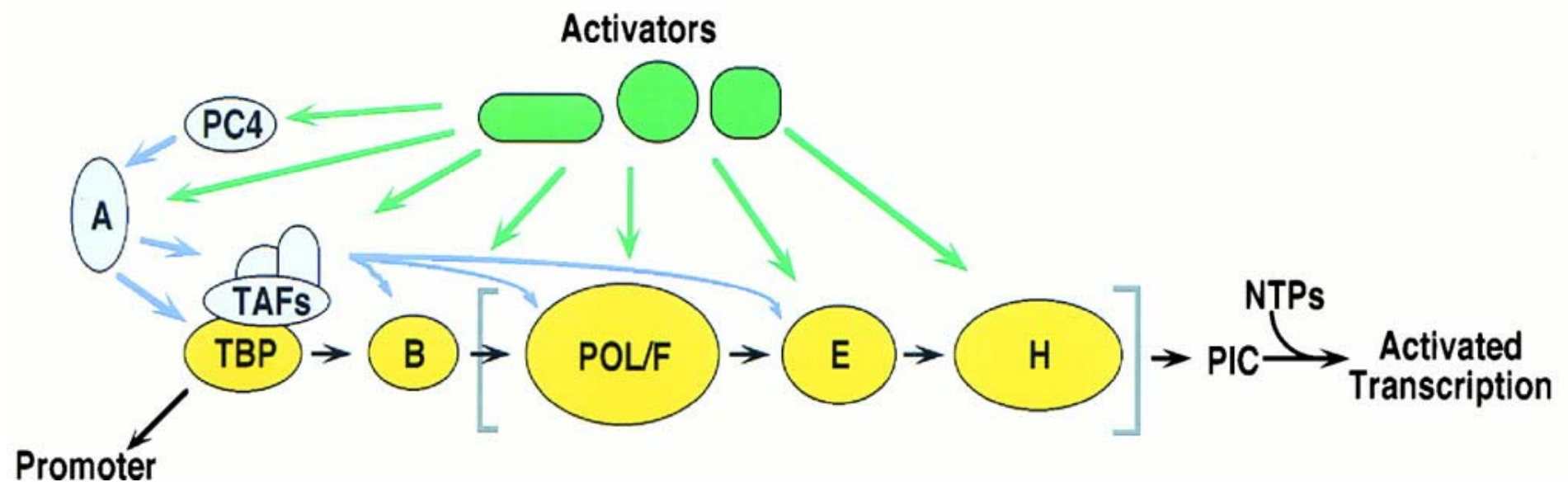
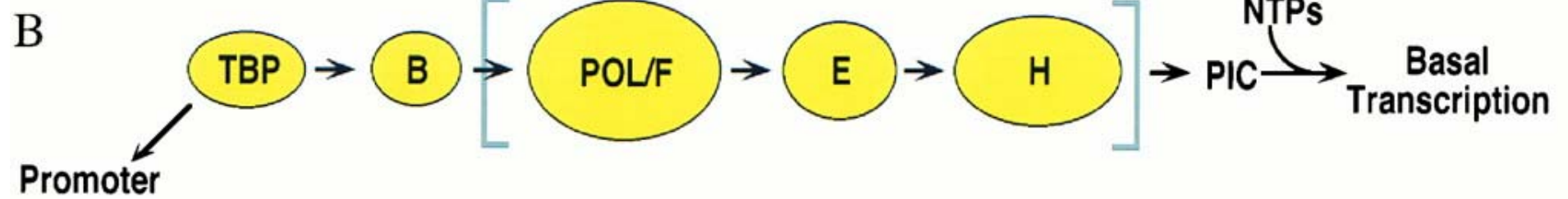
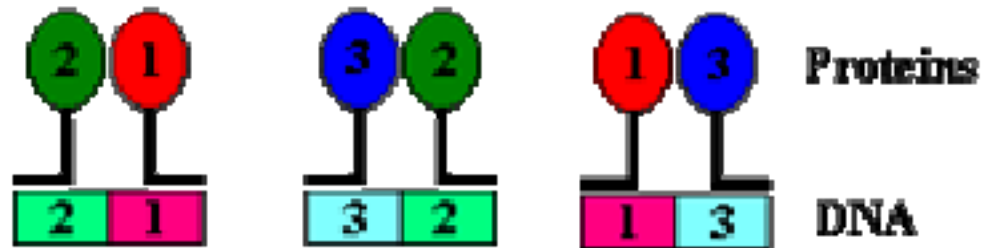
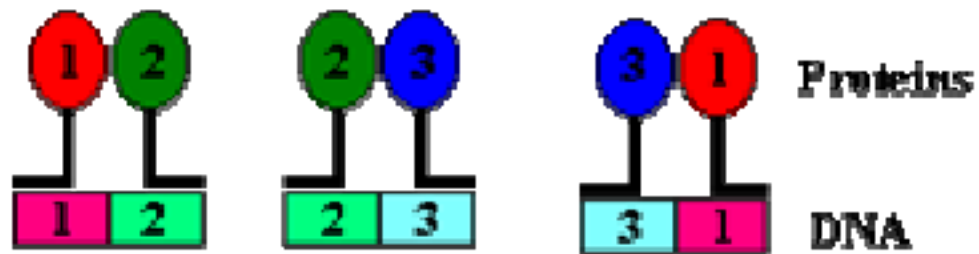
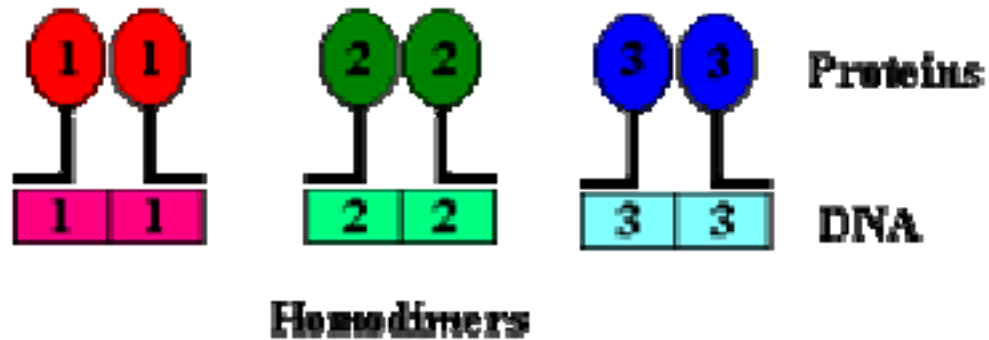


