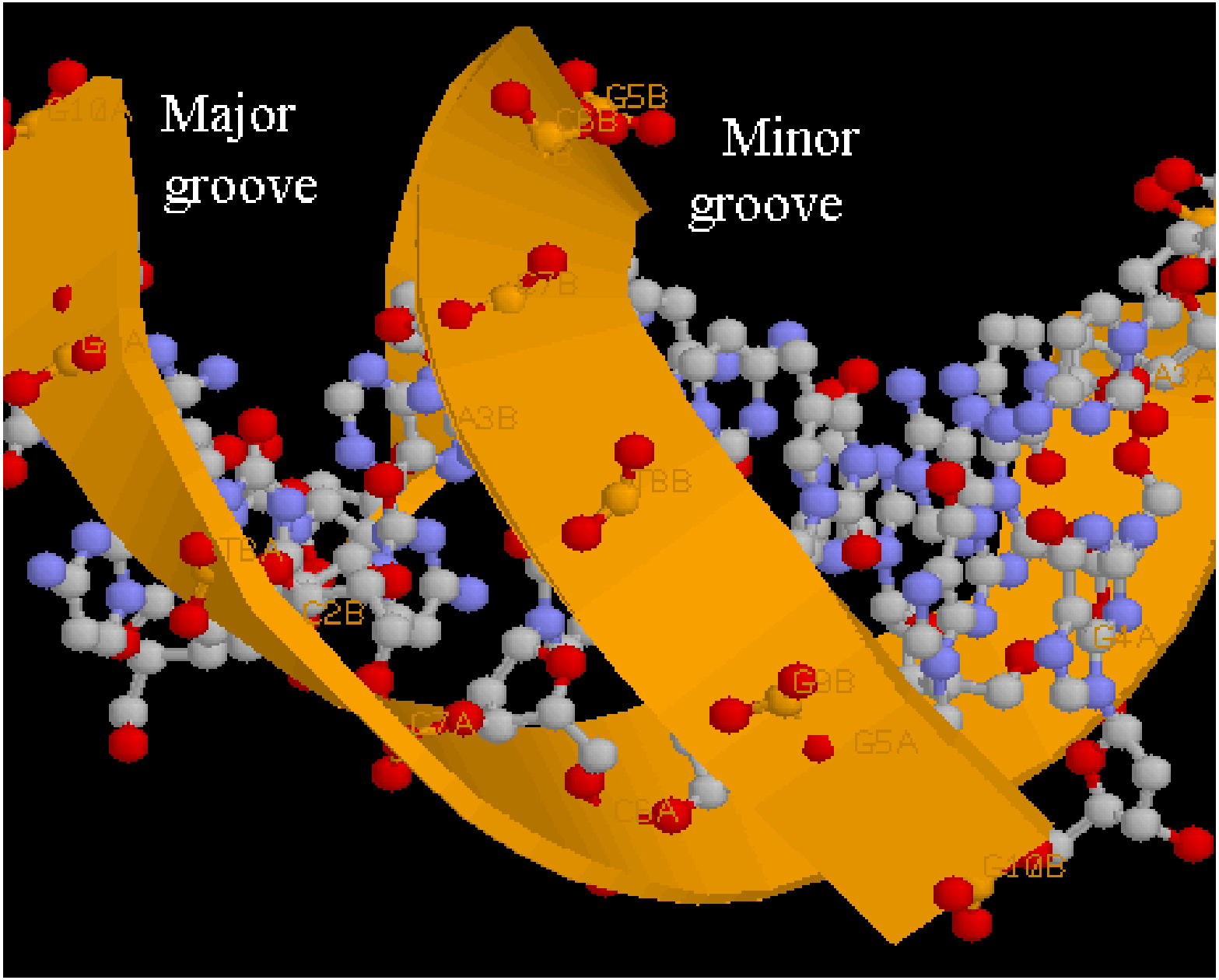
DNA bindende Proteine haben eine DNA-Bindedomäne und binden oft als Dimere



TATA-Box binding Protein

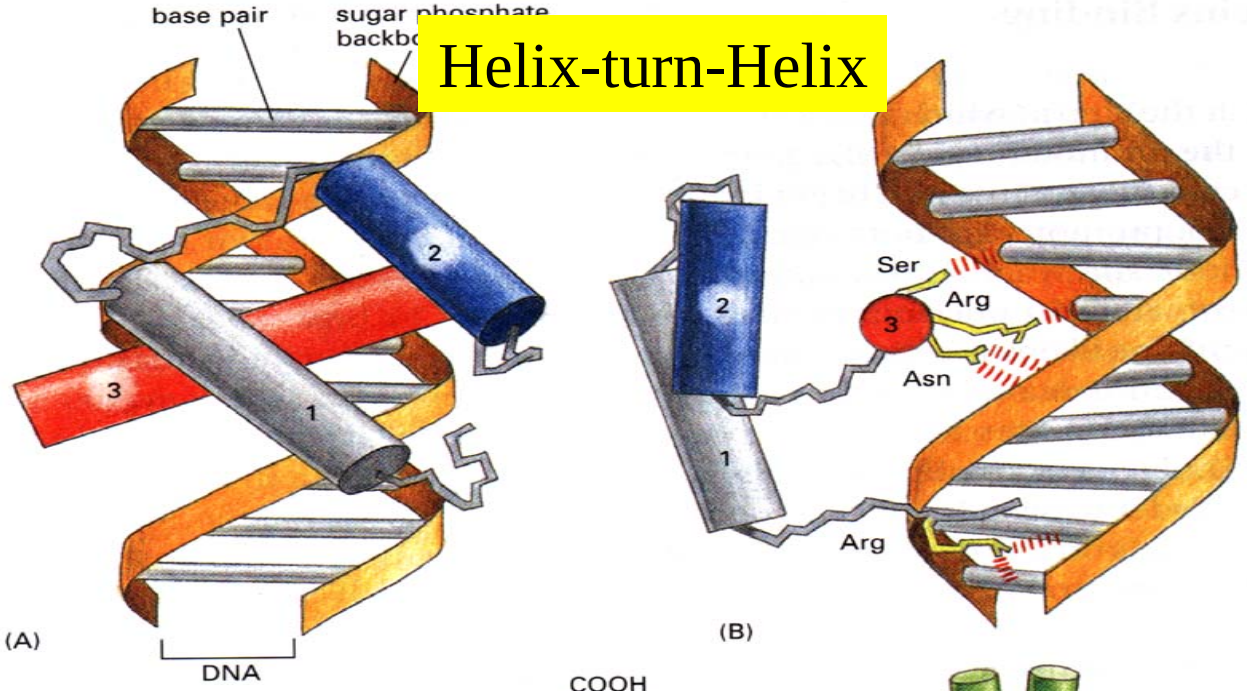


Bindung „große Grube“/kleine Grube

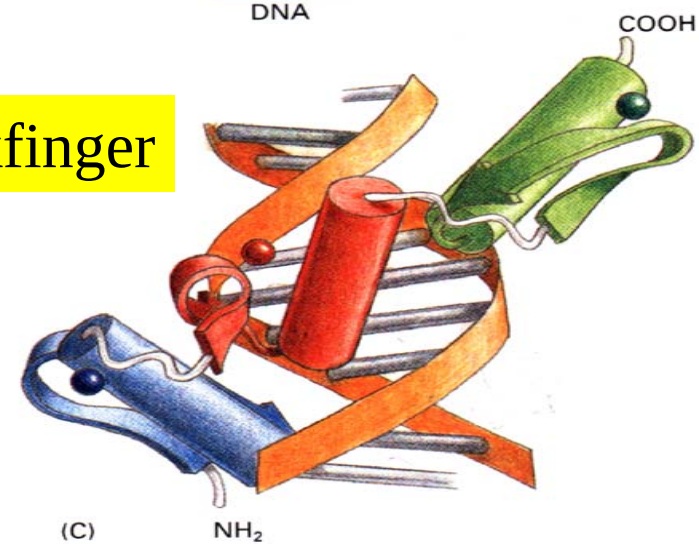


Die wichtigsten DNA-binde-Proteine

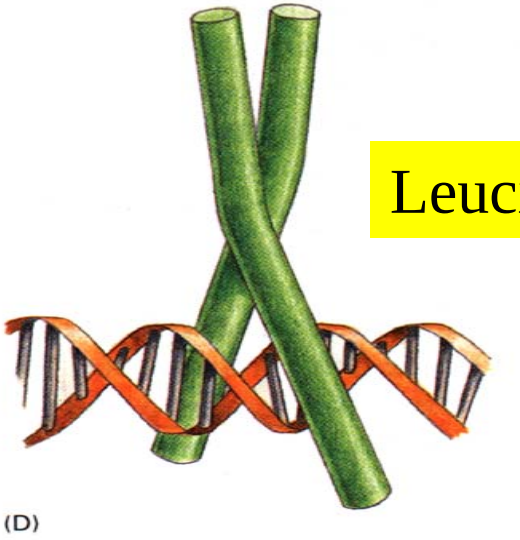
Helix-turn-Helix



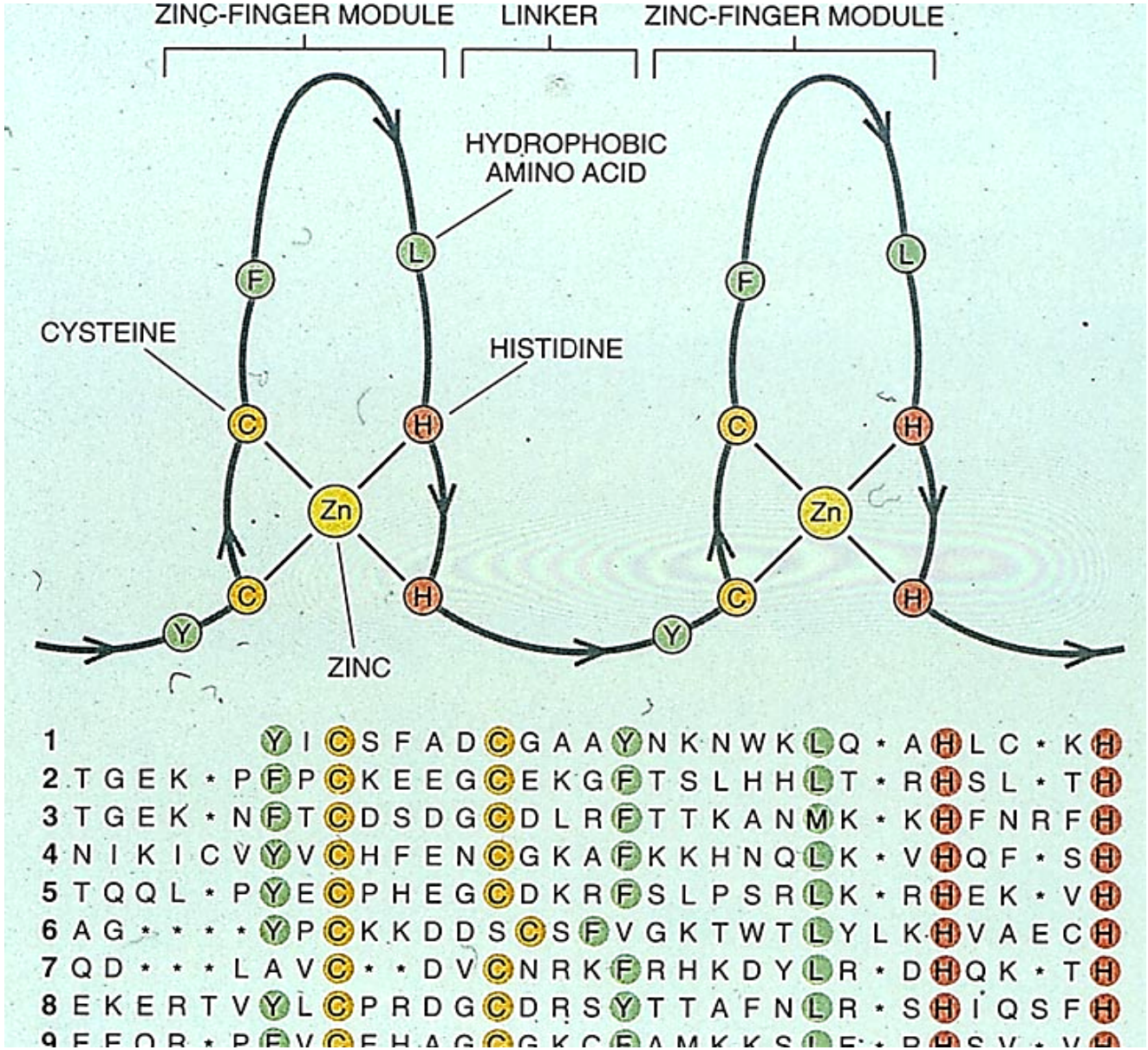
Zinkfinger



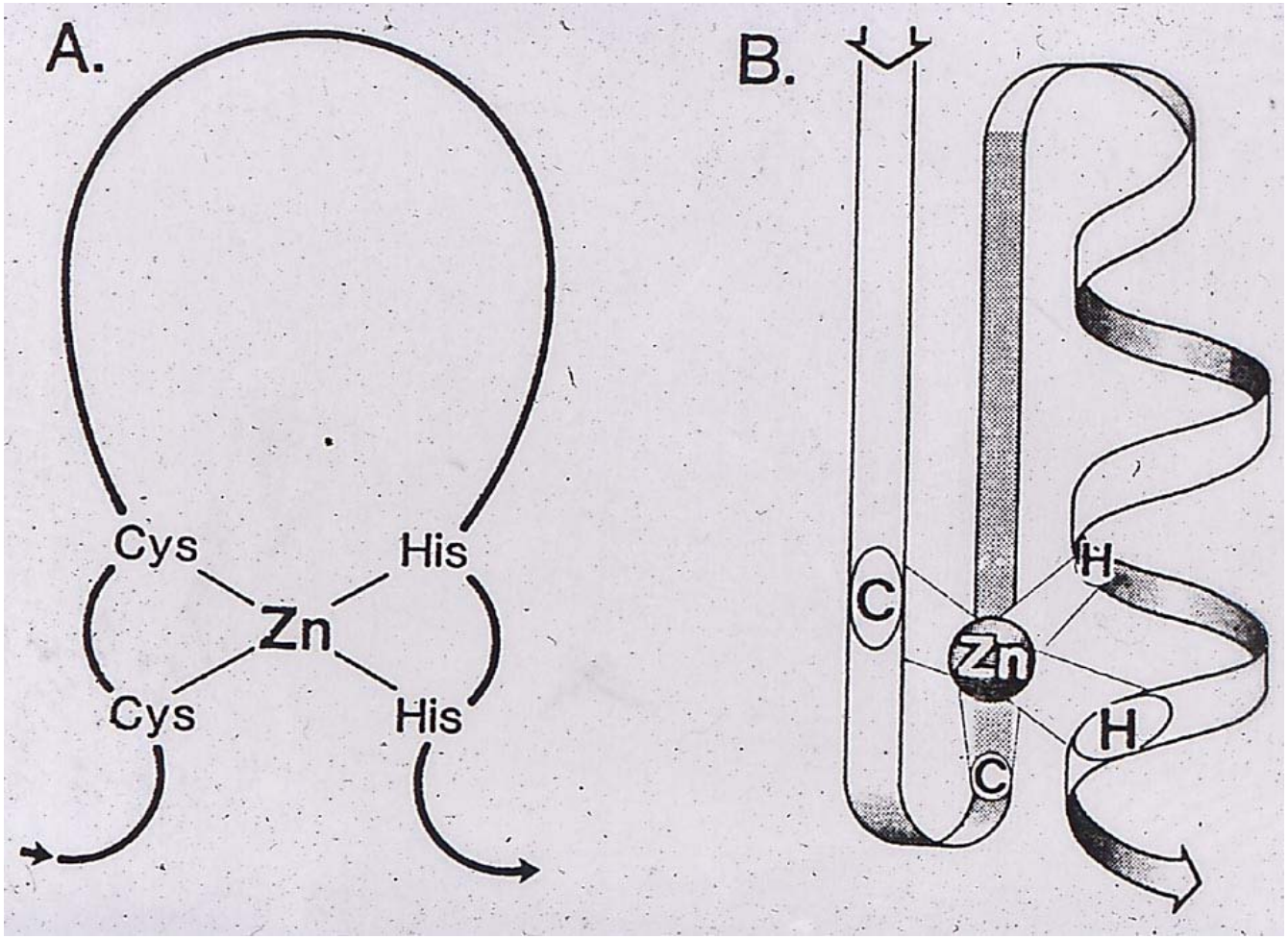
Leucin Zipper



ZinkfingerProteine (ZF)