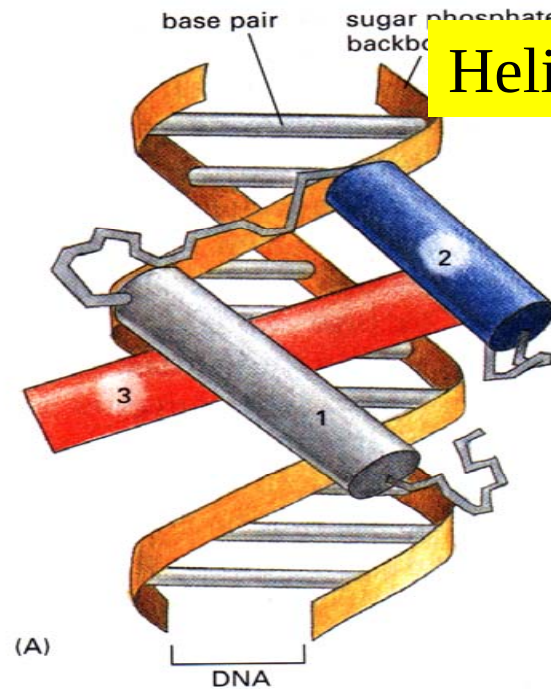


DNA bindende Proteine haben eine DNA-Bindedomäne und binden oft als Dimere

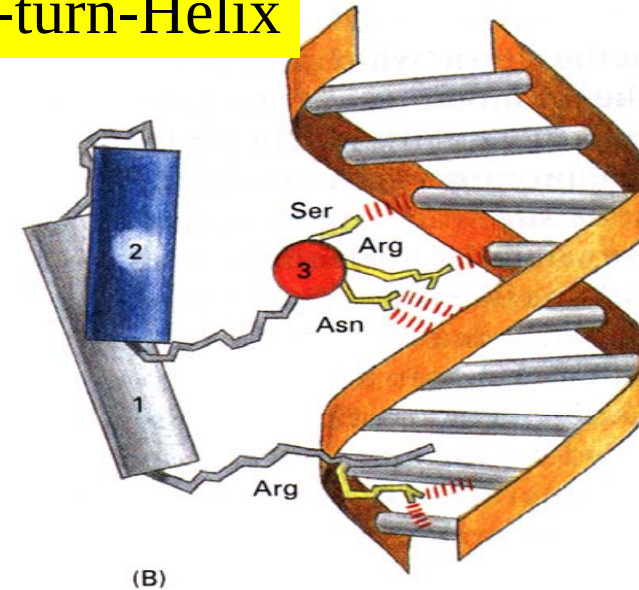


Heterodimers

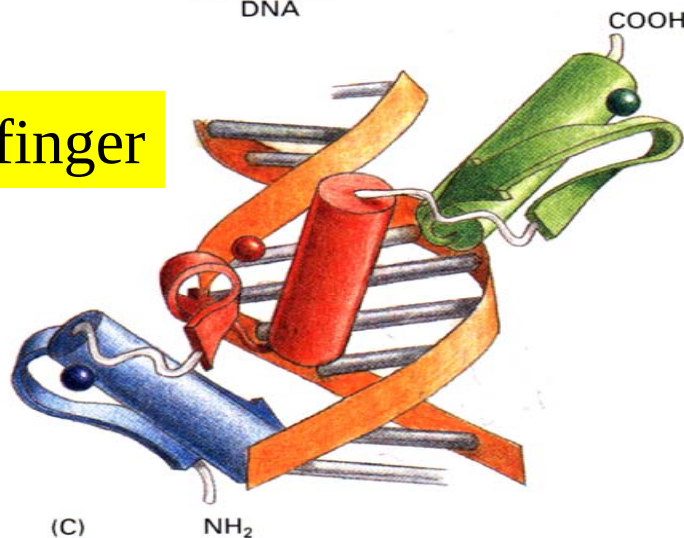
Die wichtigsten DNA-binde-Proteine



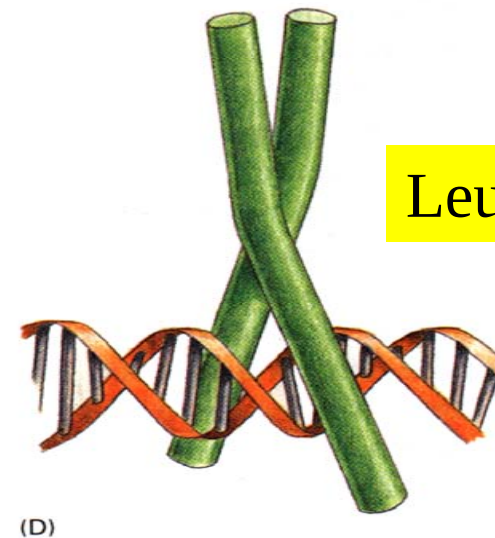
Helix-turn-Helix



Zinkfinger

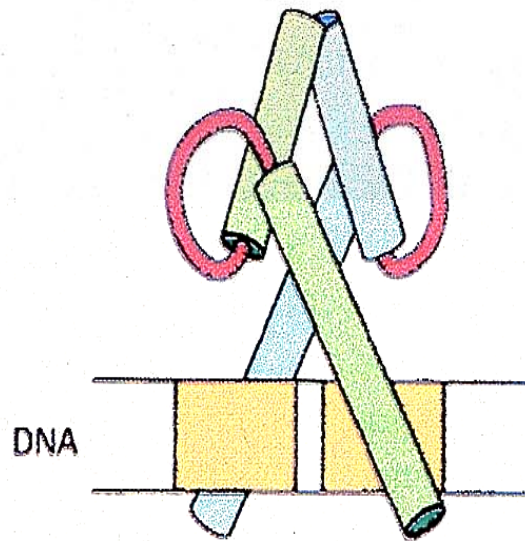


Leucin Zipper

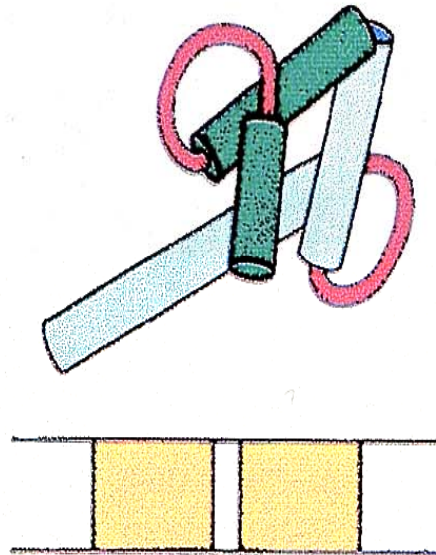


Basisches Helix-Loop-Helix

aktives HLH-Homodimer



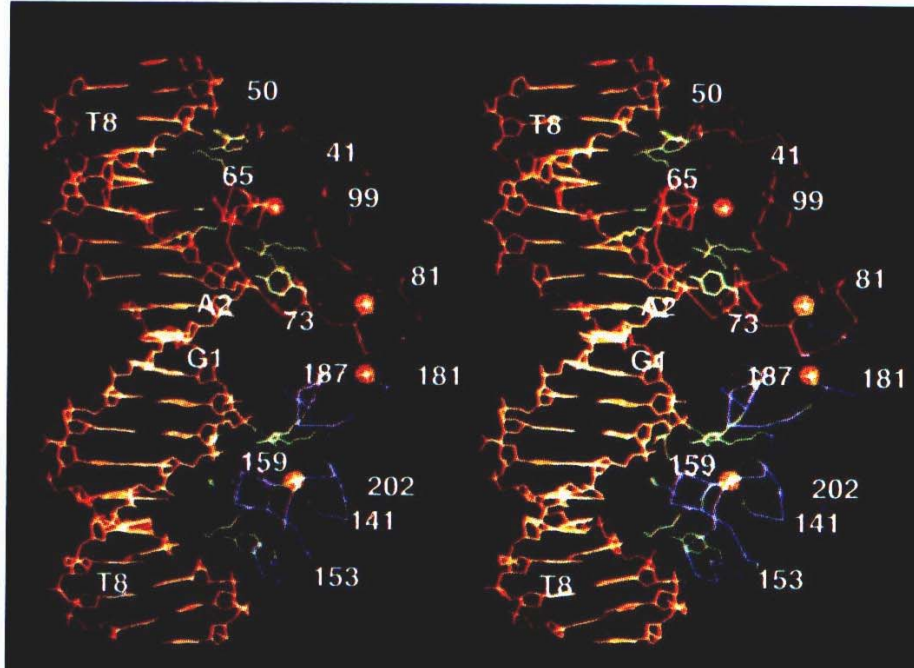
inaktives HLH-Heterodimer



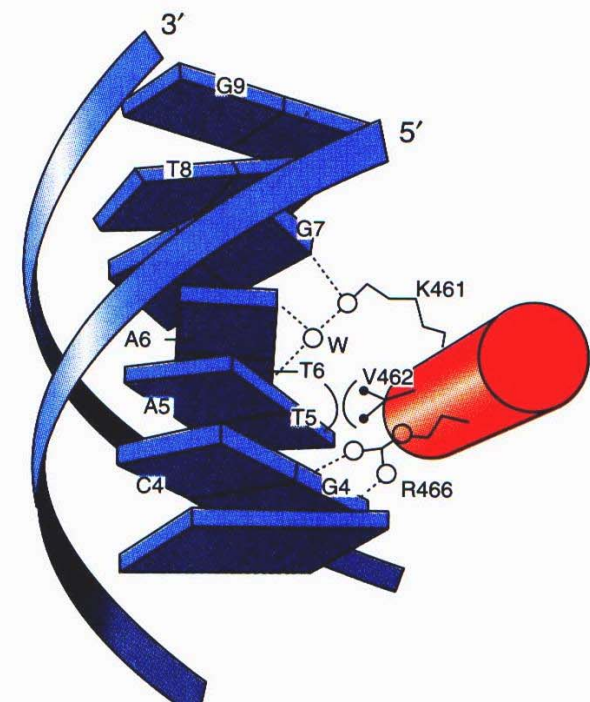
Helix-Loop-Helix

H-Brückenbindungen zwischen
Aminosäuren des Proteins und Basen der
DNA über die große Grube stellen die
sequenzspezifische Bindung sicher

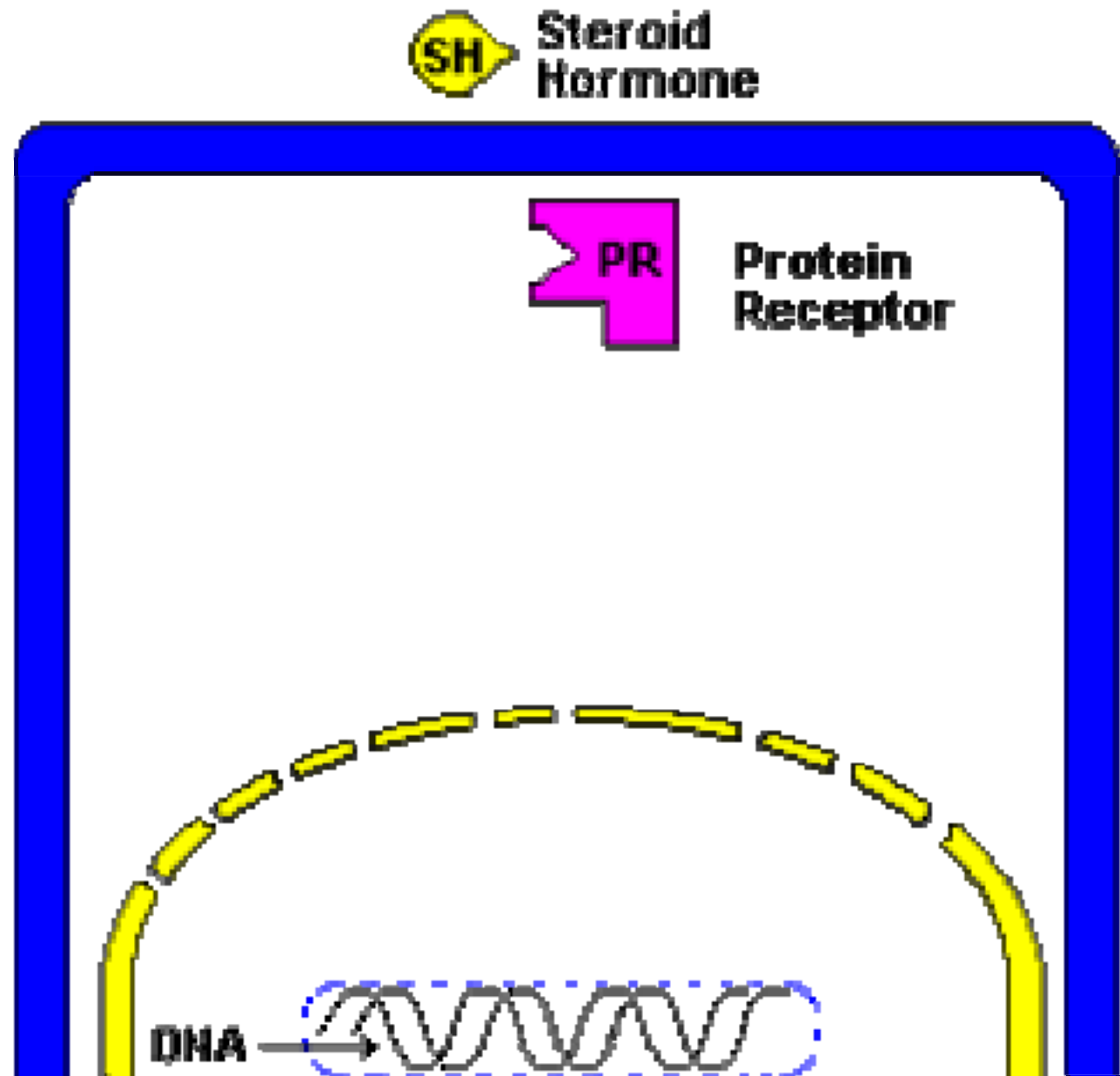
(a)



(b)

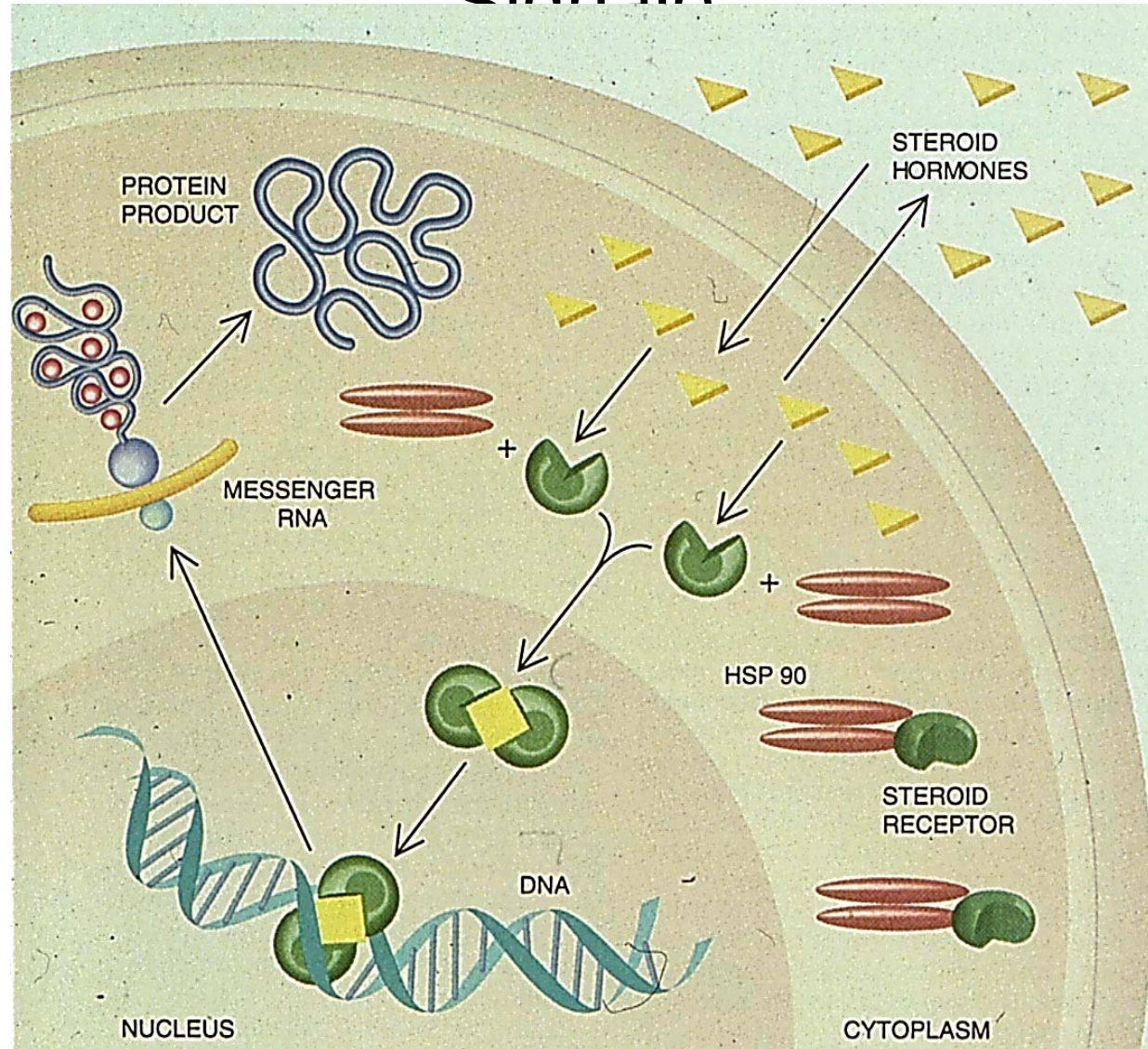


Genregulation durch Steroidhormone



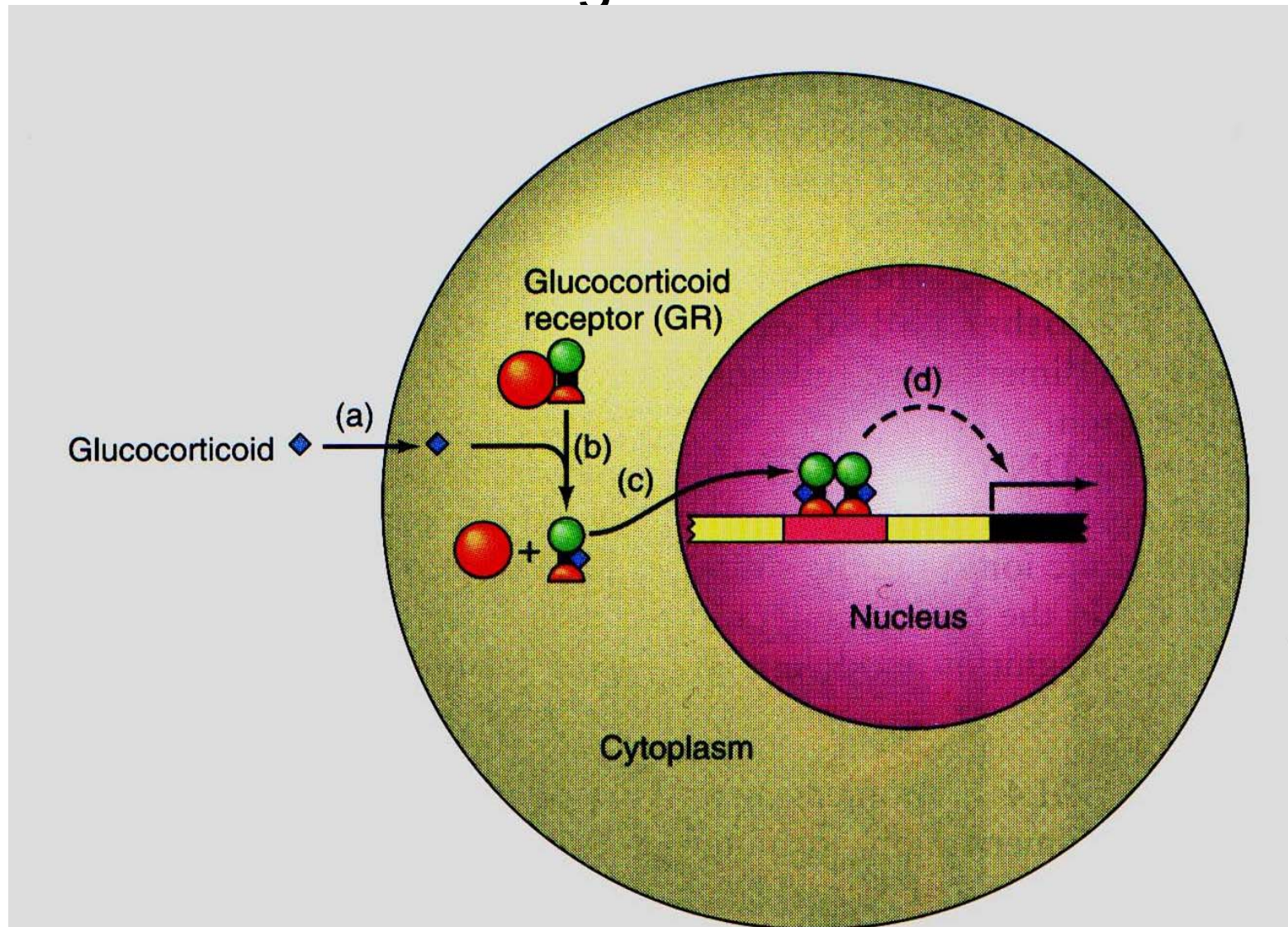
Hormon induzierte Genaktivität

Beispiel für Genaktivierung durch externe Signale



Hormon induzierte Genaktivität

Beispiel für Genaktivierung durch externe Signale



Nuclear Receptors

Palindromic Repeats

Glucocorticoid	RGRACANNNTGTYCY
Oestrogen	RGGTCANNNTGACCY
Thyroid	RGGTCA-----TGACCY