Die wichtigsten DNA-binde-Proteine: Zinkfinger-Proteine



Die wichtigsten DNA-bindeProteine: Zinkfinger-Proteine: **Cys-His-Typ**

C_2H_2: F/Y-X- C $-X_{2-4}-$ C $-X_3$-F-X_5-L-X_2- H $-X_{3-4}-$ H $-X_{\beta}$

	Repeats	Binds DNA In vitro	Trans-Acting	Organism
TFIIIA[a]	9	+	+	Xenopus
ADR1[b]	2		+	yeast
SP1[c]	3	+	+	human
NGFI-A[d]	3			rat
Krüppel h[e]	2(+)			Drosophila
Krüppel[f]	4	+		Drosophila
Hunchback[g]	4+2			Drosophila
Serendipity β[h]	5			Drosophila
Serendipity δ[h]	6+1			Drosophila
Snail[i]	4			Drosophila
MKR1[j]	7(+)			mouse
MKR2[j]	9(+)			mouse
TDF[k]	13(+)			human
Xfin[l]	6+6+8+ 7+3+5			Xenopus

The two classes of finger proteins are listed with the general primary structure of each shown. Amino acids in bold are invariant and potentially coordinate metal, where "X" indicates intervening amino acid residues. A "+" between finger repeat units represents a linker of greater than 8 amino acids separating two groups of repeat units, "(+)" indicates data from a partial coding sequence, and "Trans-Acting" denotes demonstrated ability to transcriptionally regulate a gene(s).

Die wichtigsten DNA-binde-Proteine: Zinkfinger-Proteine **Cys-Cys-Typ**

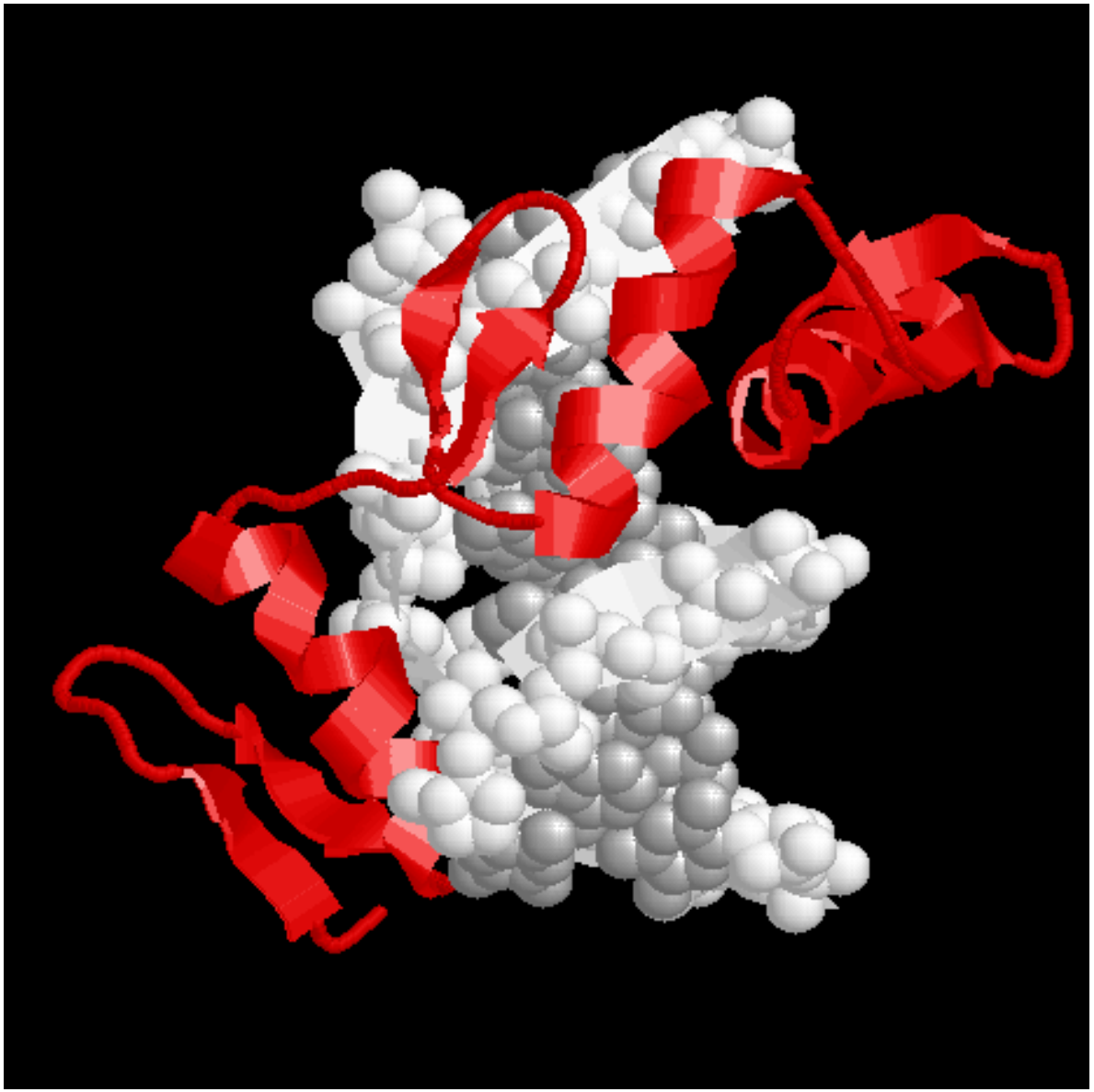
C_x C_4: C $-X_2-$ C $-X_{13}-$ C $-X_2-$ C

C_5: C $-X_5-$ C $-X_9-$ C $-X_2-$ C $-X_4-$ C

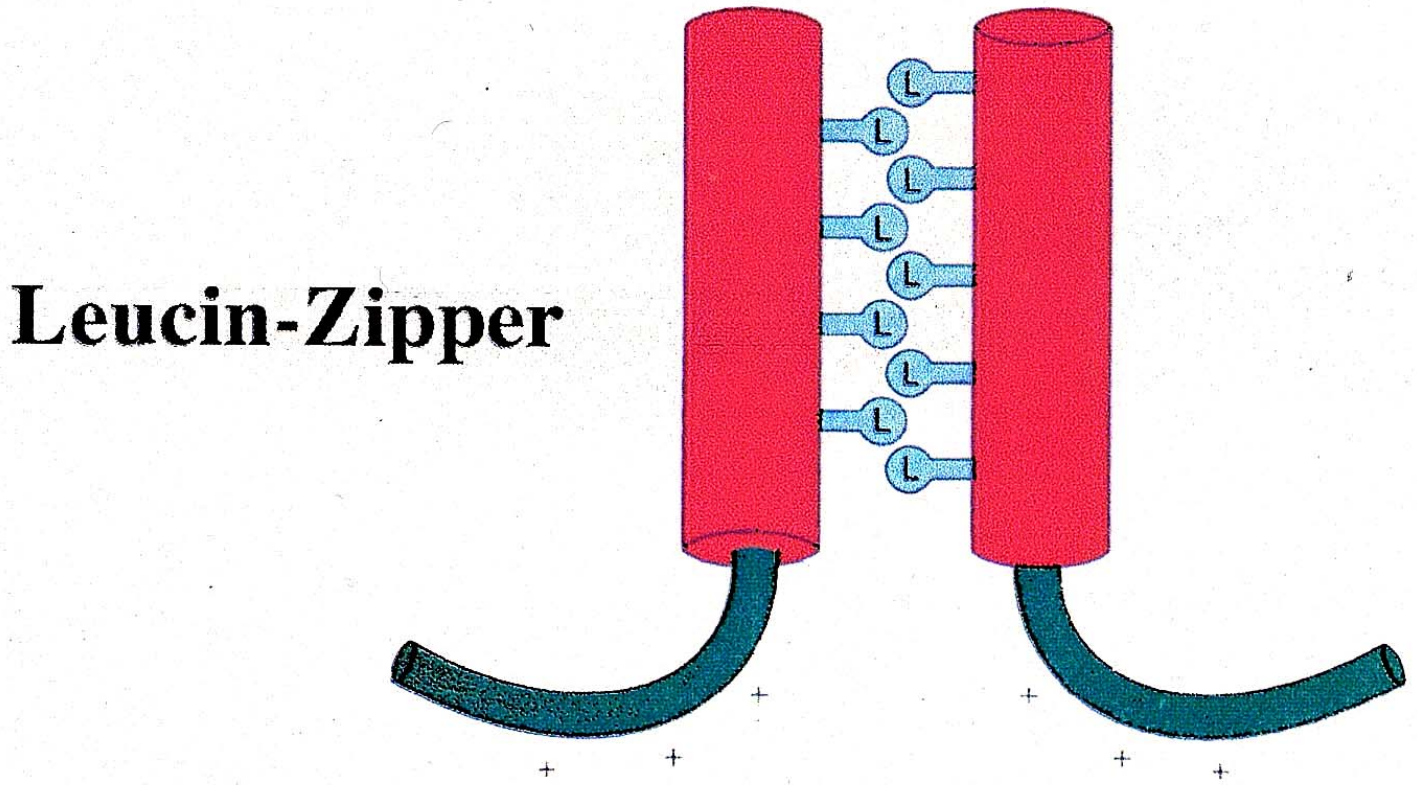
C_6: C $-X_2-$ C $-X_6-$ C $-X_6-$ C $-X_2-$ C $-X_6-$ C

	Finger Type	Binds DNA In vitro	Trans-Acting	Organism
GAL4[m] (PPR1/ARGR11/ LAC9/qa-1F)	C_6	+	+	yeast
E1A[n]	C_4	-	+	adenovirus
Steroid Hormone Receptor Superfamily[o]	C_4+C_5	+	+	human/rat/ mouse/ chicken

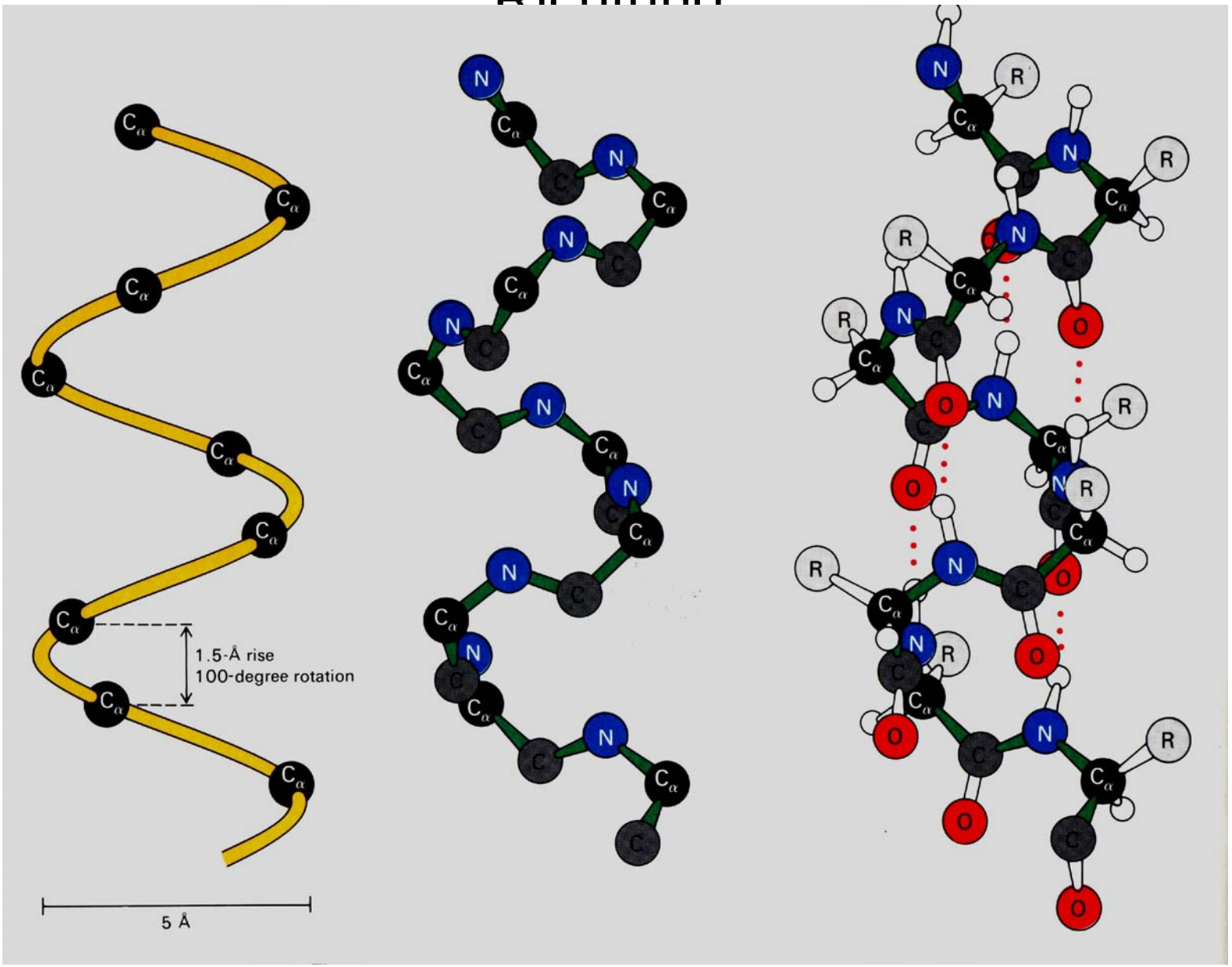
Zinkfinger-Motiv



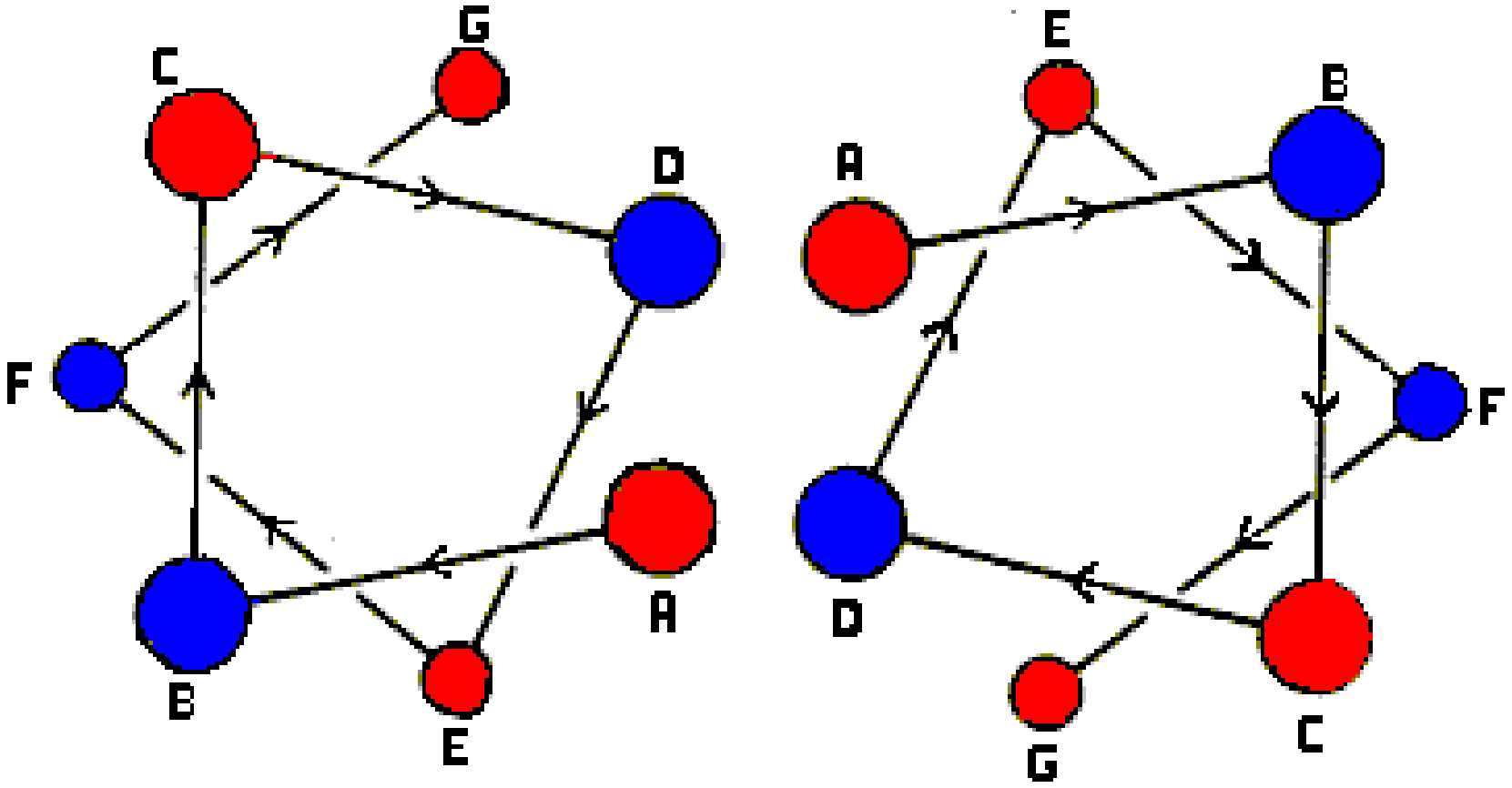
Leucin „Zipper“



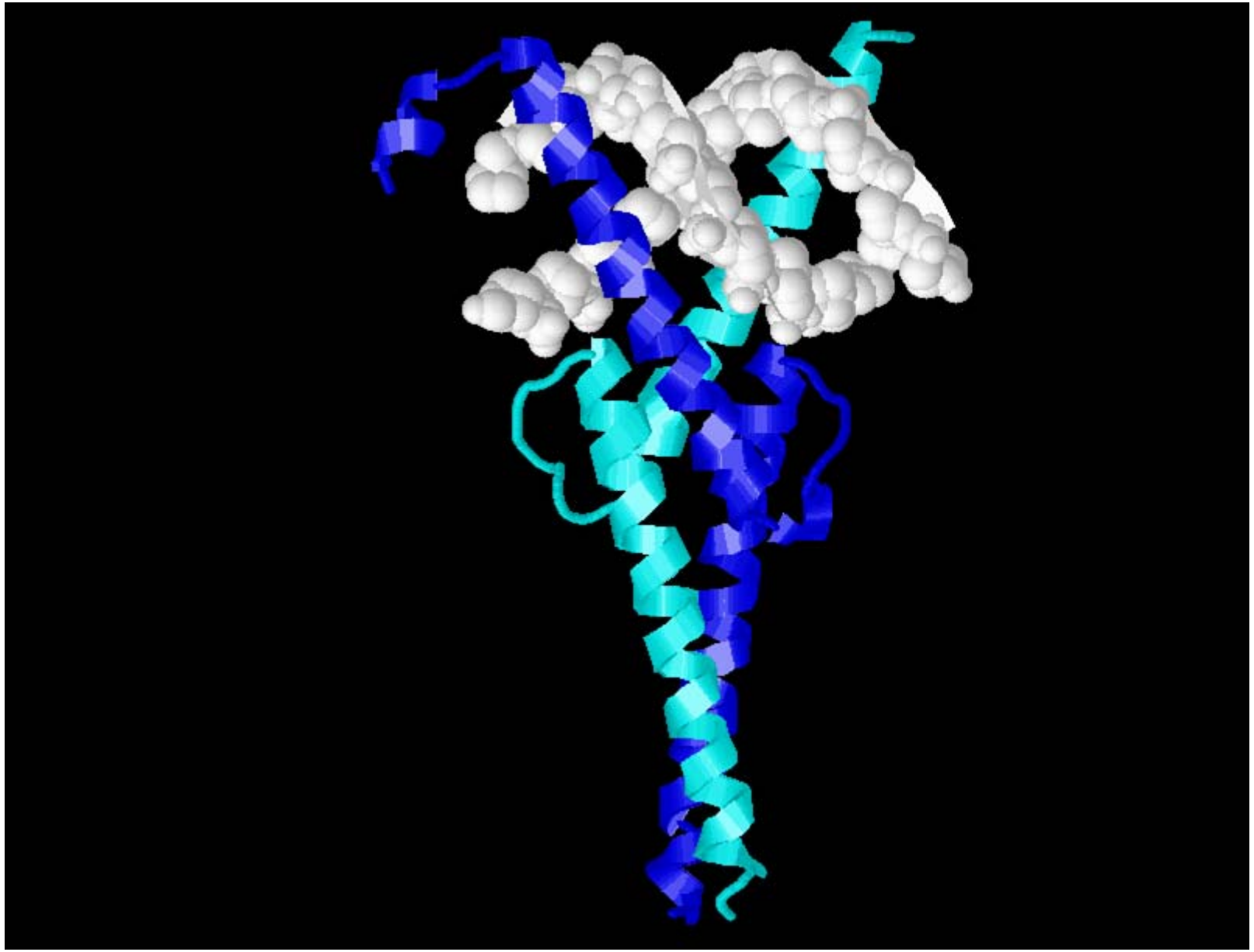
alpha-Helix der Proteine:
alle 7 Aminosäuren weist die Helix in die gleiche Richtung