Direct Repeats

6-cis retinoic acid	AGGTCAN ₁ AGGTCA
All trans retinoic acid	AGGTCAN ₂ AGGTCA
Thyroid hormone	AGGTCAN ₄ AGGTCA

N indicates any nucleotide

R indicates a purine ie. A or G

Y indicates a pyrimidine ie. C or T

Nuclear Receptors

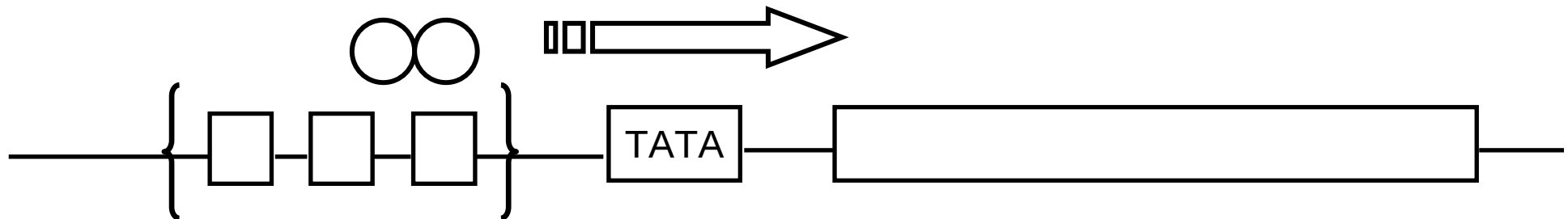
Binding of hormone

Dissociation from hsp90

Dimerisation

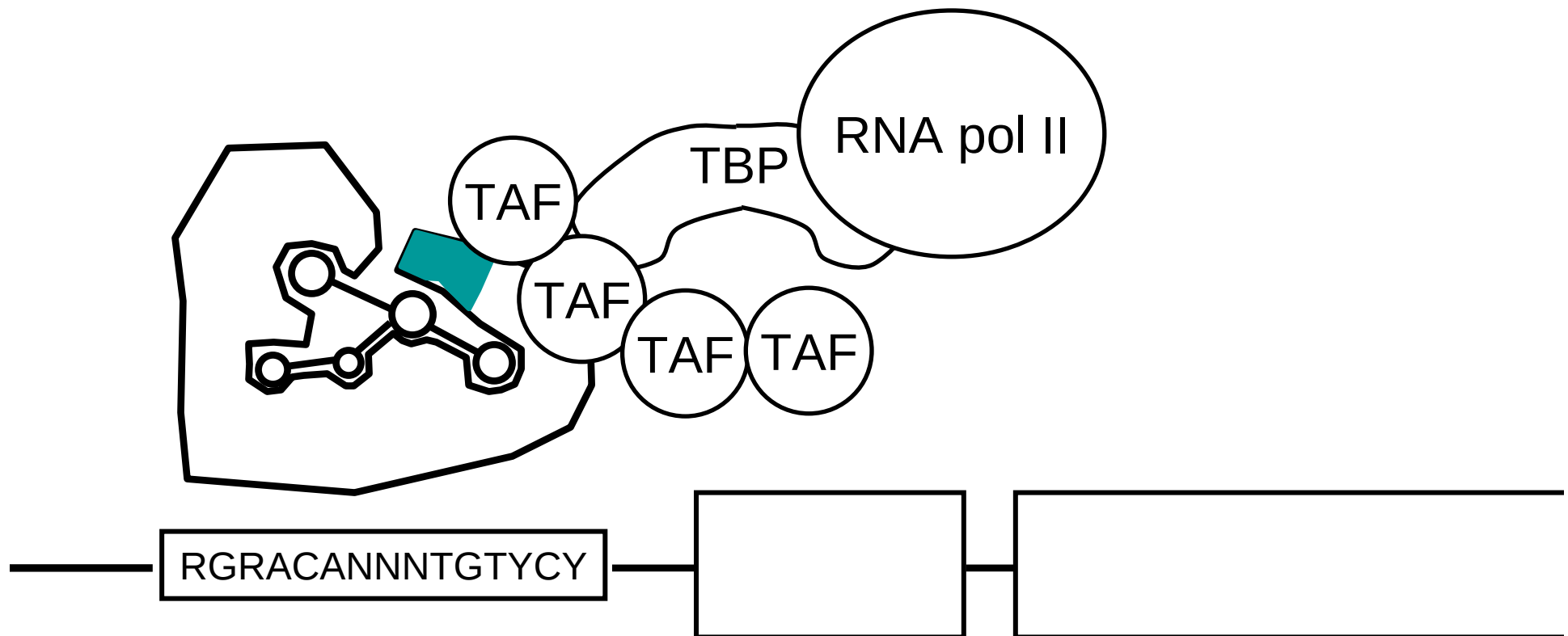
Migration to nucleus

Binding to URE

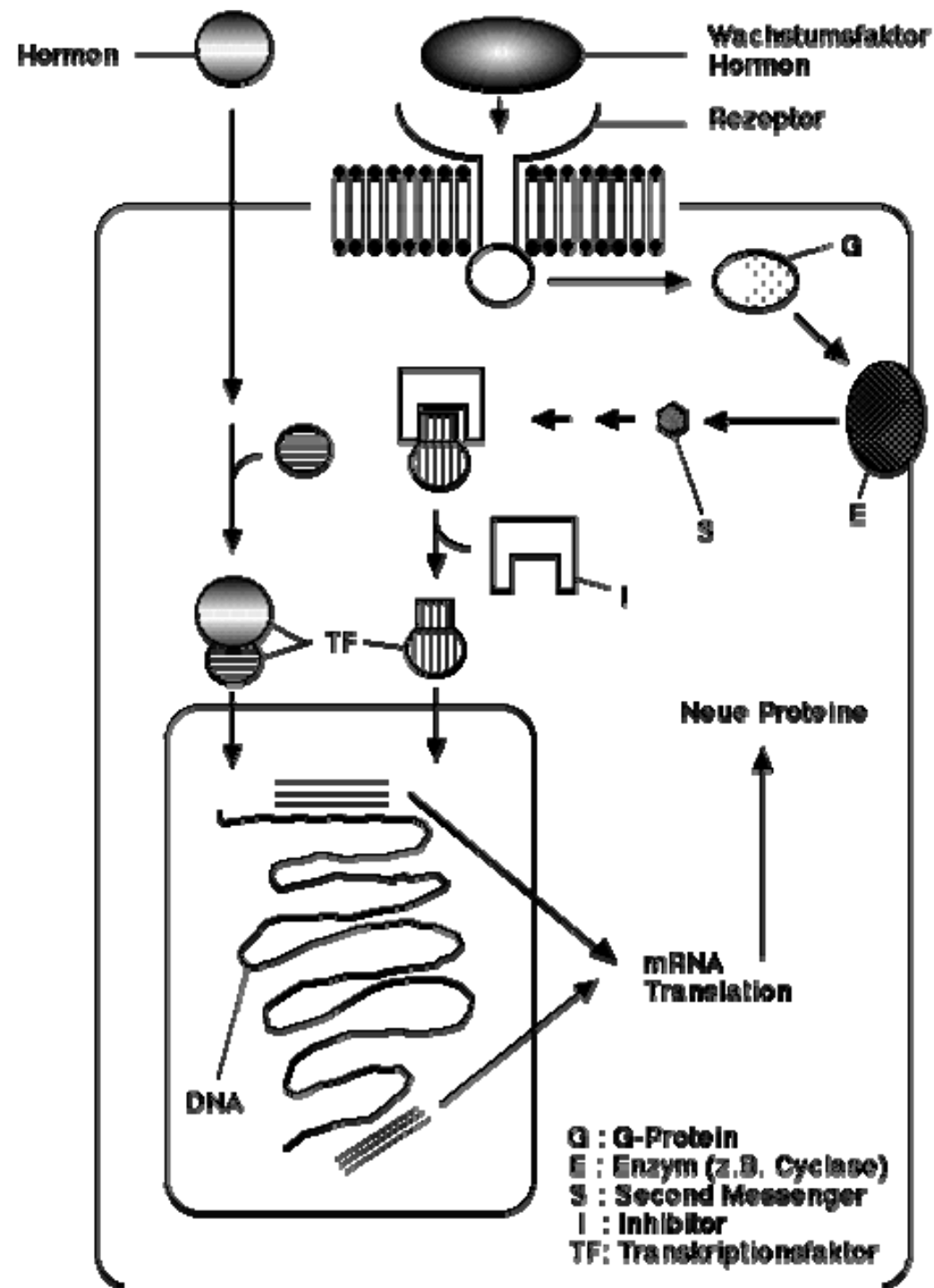


Nuclear Receptors

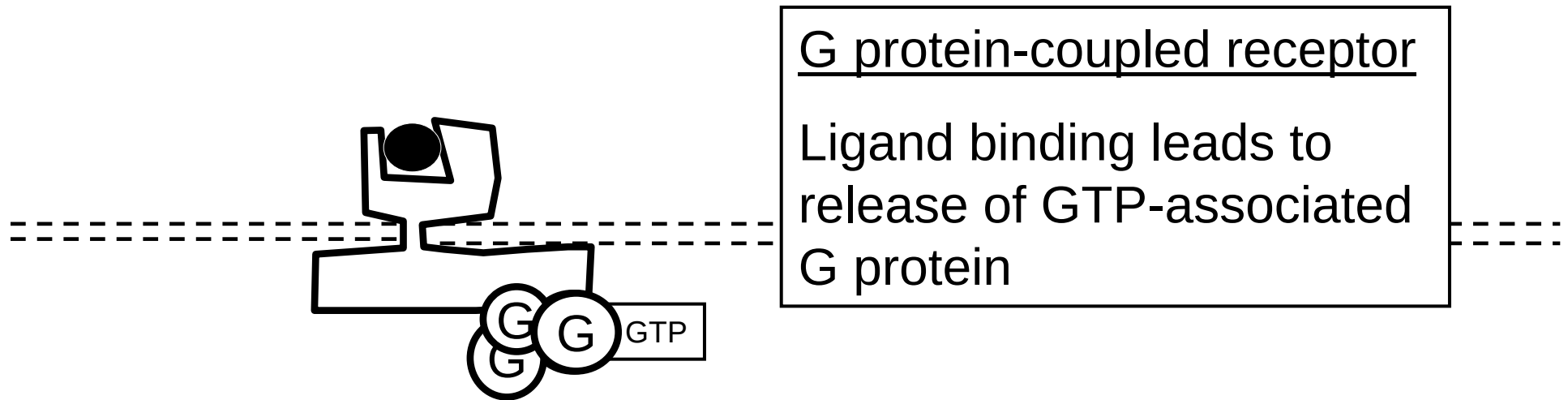
Binding of ligand causes conformational change allowing transactivation domain to interact with transcriptional machinery