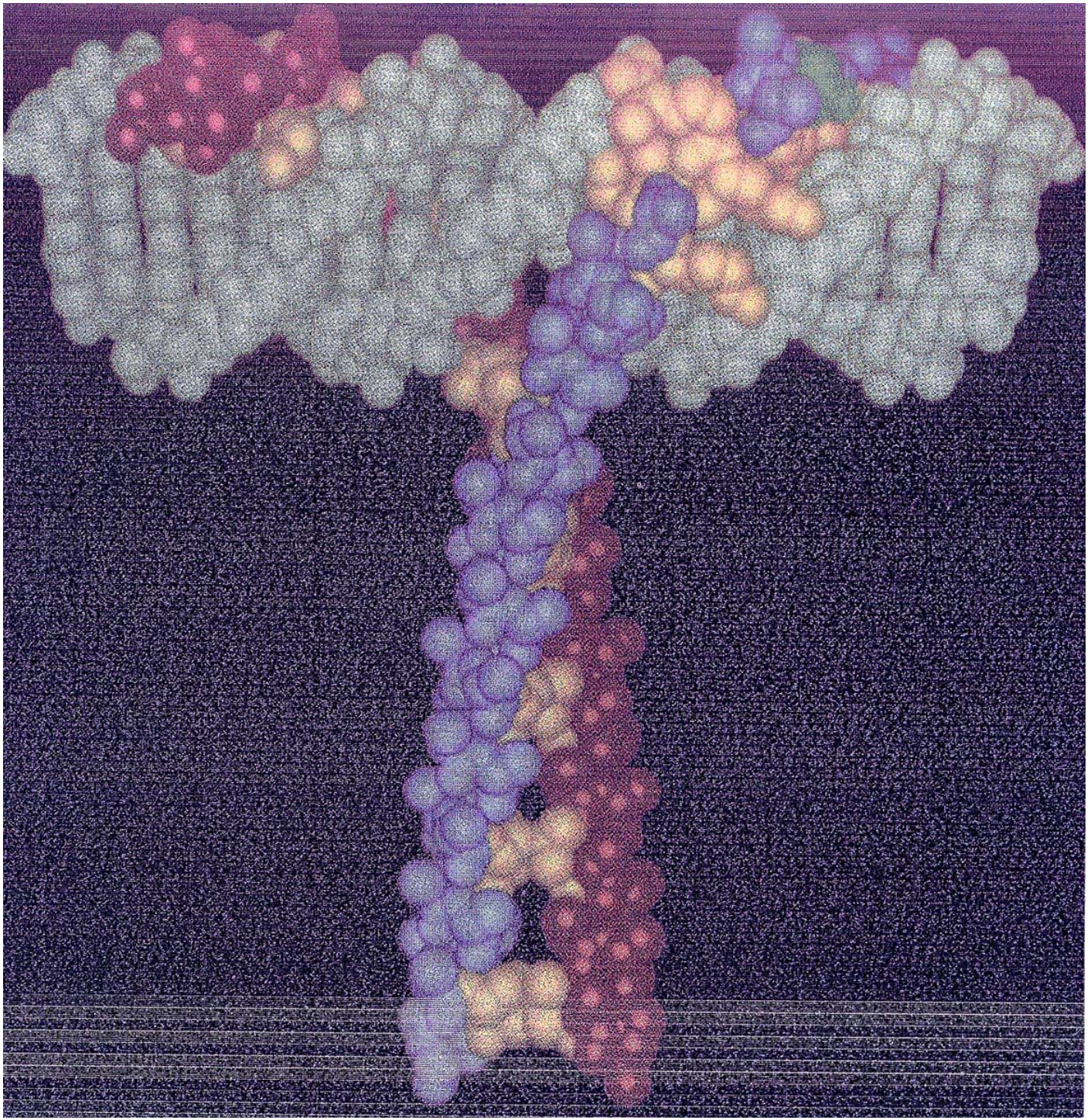
Alpha-Helix



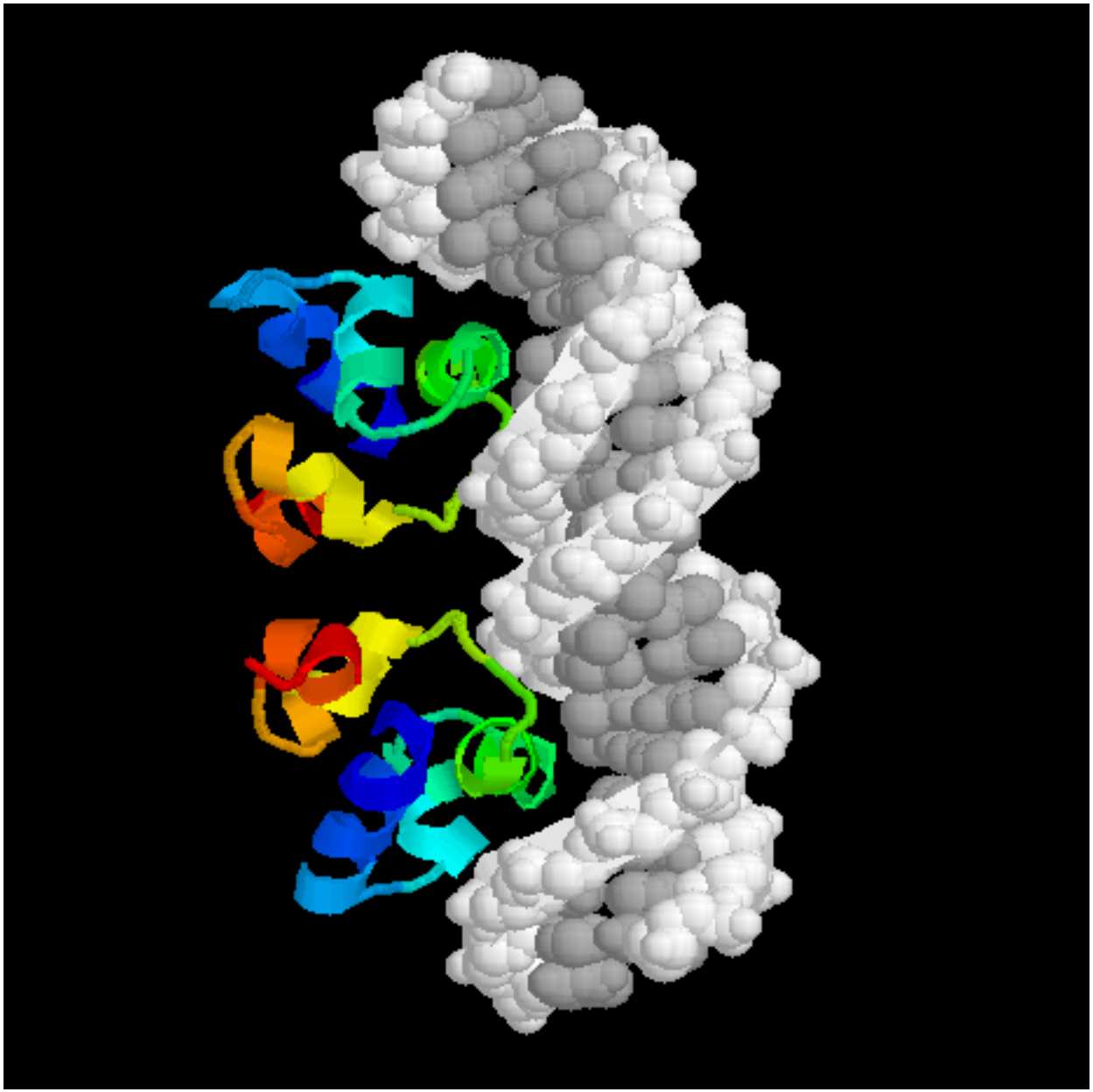
Leucin-Zipper



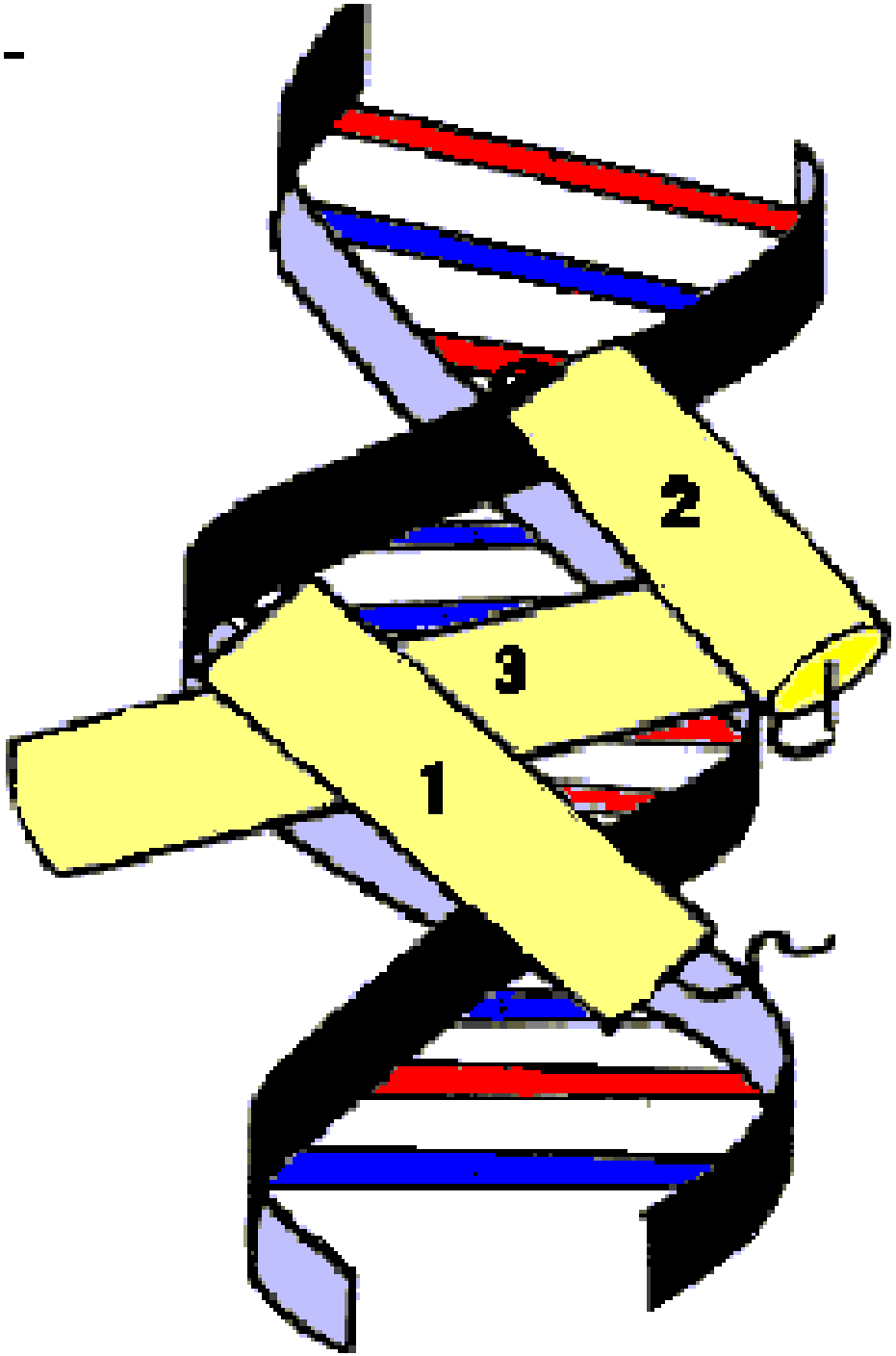
Leucin „Zipper“



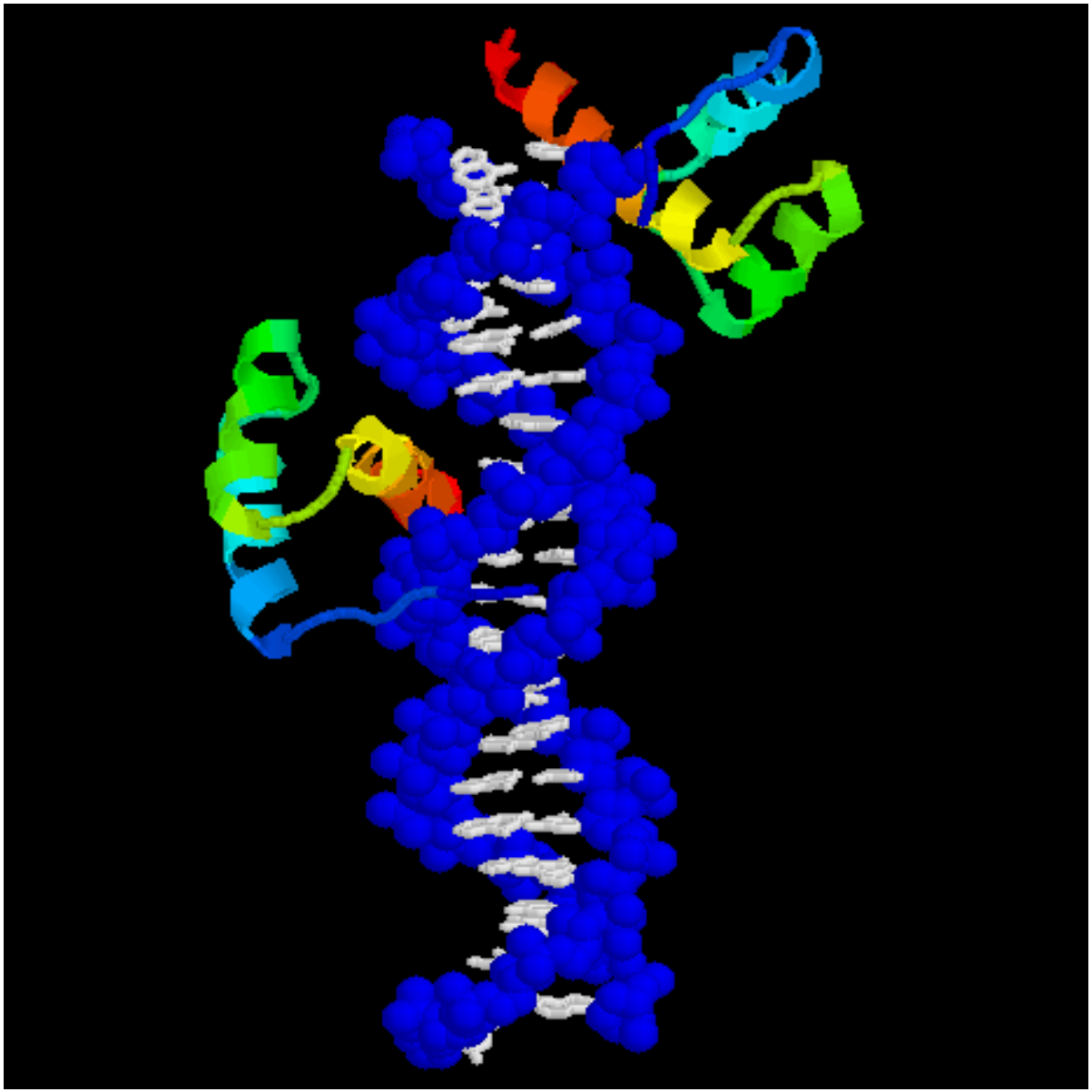
Helix-turn-Helix-Protein



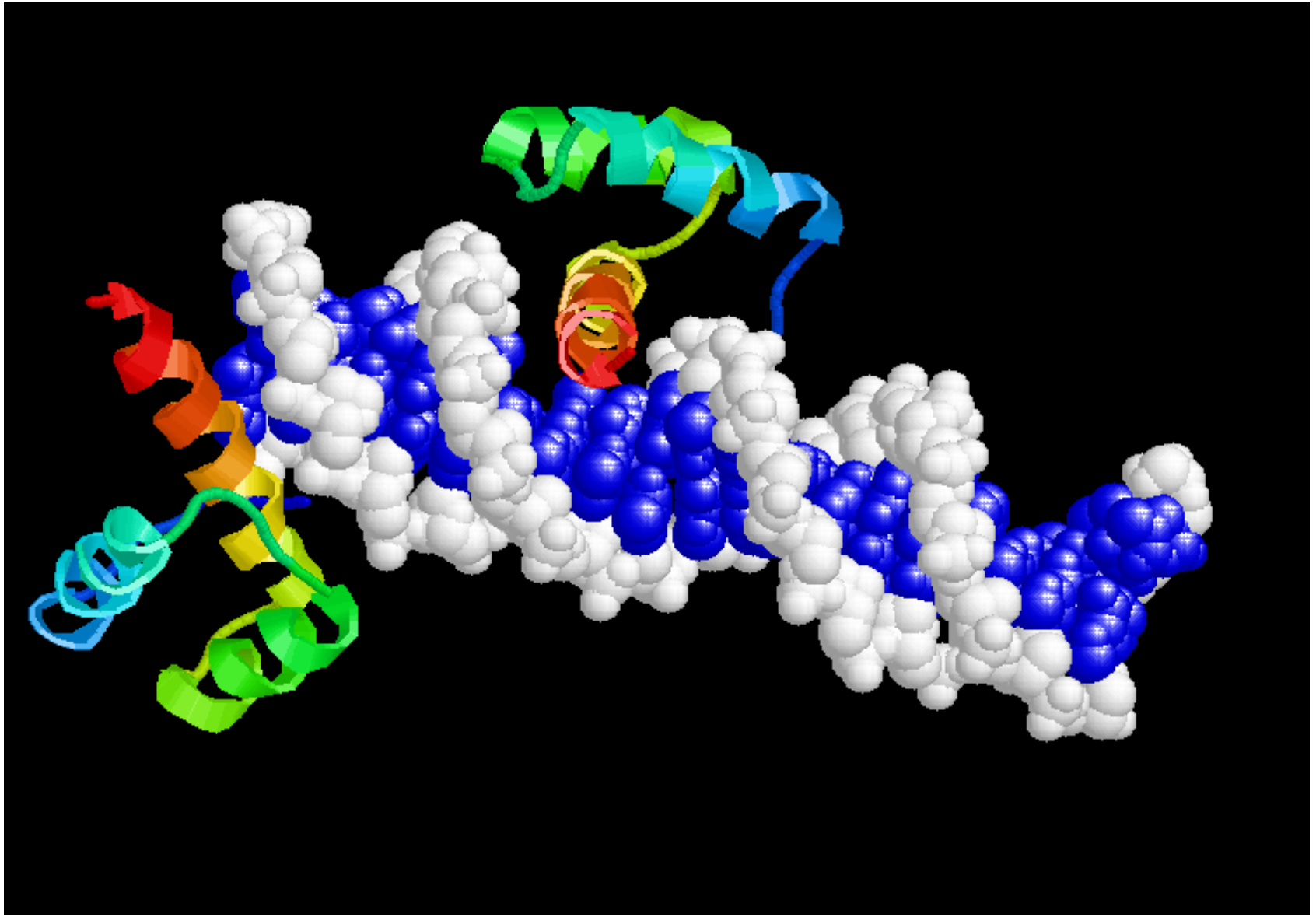
Helix-turn-Helix-Proteine



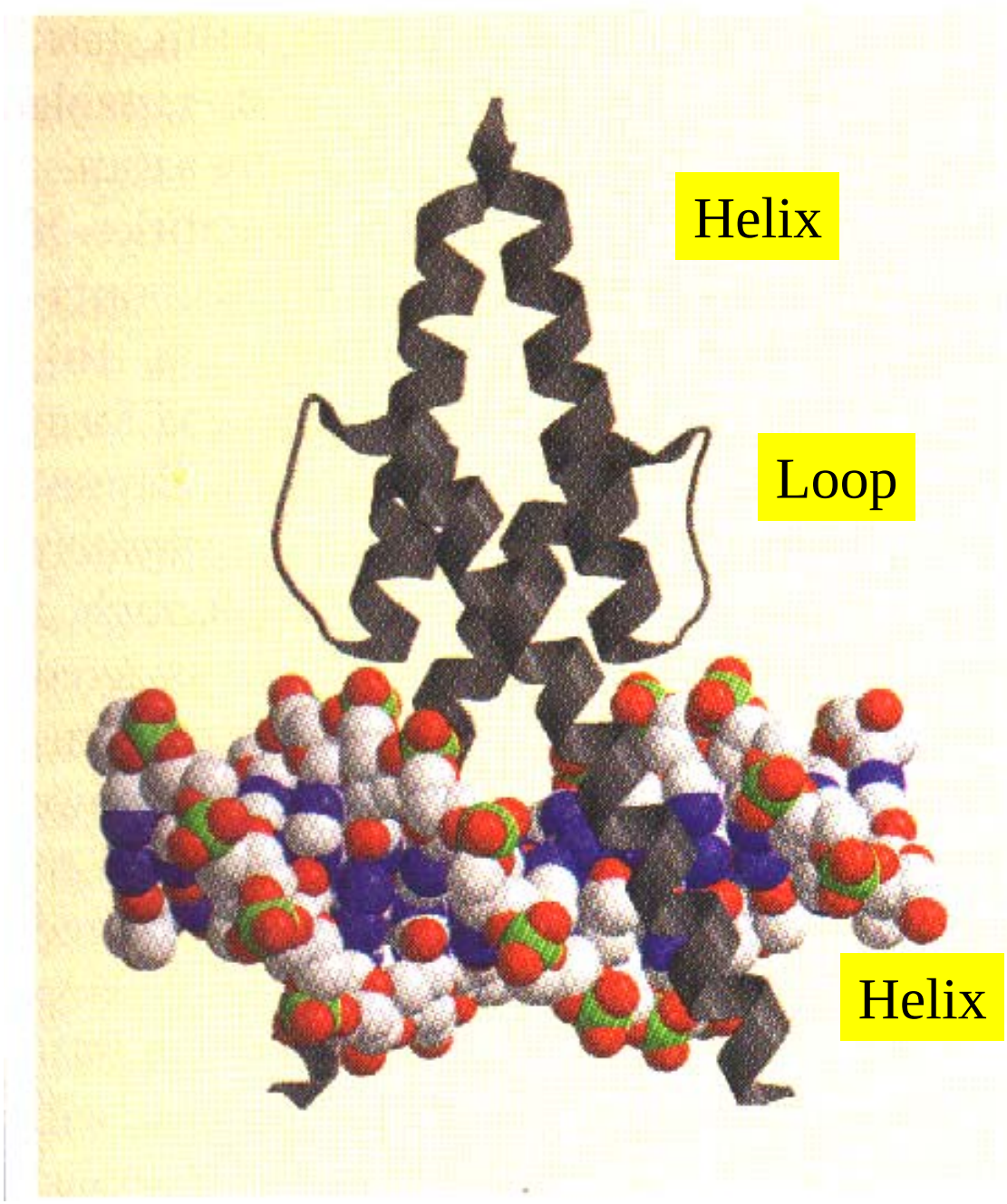
Homeodomän-Protein



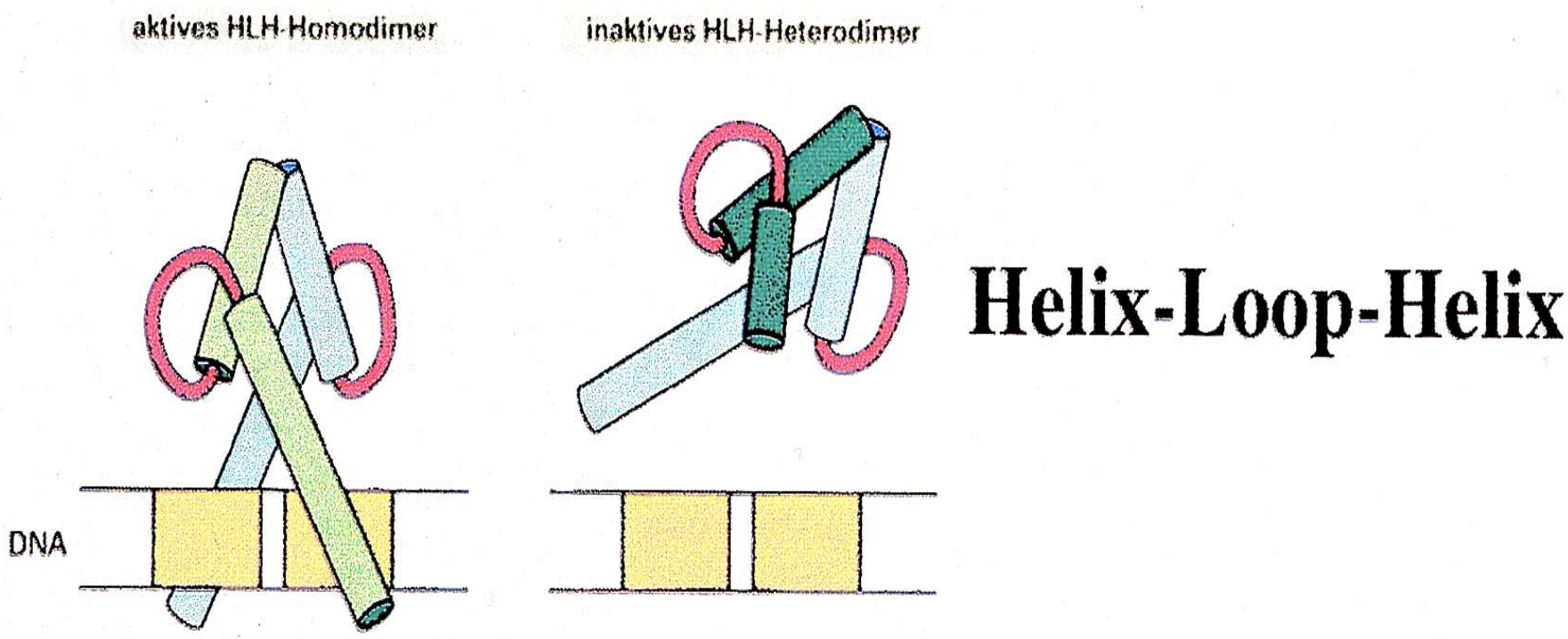
Drosophila TF „engrailed“



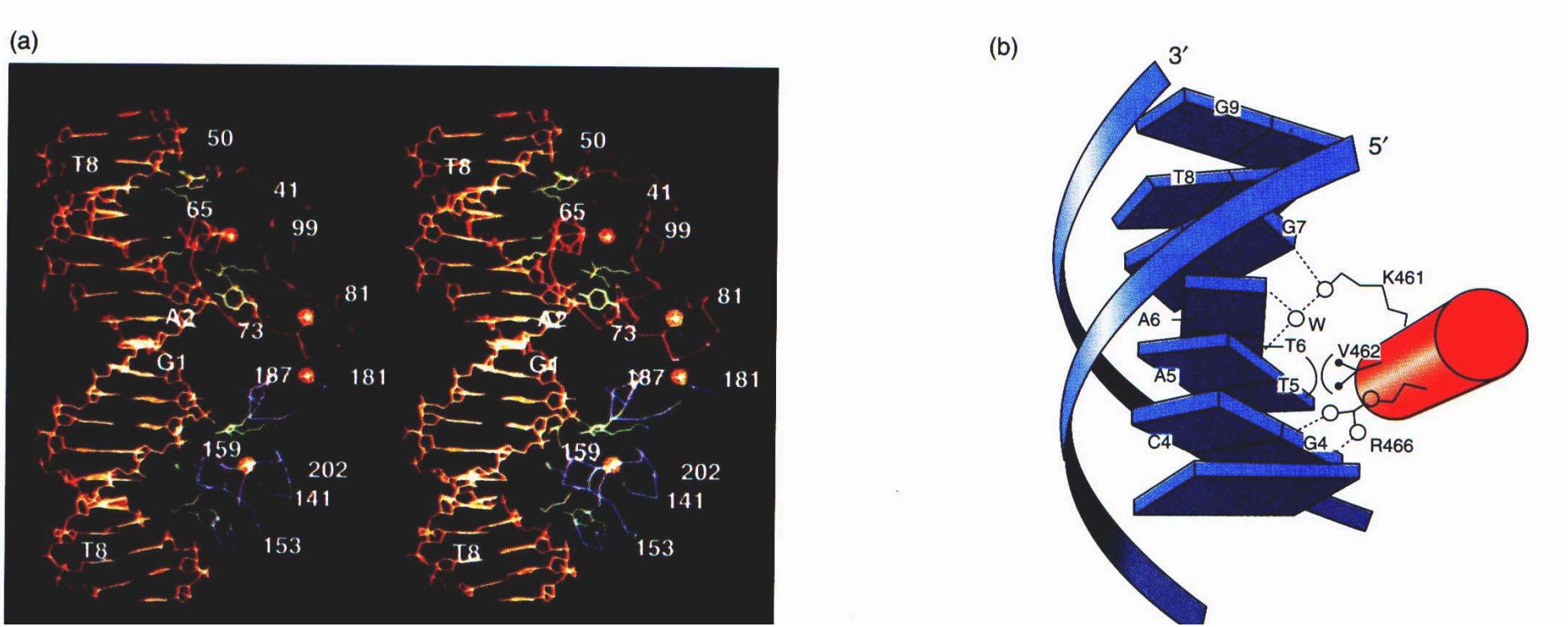
DNA-binde-Proteine: Basisches Helix-loop-Helix-Protein (bHLH)



Basisches Helix-Loop-Helix

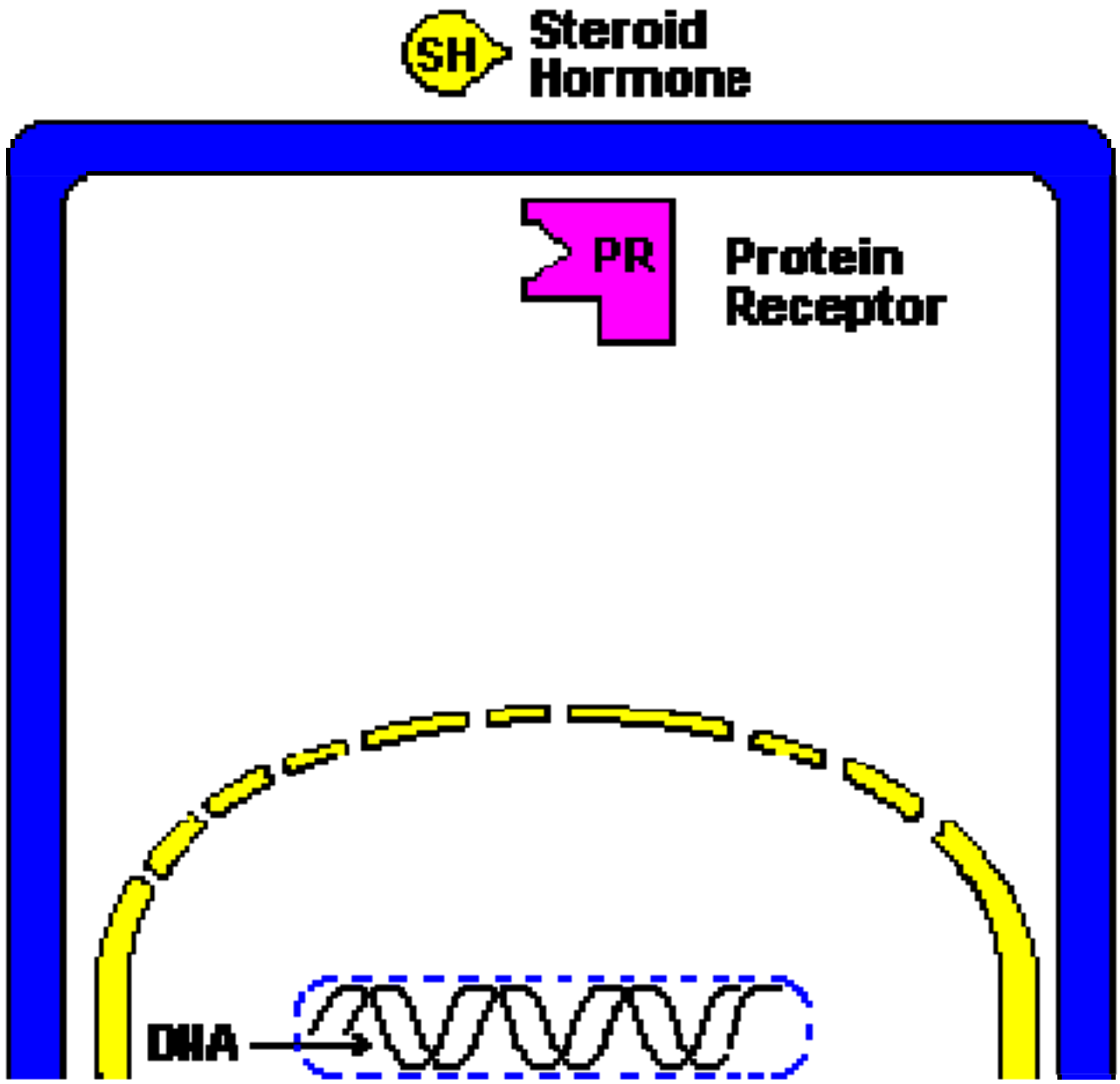


H-Brückenbindungen zwischen Aminosäuren des Proteins und Basen der DNA über die große Grube stellen die sequenzspezifische Bindung sicher



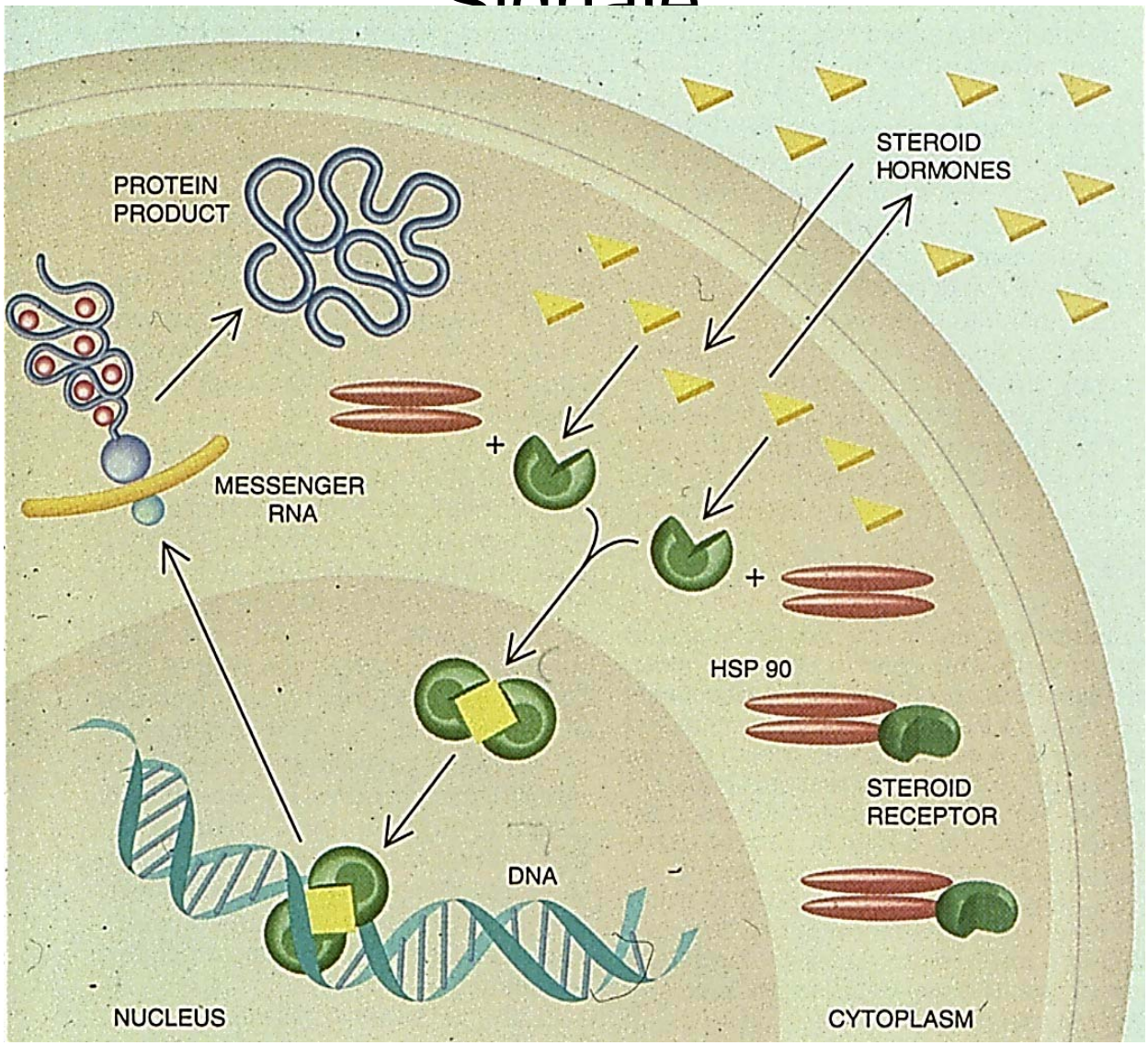
Die Aminosäuren der DNA-bindenProteine interagieren über die große Grube direkt mit den Basen der DNA über H-Brückenbindungen