Hormon- regulation von Genen



cAMP signalling

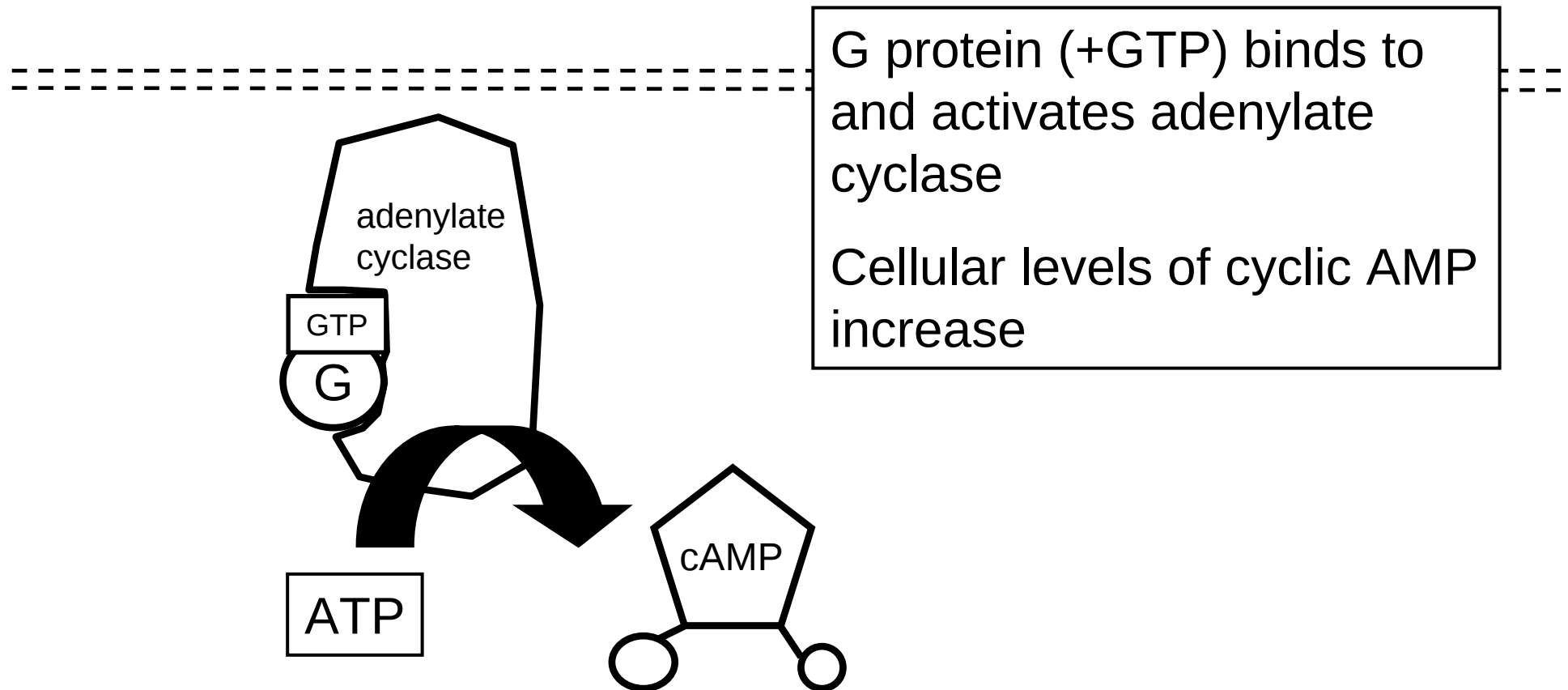


Small GTTP-binding proteins

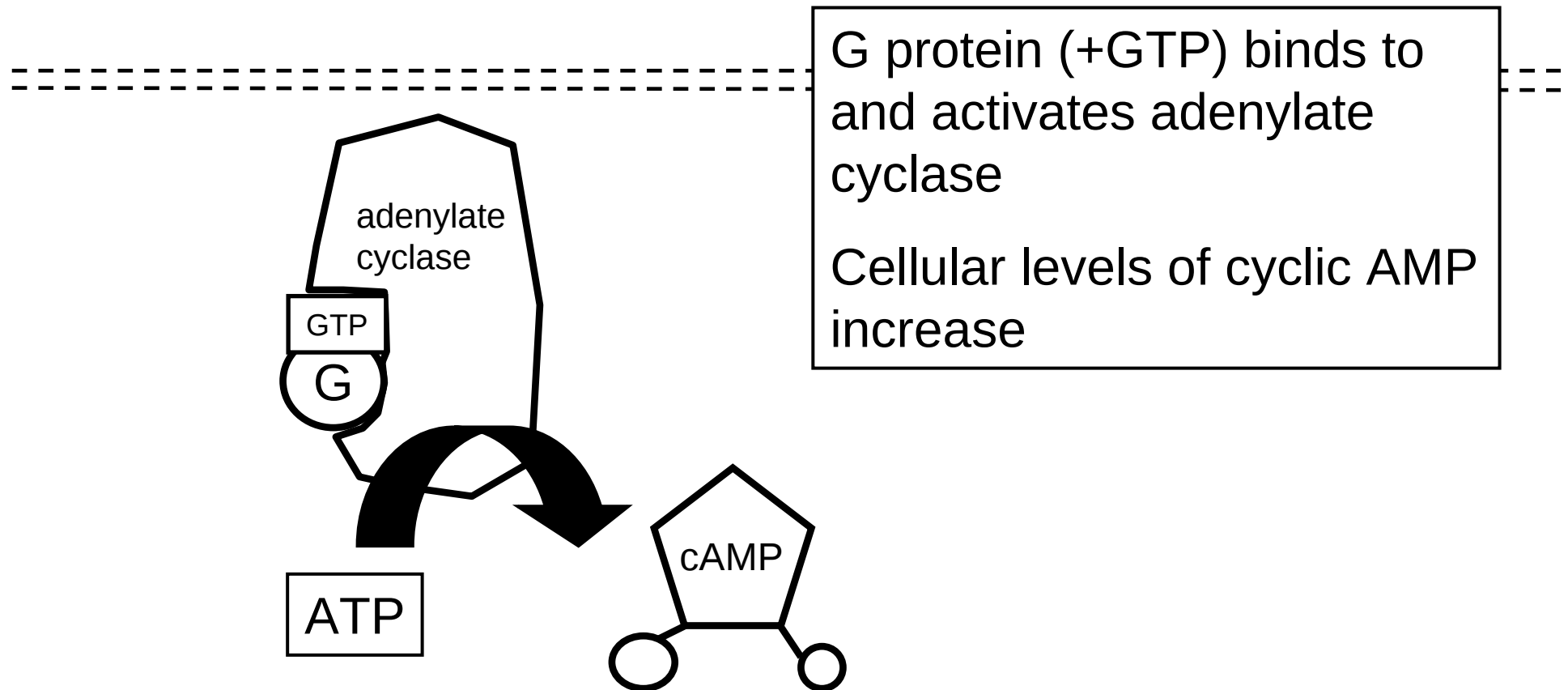
Example: ras – covered in earlier lecture

Alternate between inactive GDP and active GTP bound forms

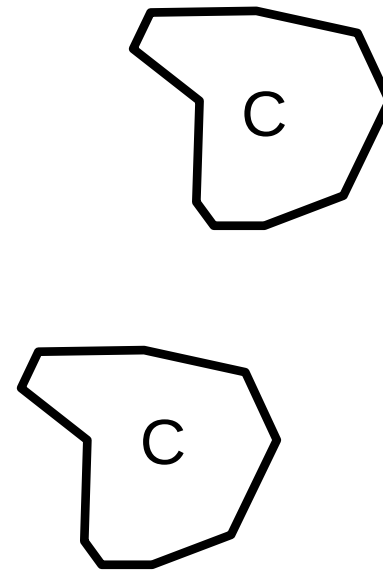
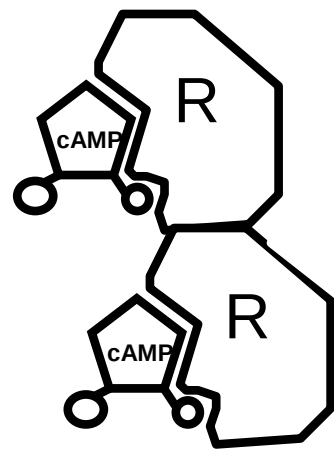
cAMP signalling



cAMP signalling



cAMP signalling

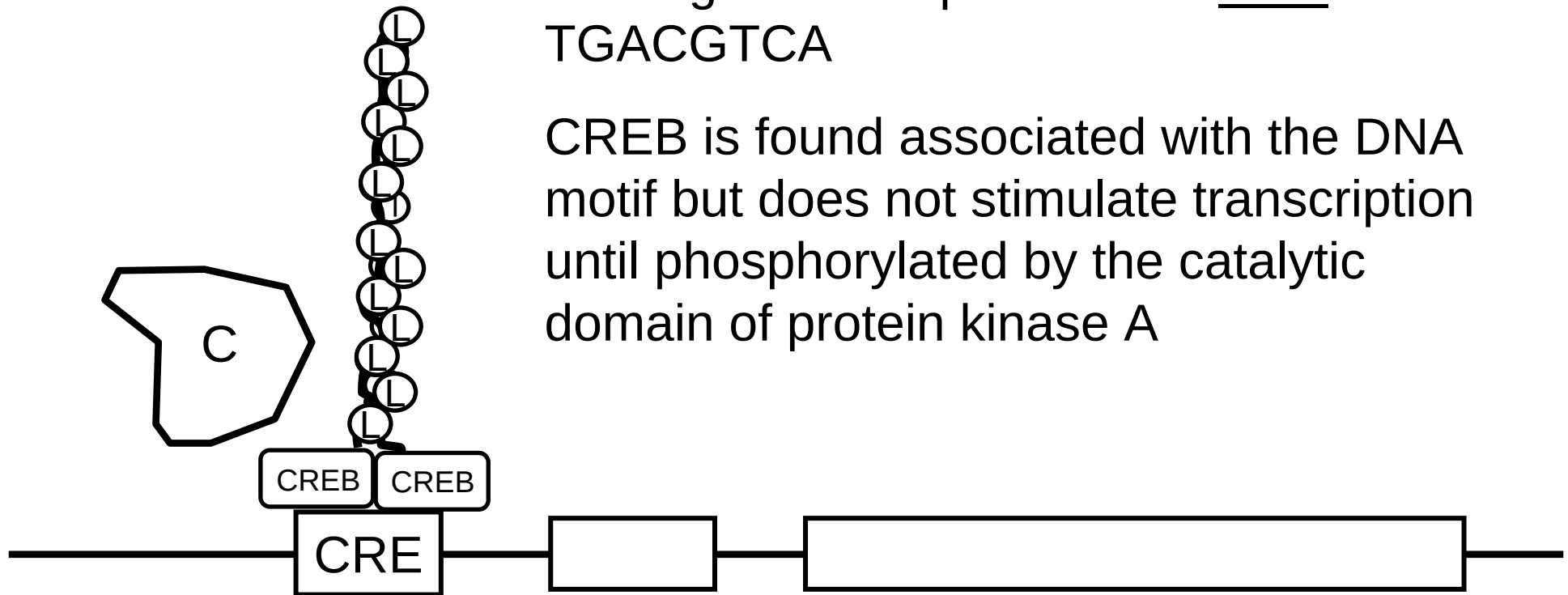


Cyclic AMP Response Element Binding protein

CREB is a member of the bZIP family of transcription factors

It recognizes the palindromic CRE motif TGACGTCA

CREB is found associated with the DNA motif but does not stimulate transcription until phosphorylated by the catalytic domain of protein kinase A

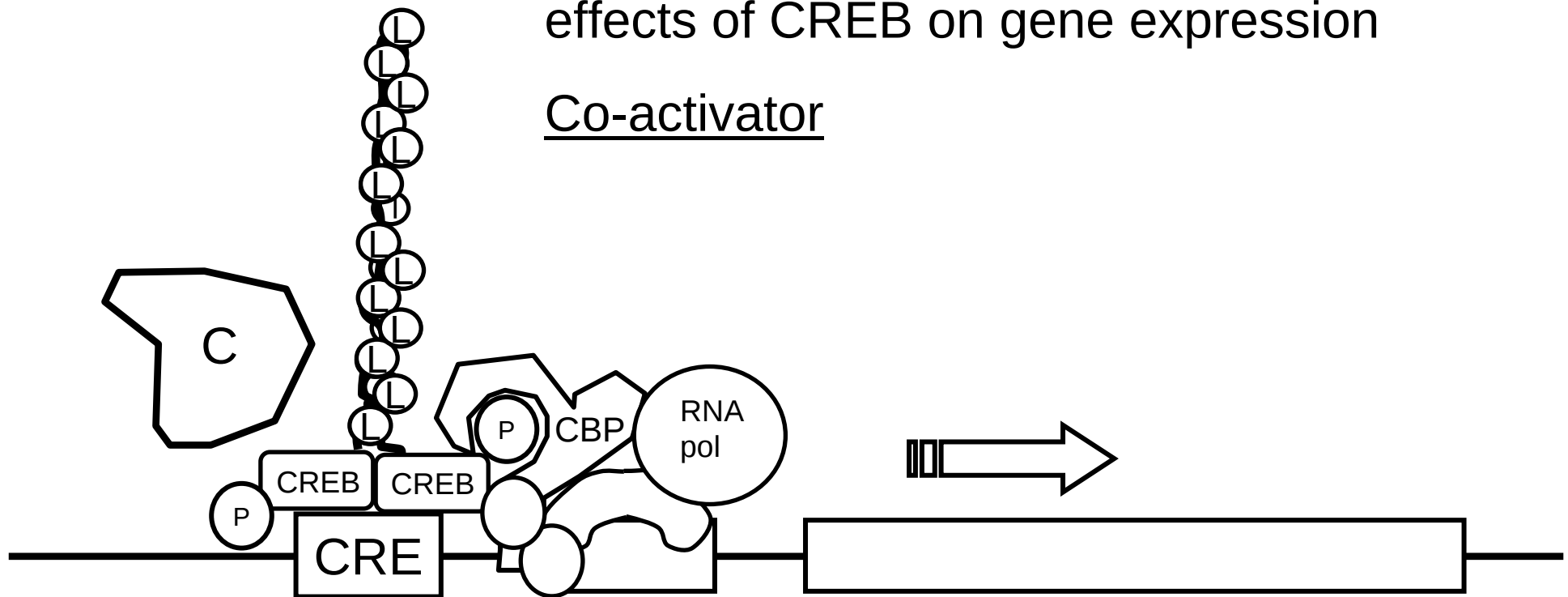


CREB binding protein

CBP recognises and binds the
phosphorylated form of CREB

CBP interacts with the basal
transcription complex to mediate the
effects of CREB on gene expression

Co-activator



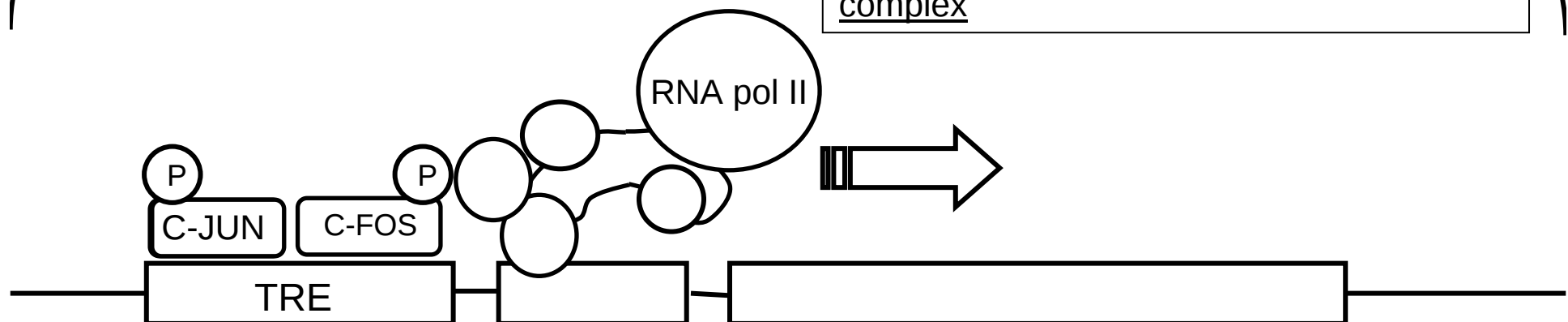
Kinase Cascades

Active JNK travels to the nucleus and phosphorylates the bZIP transcription factor C-JUN

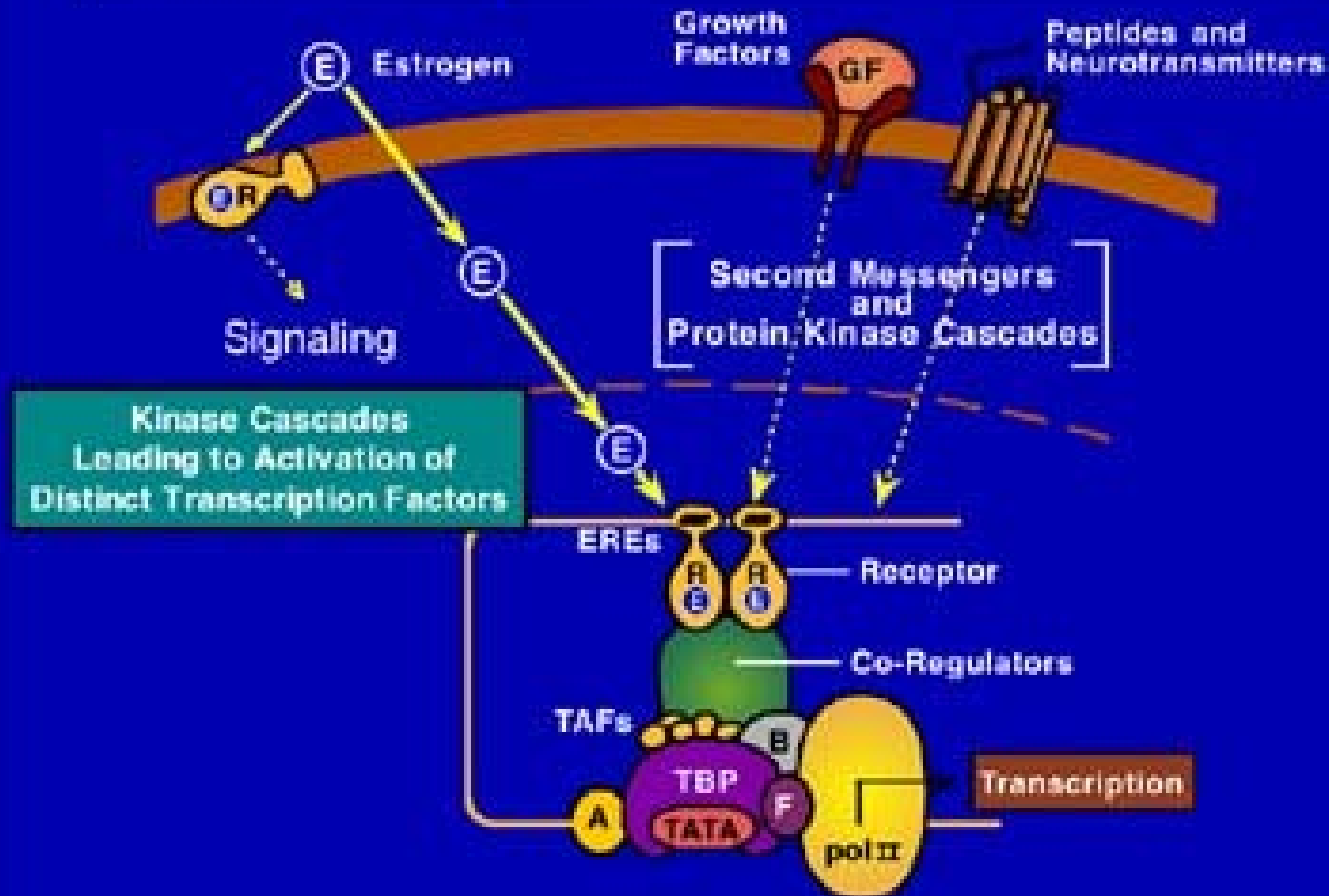
nucleus

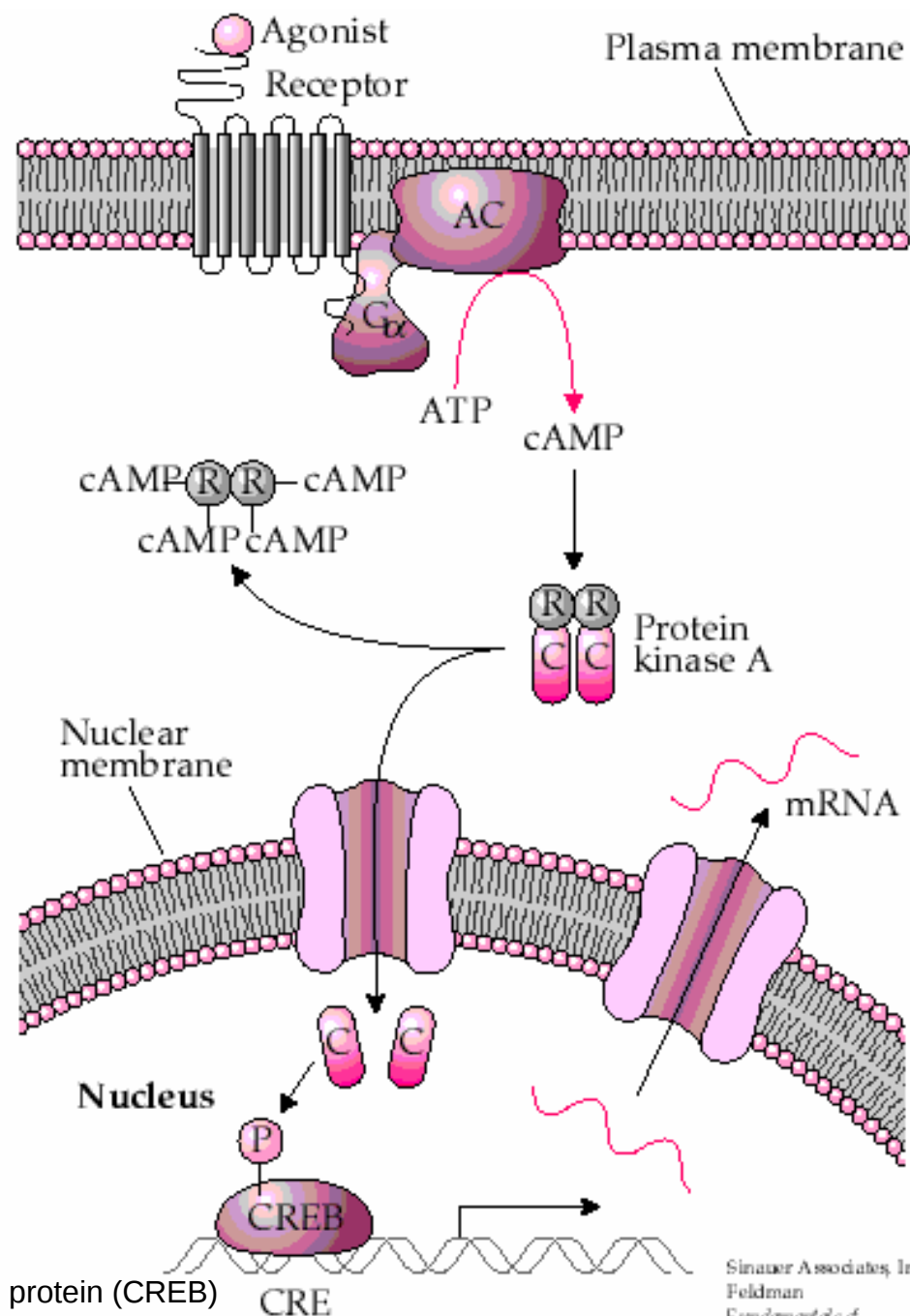
C-JUN dimerizes with C-FOS to form the AP-1 transcription factor

AP-1 activates transcription by binding to the -TGA(C/G)TCA- TPA response element (TRE) (seen in lecture 1) and interacting with the basal transcription complex