Genregulation durch Steroidhormon



Hormon induzierte Genaktivität

Beispiel für Genaktivierung durch externe Signale



Hormon induzierte Genaktivität

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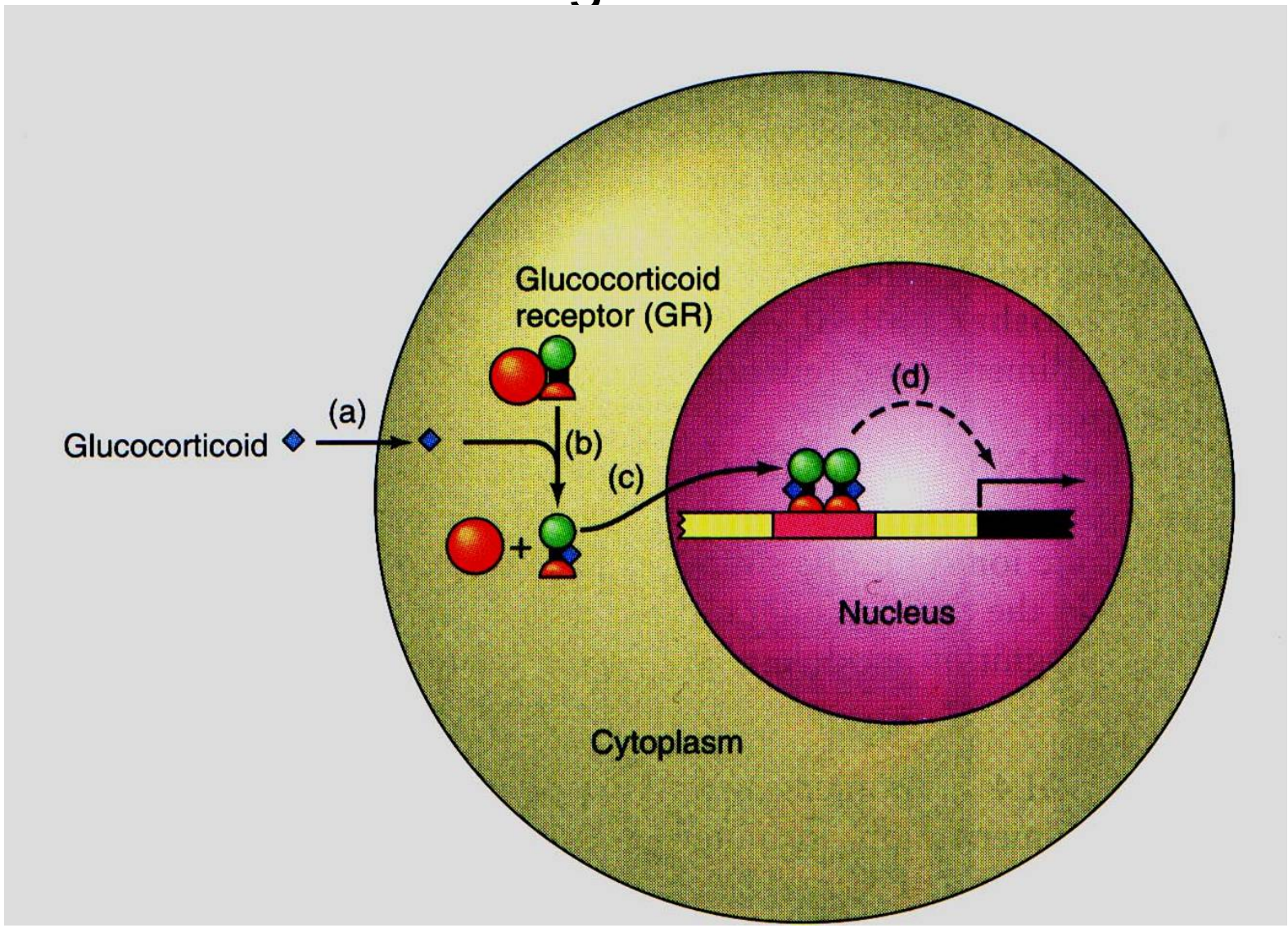


Nuclear Receptors

Palindromic Repeats

Glucocorticoid	RGRACANNNTGTYCY
Oestrogen	RGGTCANNNTGACCY
Thyroid	RGGTCA------TGACCY

Direct Repeats

6-cis retinoic acid	$AGGTCAN_1AGGTCA$
All trans retinoic acid	$AGGTCAN_2AGGTCA$
Thyroid hormone	$AGGTCAN_4AGGTCA$

N indicates any nucleotide
R indicates a purine ie. A or G
Y indicates a pyrimidine ie. C or T

Nuclear Receptors

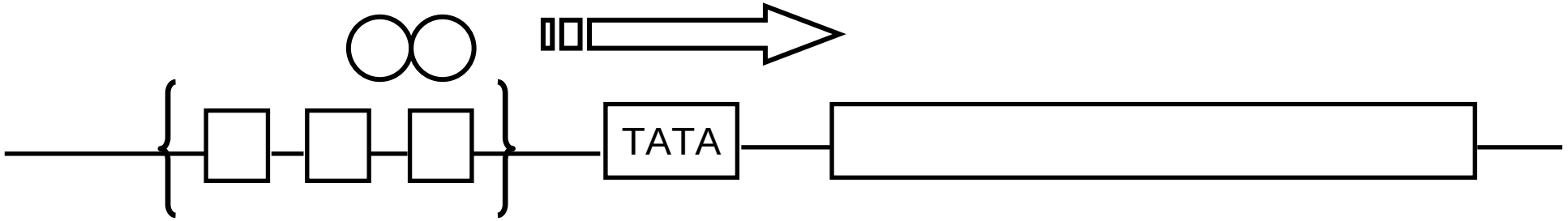
Binding of hormone

Dissociation from hsp90

Dimerisation

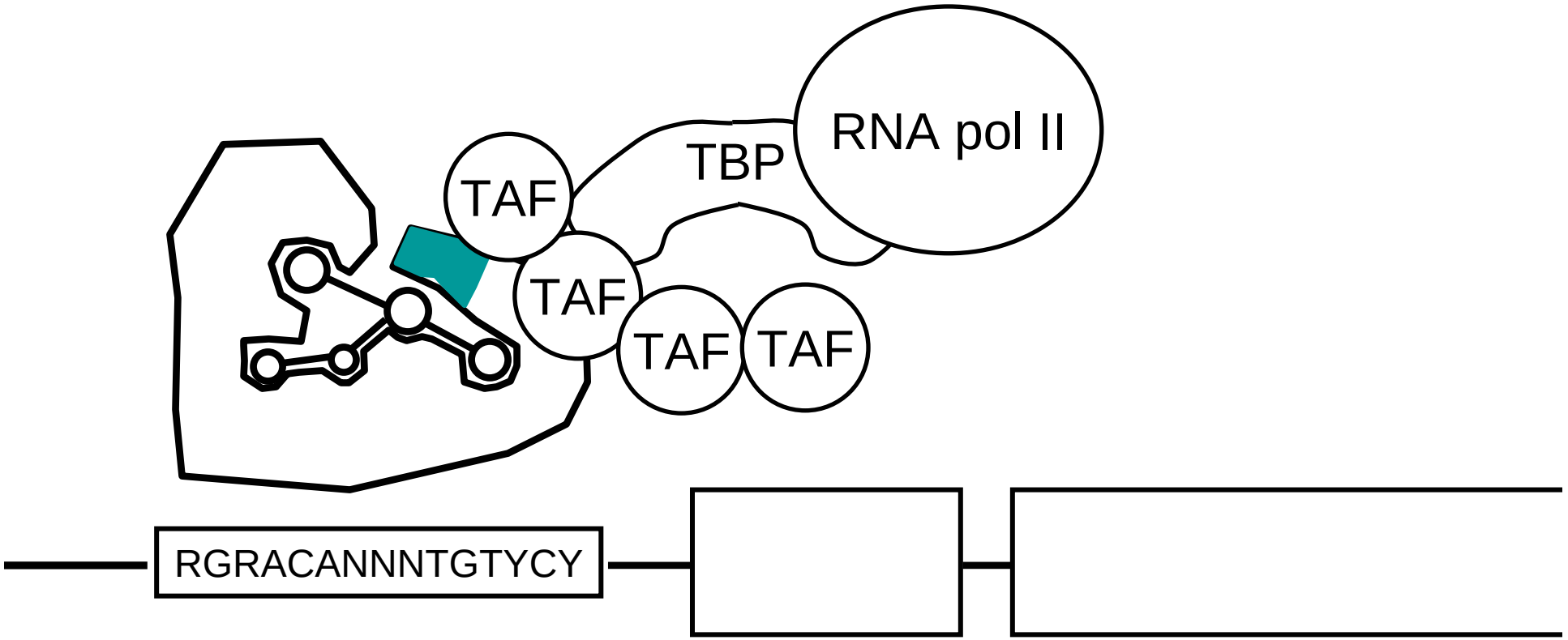
Migration to nucleus

Binding to URE

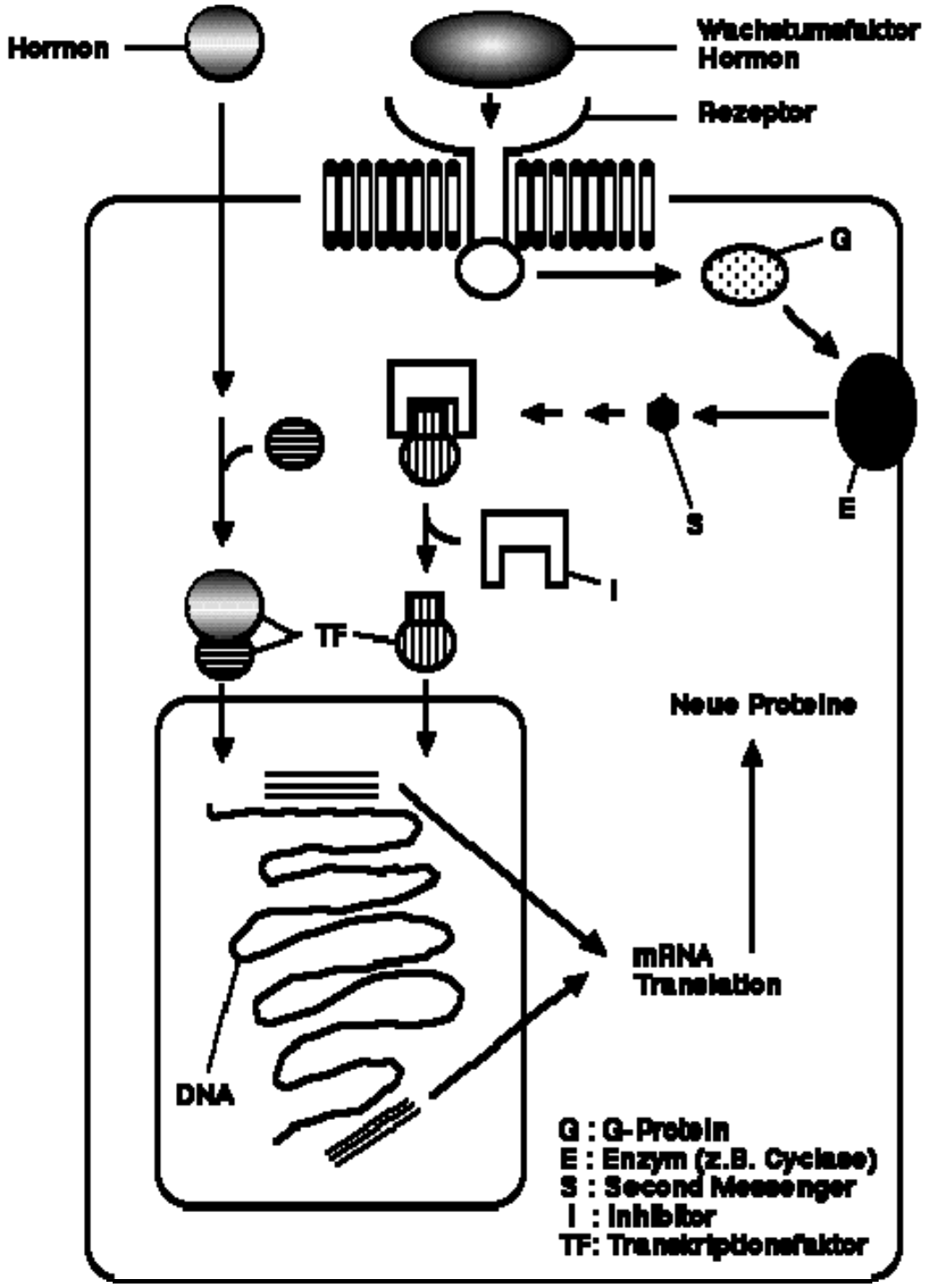


Nuclear Receptors

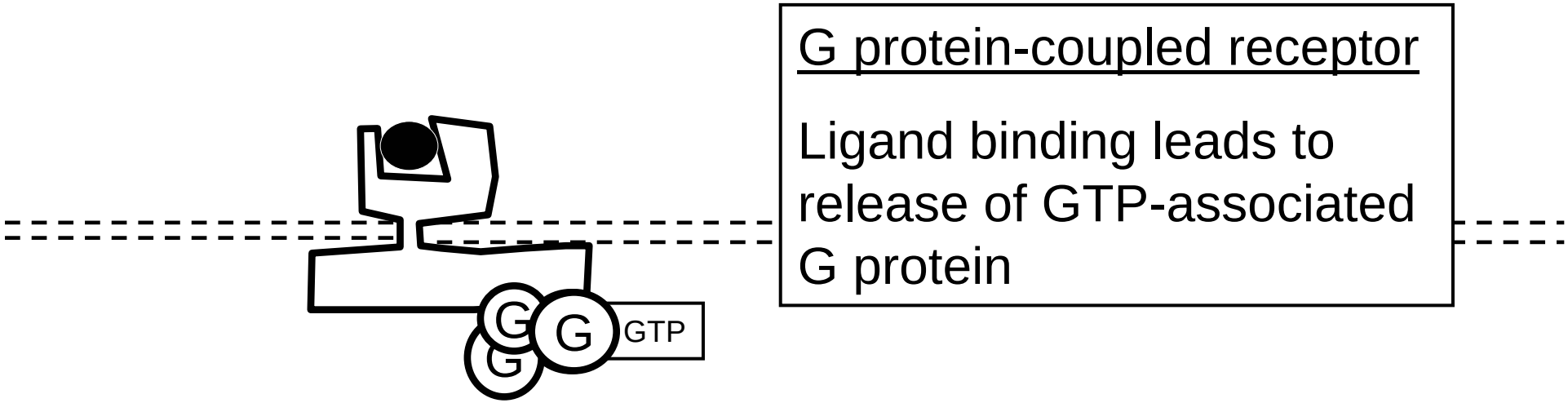
Binding of ligand causes conformational change allowing transactivation domain to interact with transcriptional machinery