Receptor Actions in Cells

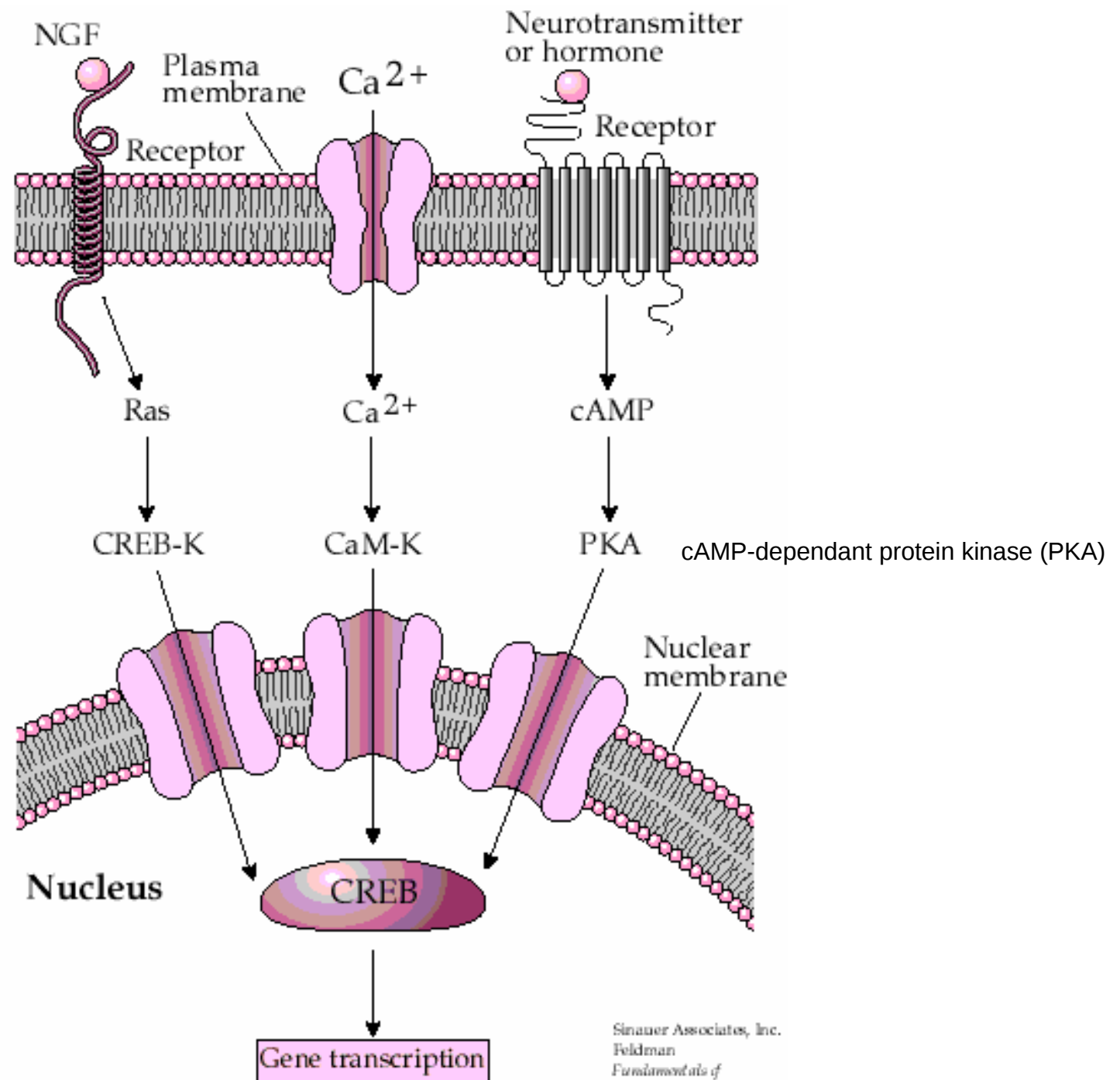


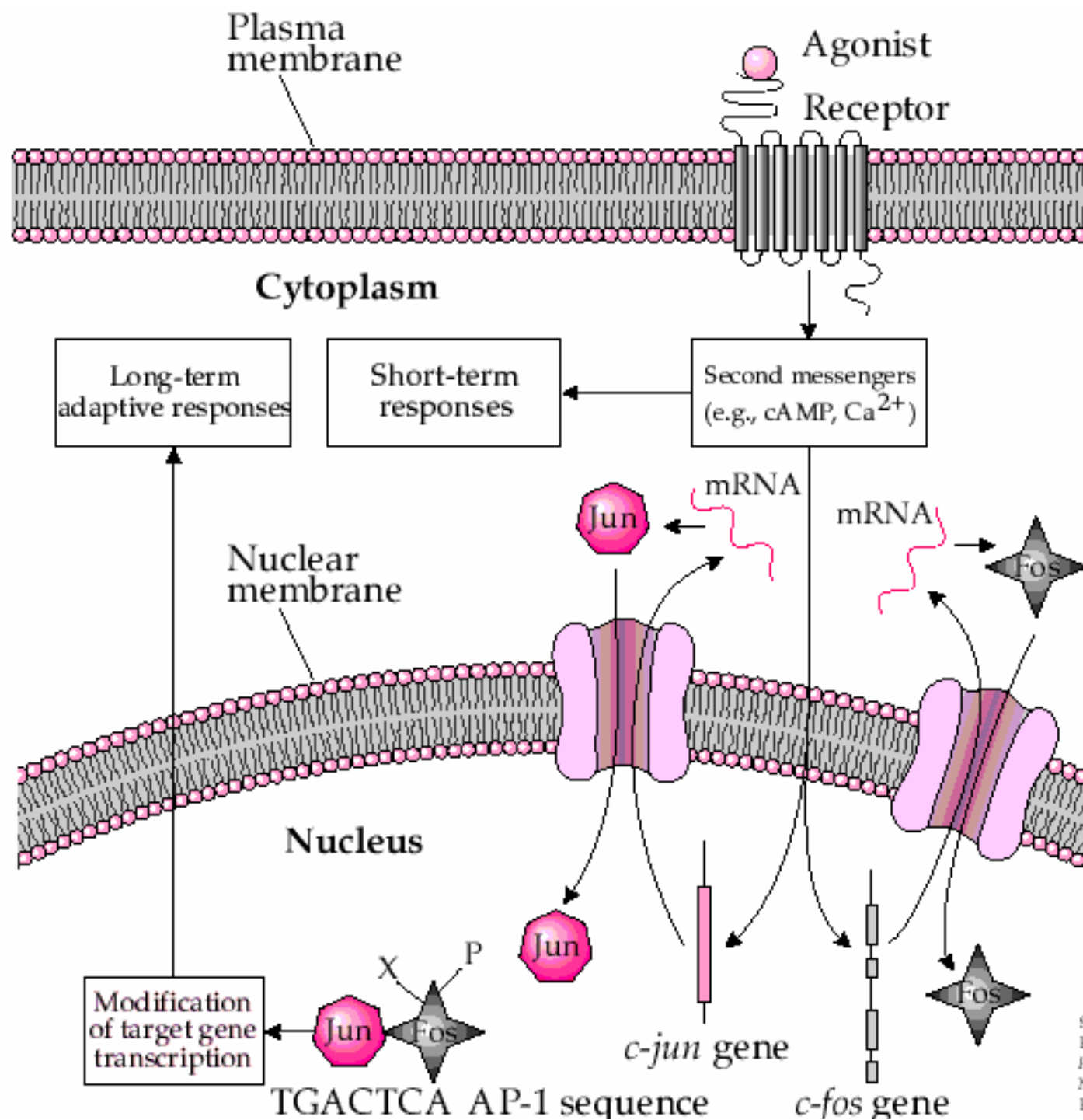


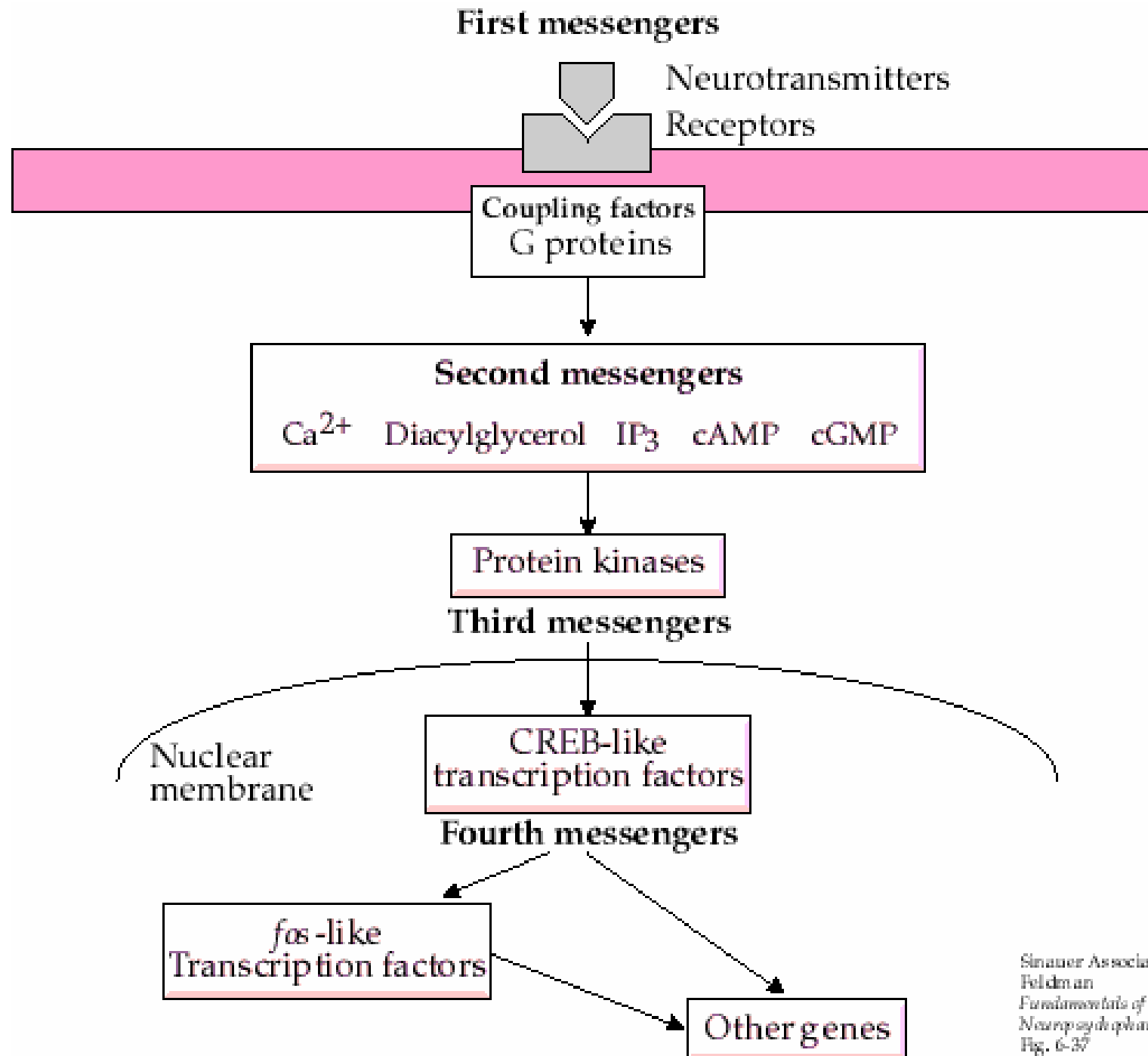
cAMP response element binding protein (CREB)

AC Adenylate cyclase

Sinatra Associates, Inc.
Feldman
*Fundamentals of
Neuropharmacology*
Fig. 6-34







Chromatin und Genregulation

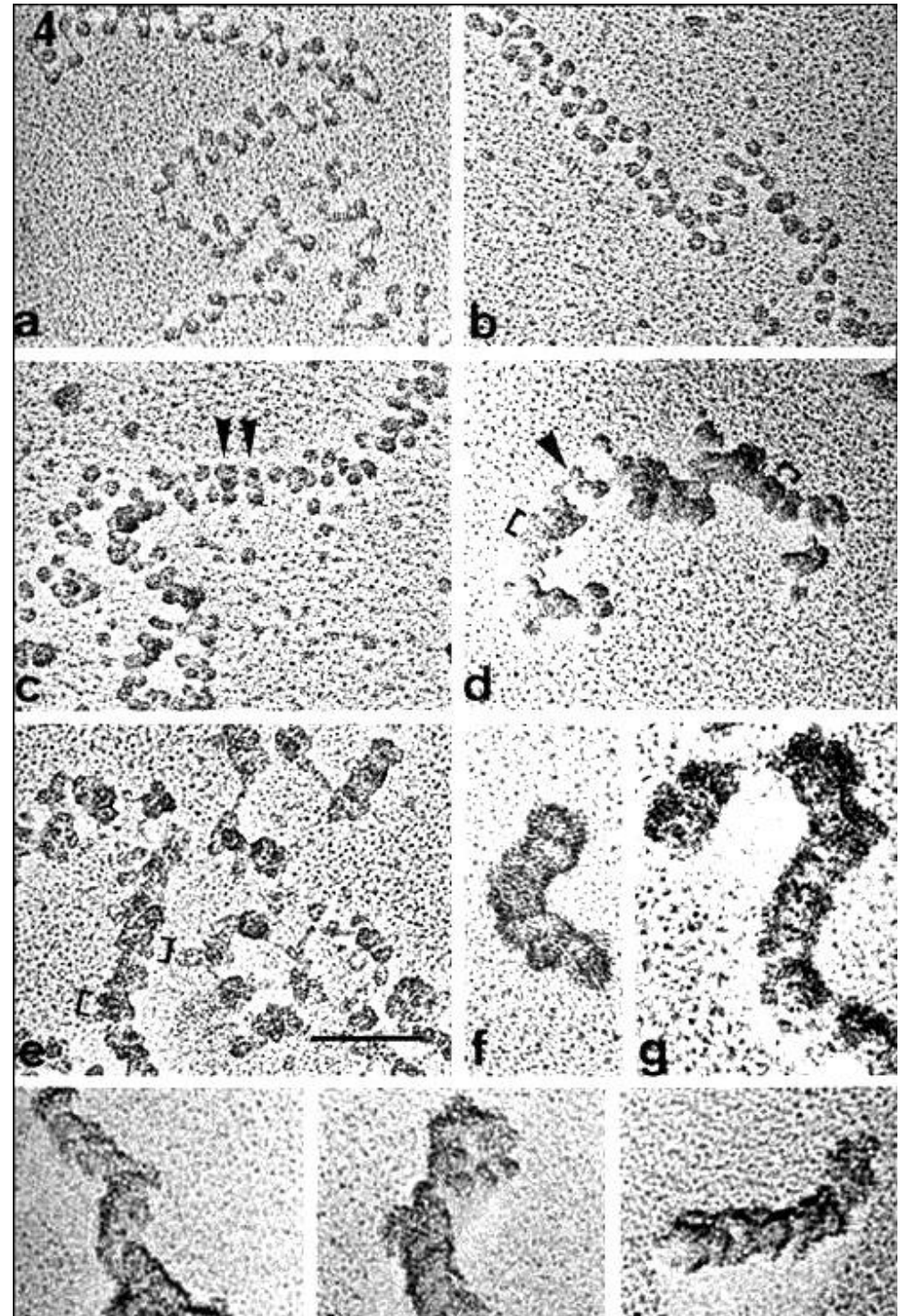
Chromatin ist eine wesentliche Ebene der Genregulation bei Eukaryoten

Beispiele sind Eu- und Heterochromatin

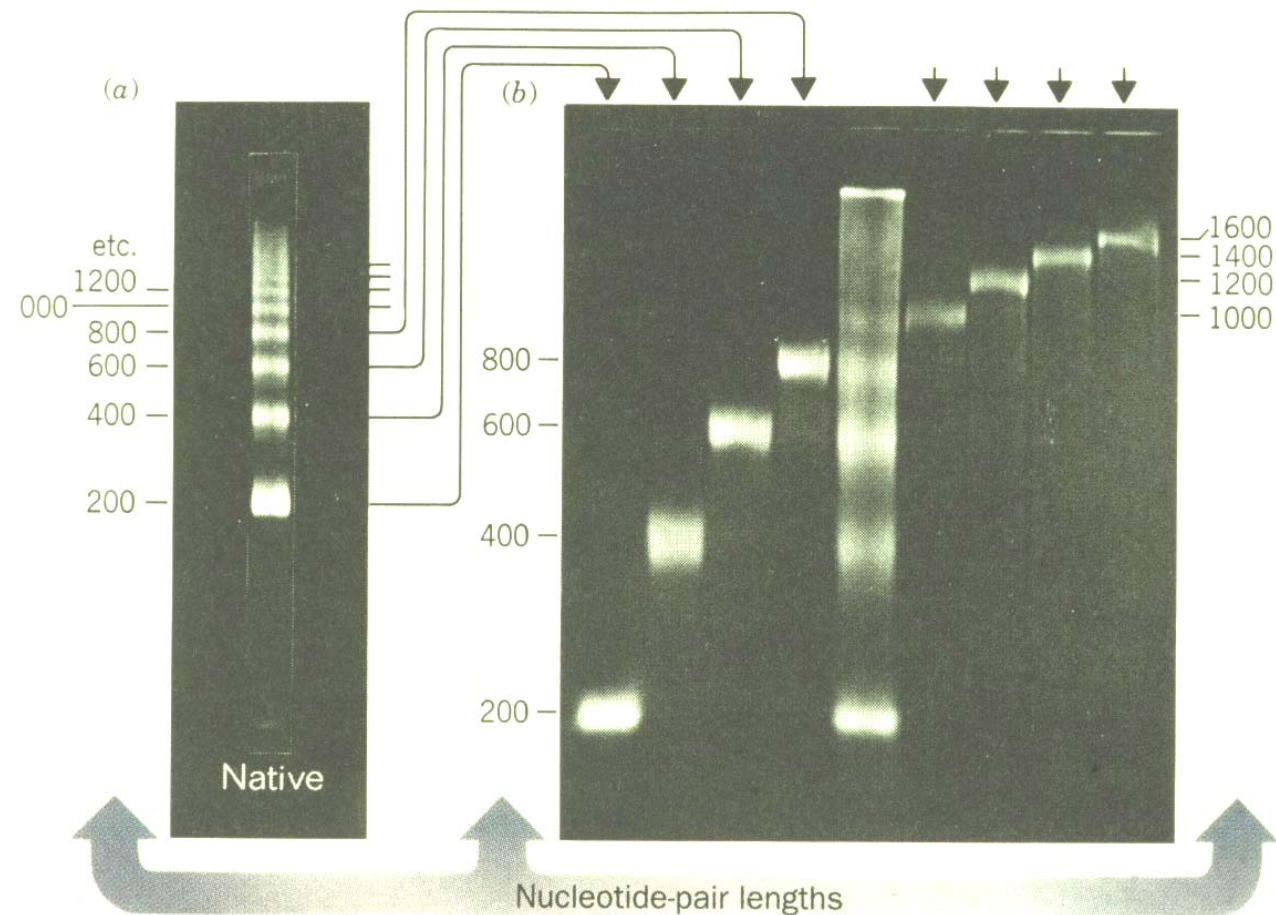
Aktives und inaktives Chromatin entstehen durch Proteinmodifikationen und unterschiedliche Zusammensetzung des Chromatins

Spezifische Proteine sind für die Bildung von Heterochromatin verantwortlich (z.B. HP1, SuVar-Proteine, Histonacetyltransferasen etc)