Hormon-regulation von Genen



cAMP signalling

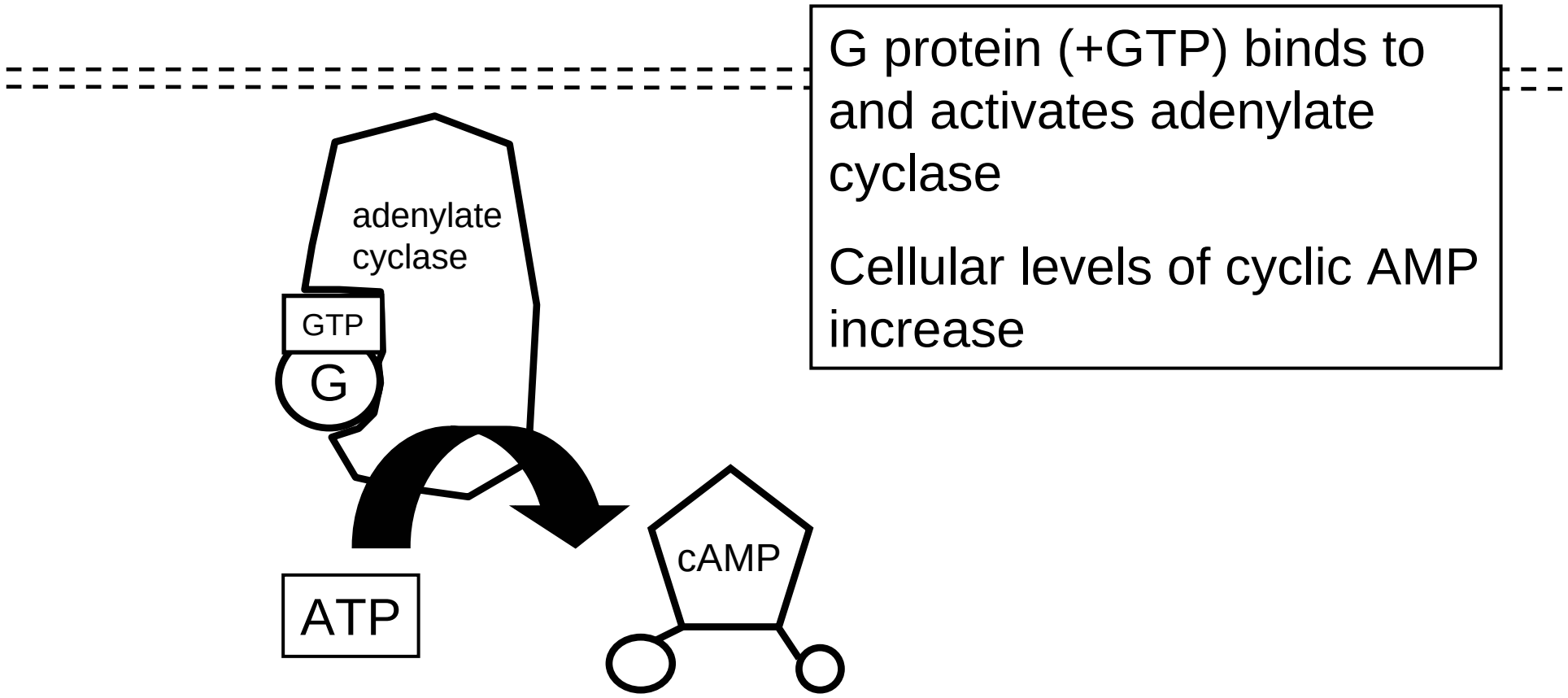


Small GTP-binding proteins

Example: ras – covered in earlier lecture

Alternate between inactive GDP and active GTP bound forms

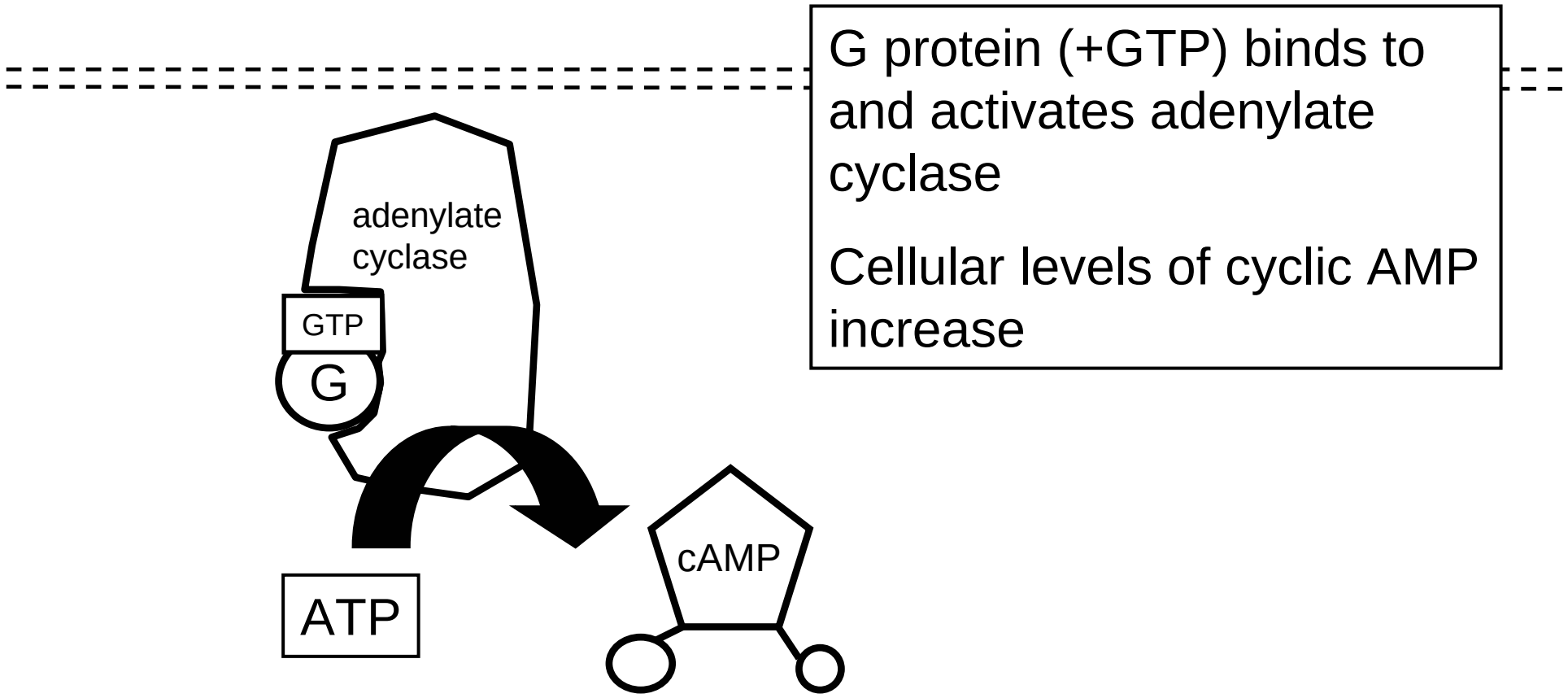
cAMP signalling



G protein (+GTP) binds to and activates adenylate cyclase

Cellular levels of cyclic AMP increase

cAMP signalling



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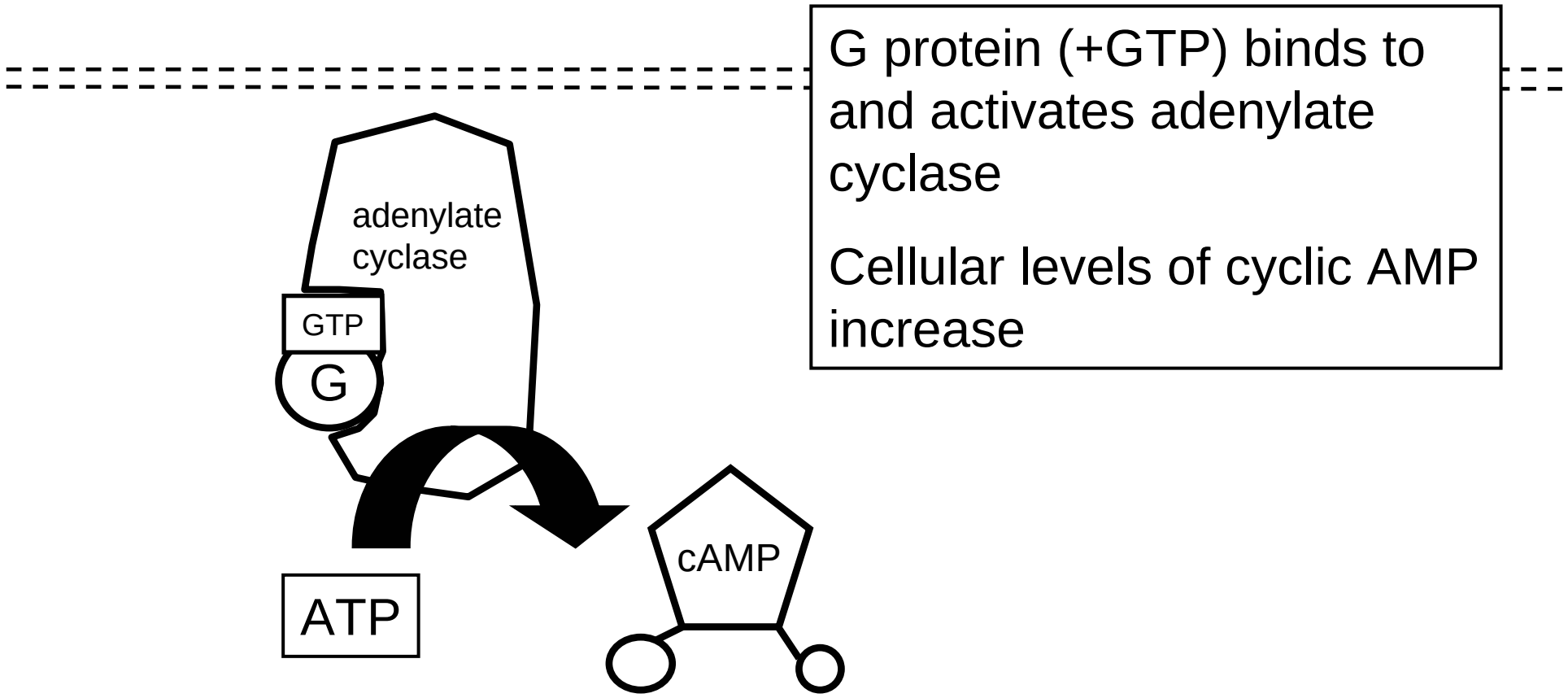

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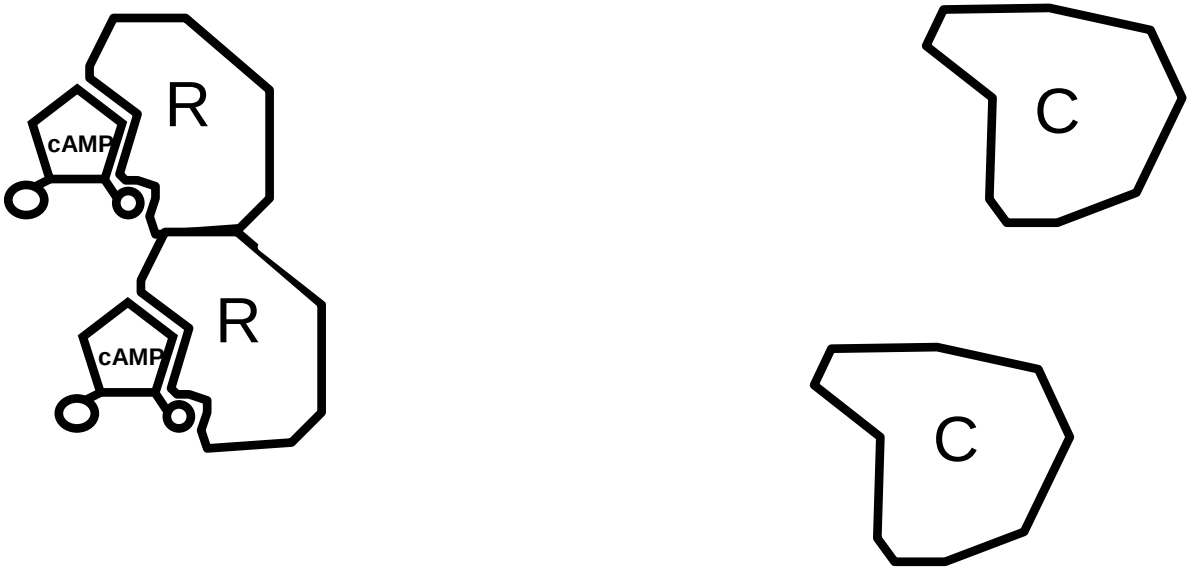
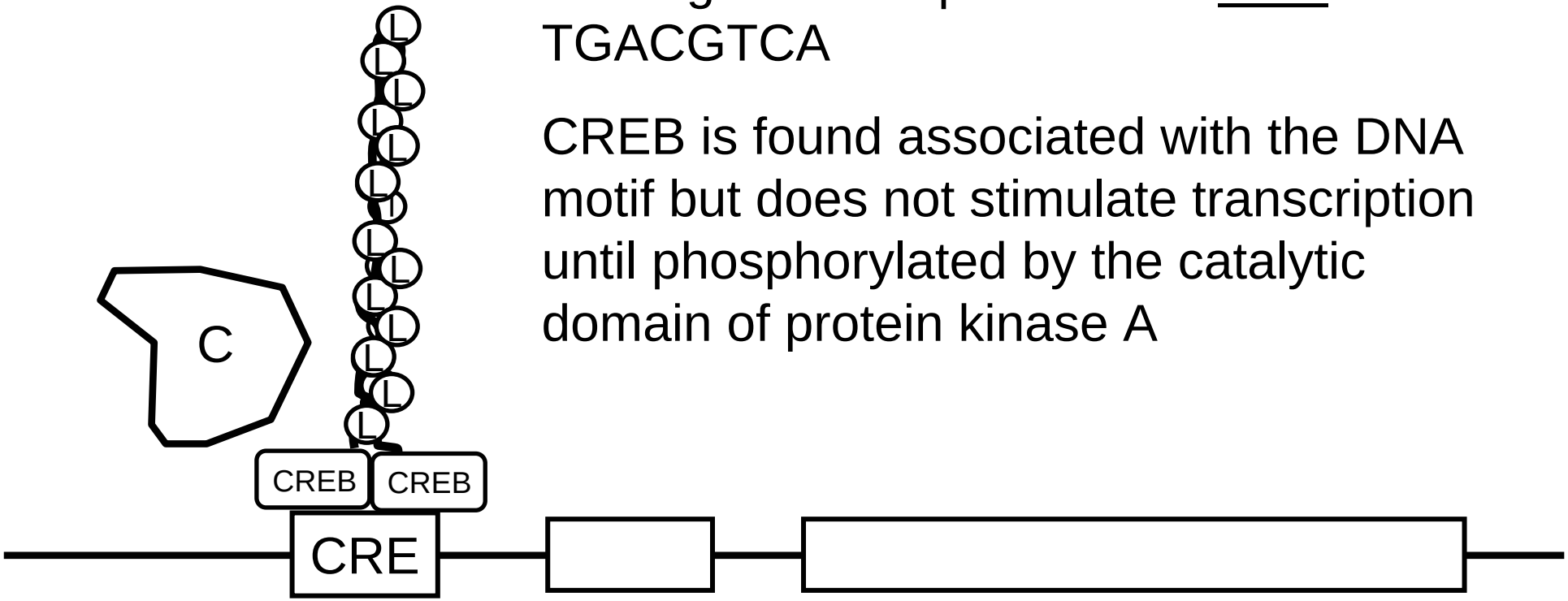


Cyclic AMP Response Element Binding protein

CREB is a member of the bZIP family of transcription factors

It recognizes the palindromic CRE motif TGACGTCA

CREB is found associated with the DNA motif but does not stimulate transcription until phosphorylated by the catalytic domain of protein kinase A

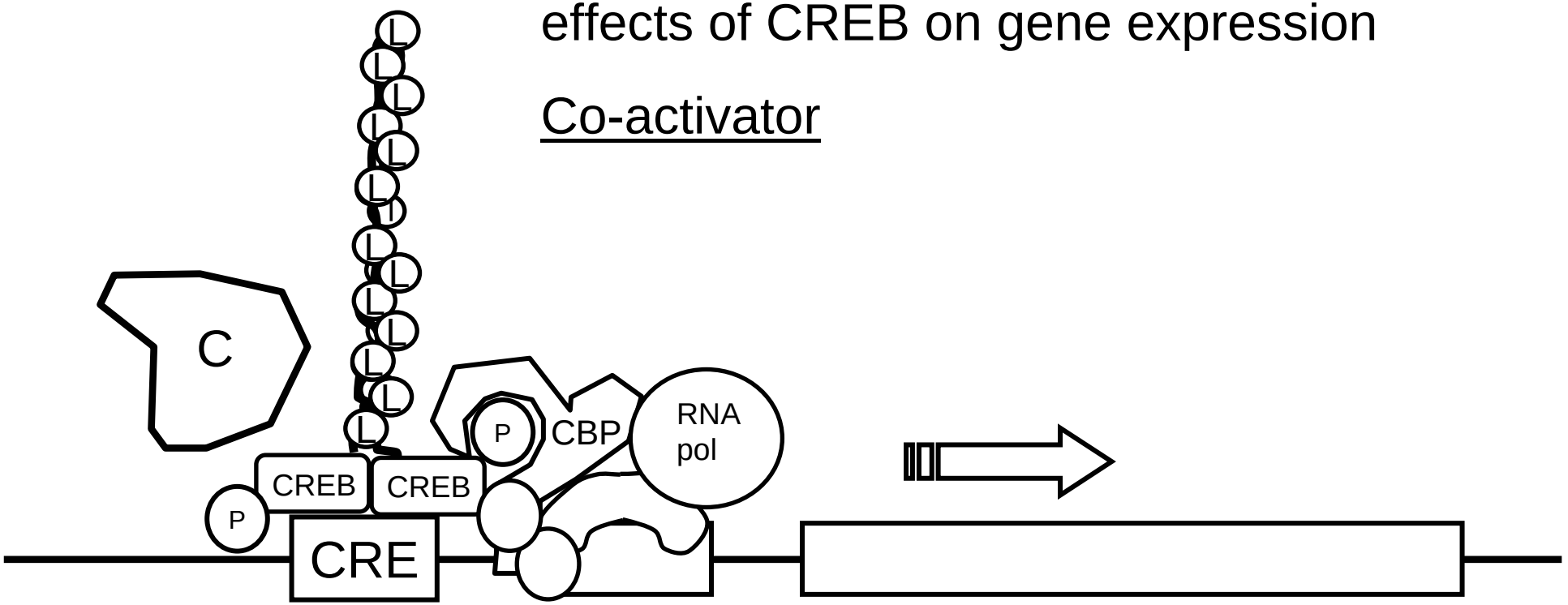


CREB binding protein

CBP recognises and binds the phosphorylated form of CREB

CBP interacts with the basal transcription complex to mediate the effects of CREB on gene expression

Co-activator

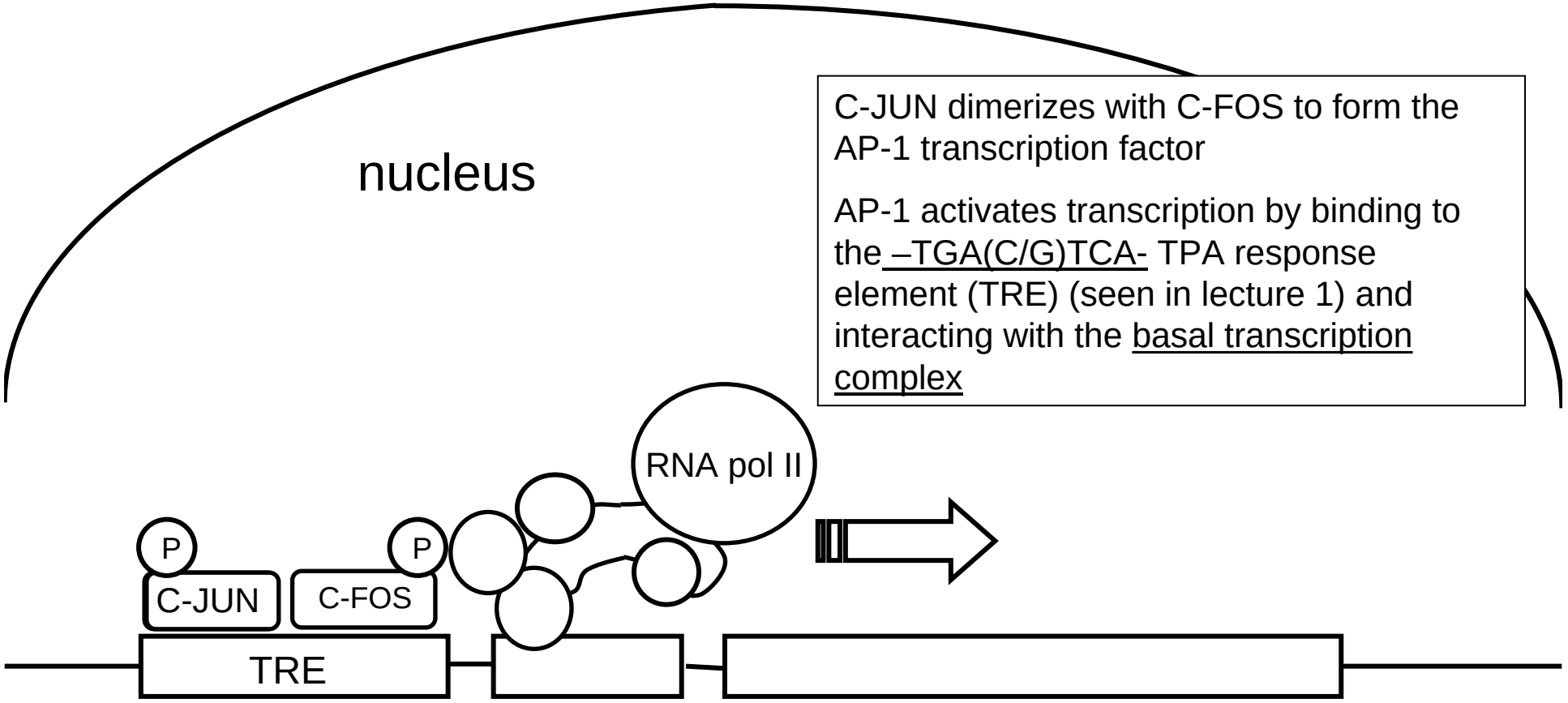


Kinase Cascades

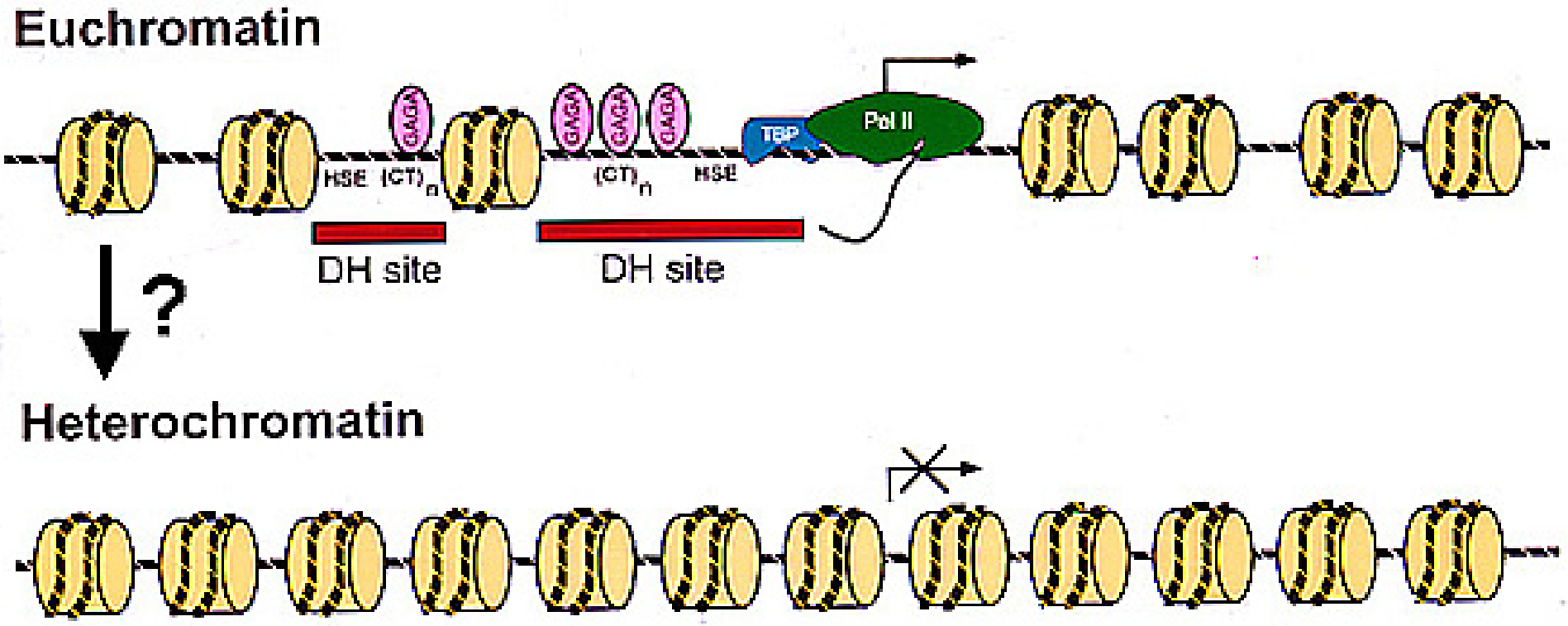
Active JNK travels to the nucleus and phosphorylates the bZIP transcription factor C-JUN

C-JUN dimerizes with C-FOS to form the AP-1 transcription factor

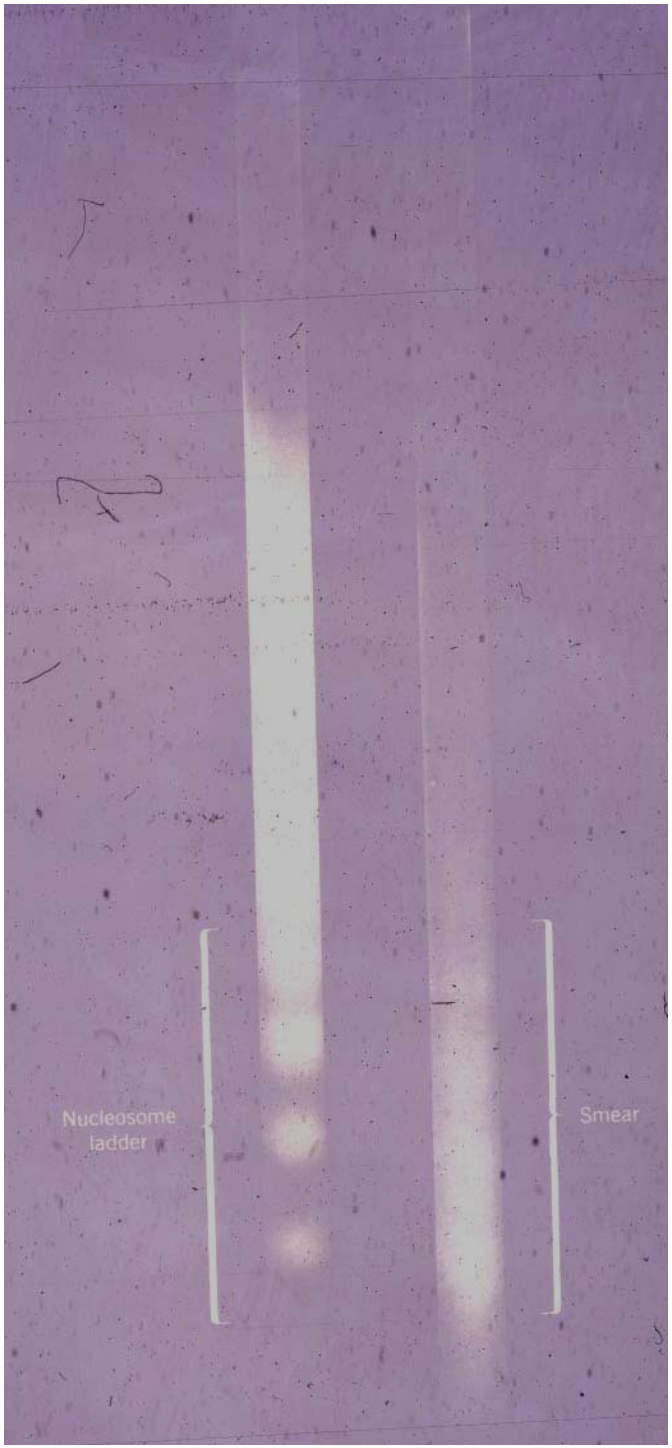
AP-1 activates transcription by binding to the –TGA(C/G)TCA- TPA response element (TRE) (seen in lecture 1) and interacting with the basal transcription complex



Chromatin und Genaktivität



Aktives Chromatin ist nicht in Nukleosomen organsisiert



Inaktive / Aktives Chromatin