DNA in Chromatin: verschiedene Verpackungs- ebenen

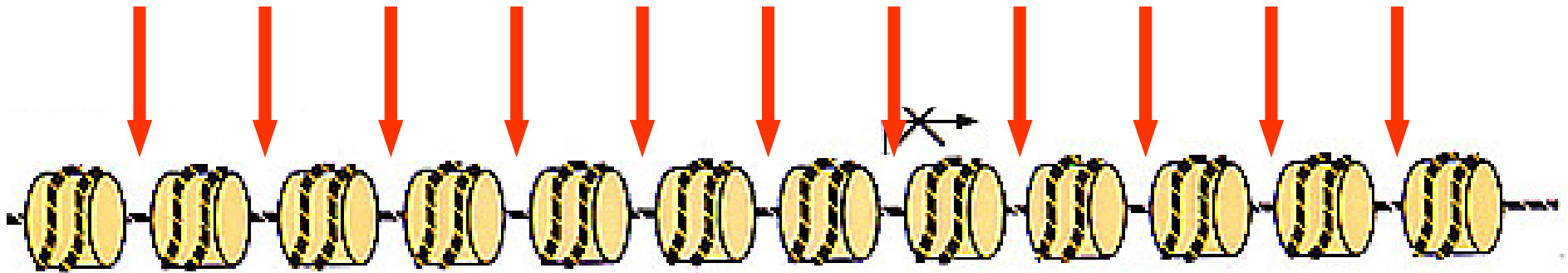


Nuclease-Verdau von Chromatin ergibt diskrete DNA-Fragmente von ca. 180 Bp Länge

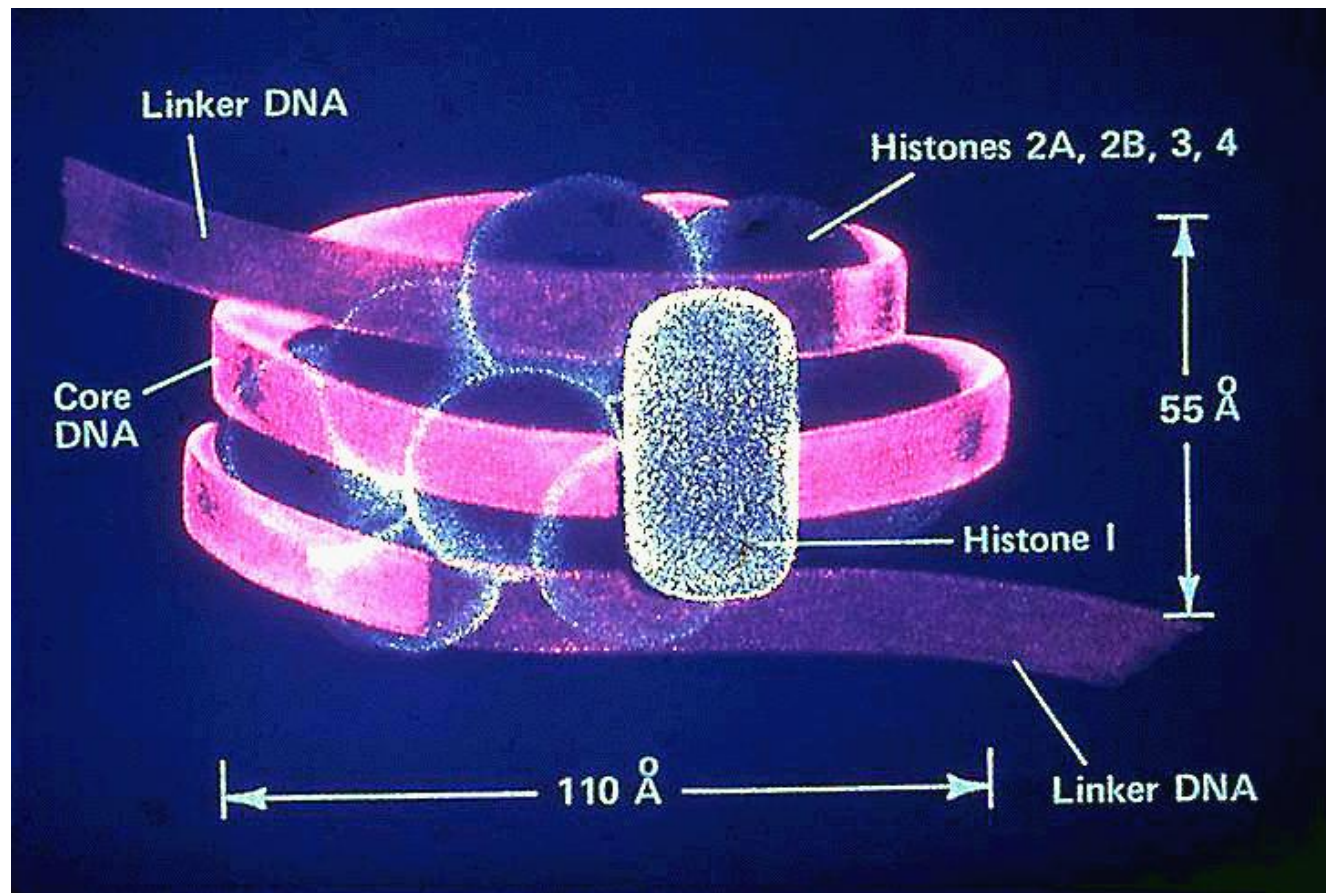


Chromatin und Genaktivität

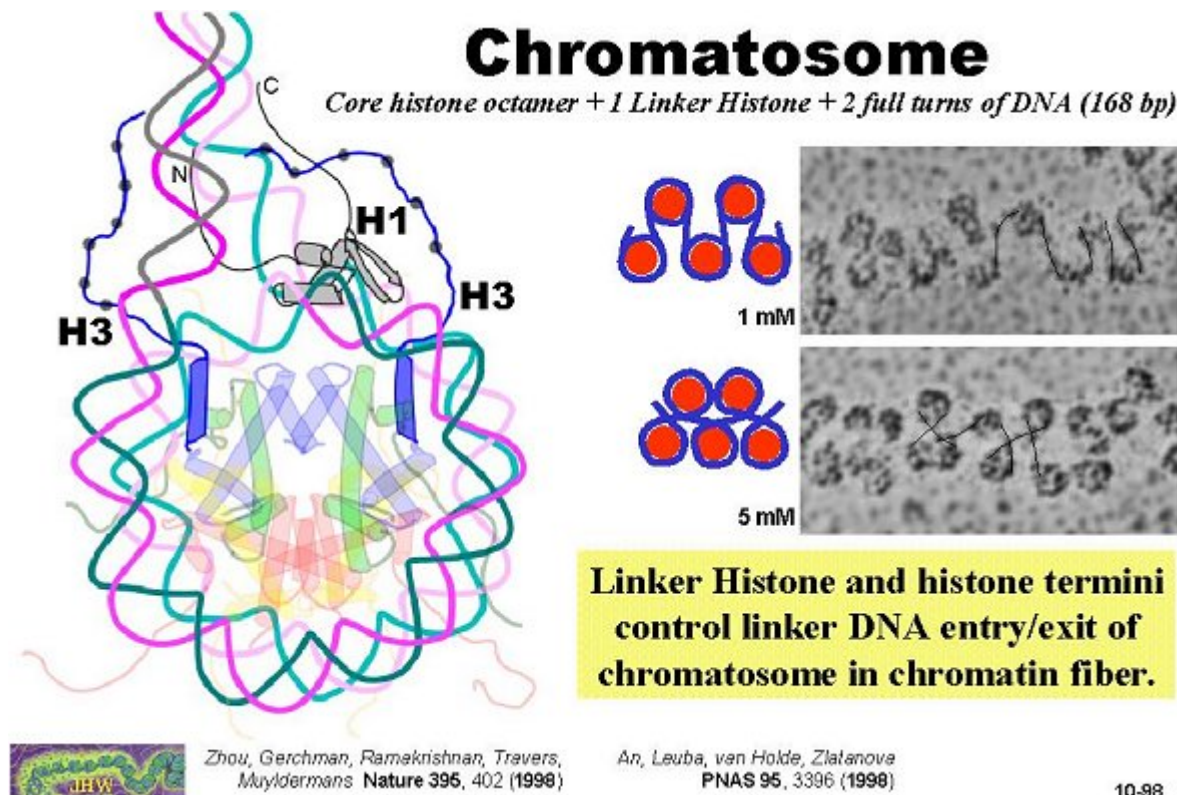
Nuklease zugängliche Chromatinbereiche



Organisation des Mononukleosoms

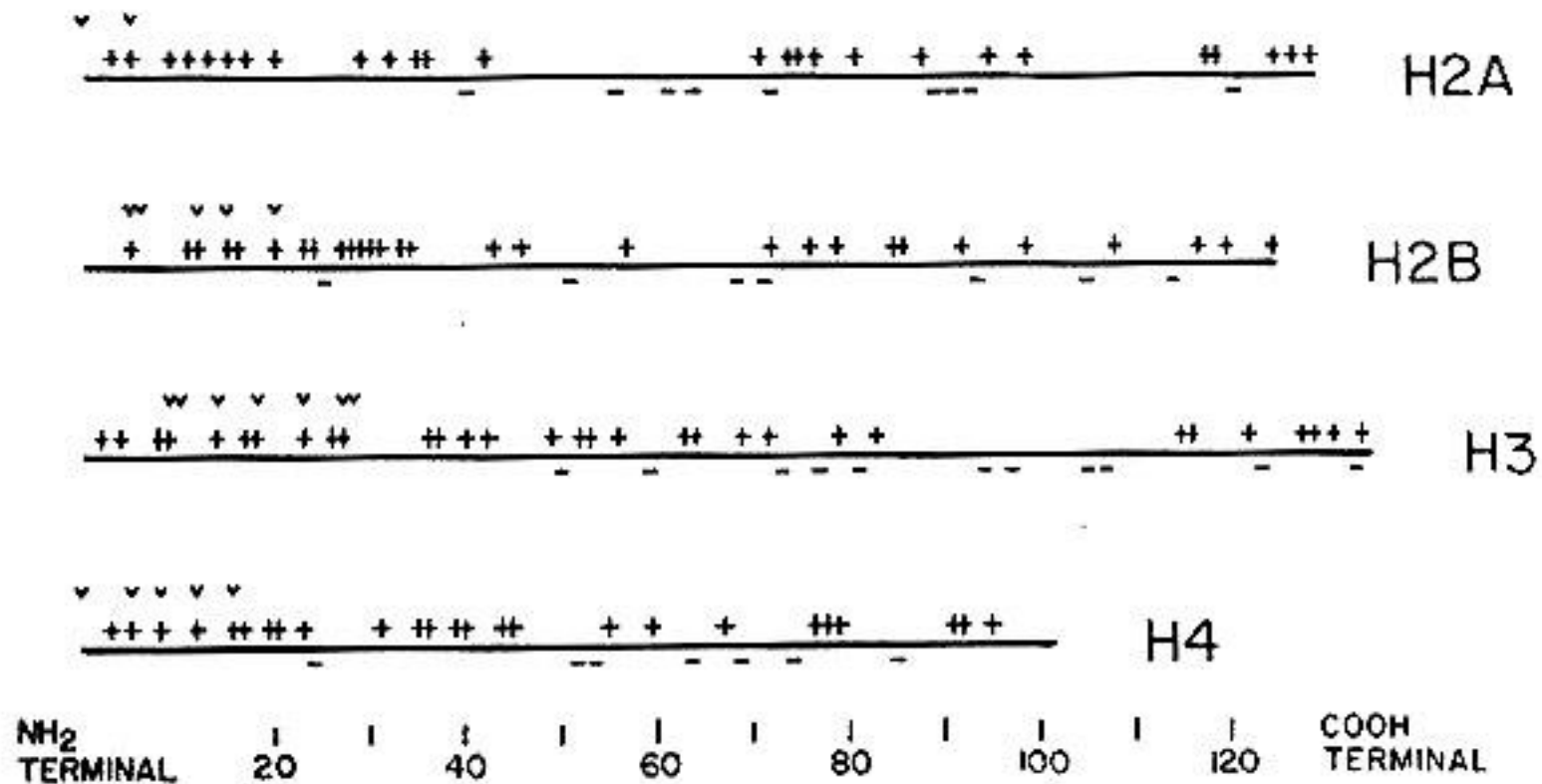


Histon H1 – „Linker Histon“

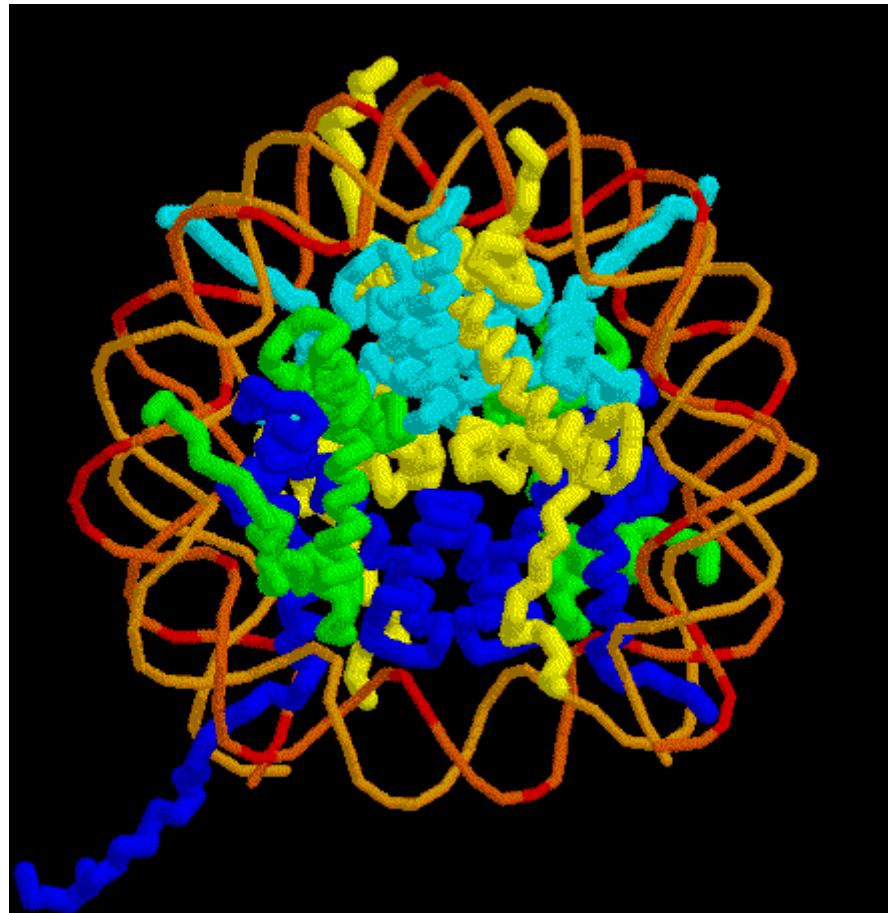


Die Core Histone: H2A, H2B, H3, H4

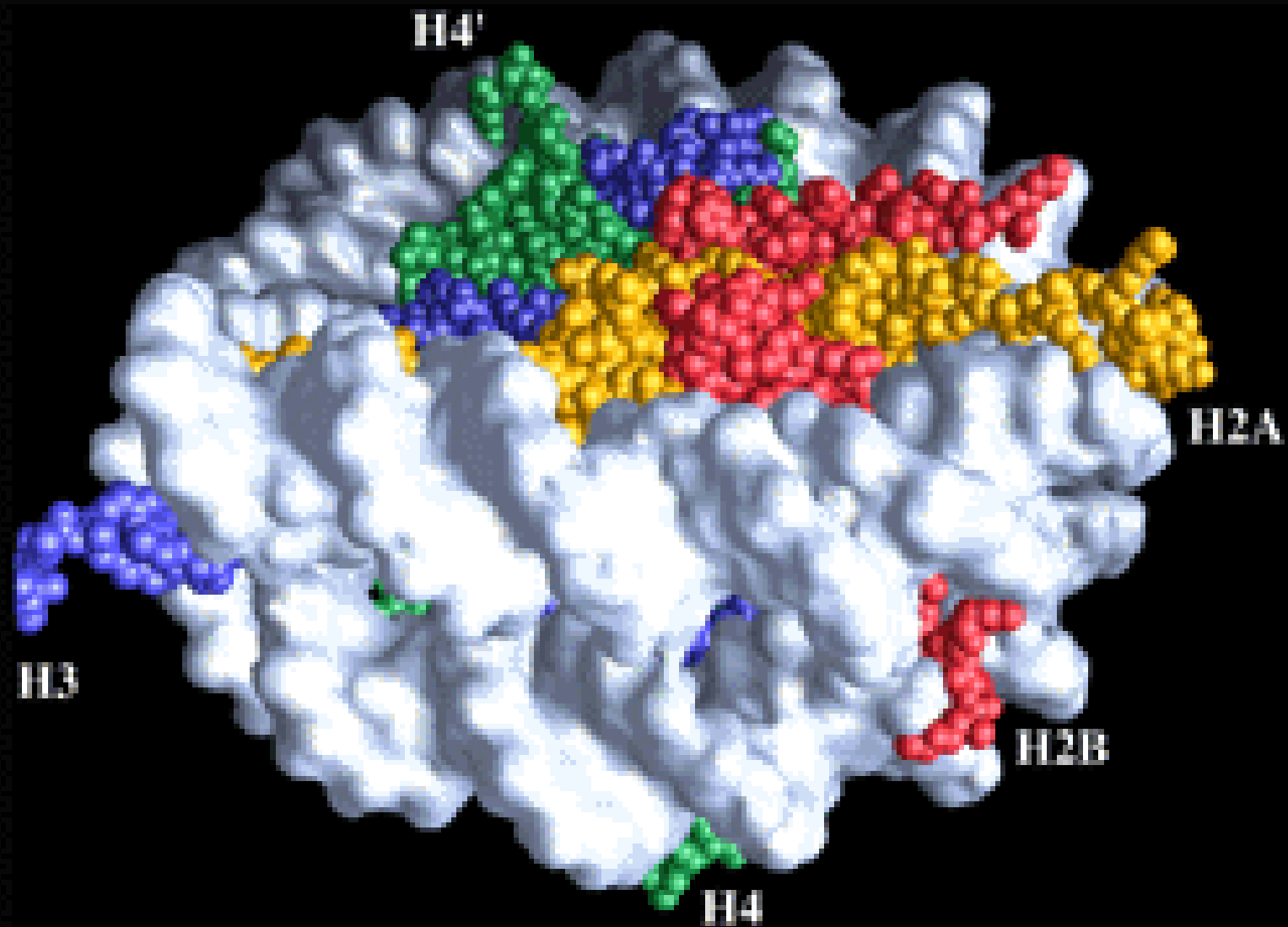
DISTRIBUTION OF CHARGED RESIDUES
WITHIN THE FOUR SMALLER HISTONES



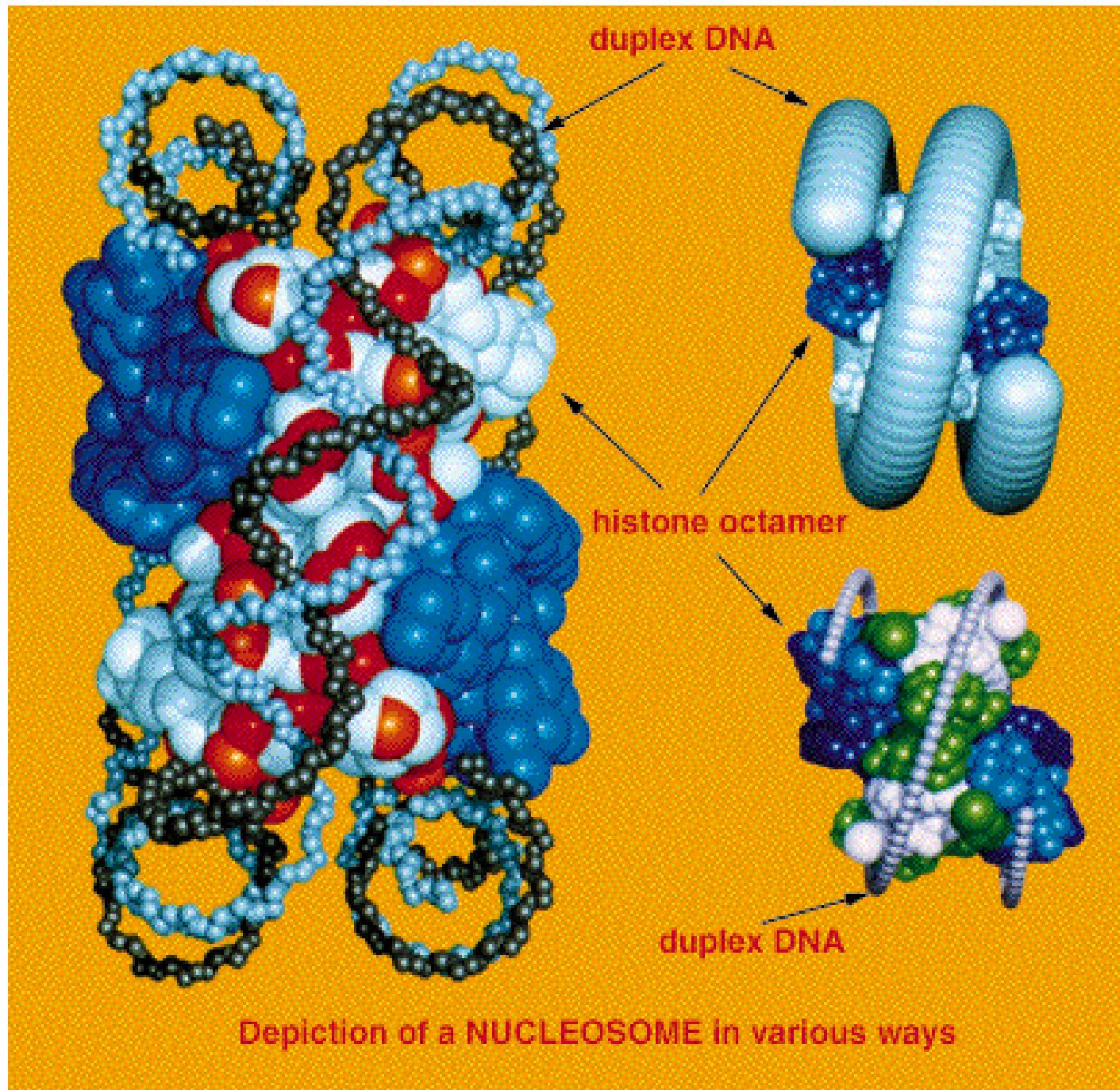
Der Aufbau des „Core“-
Nukleosoms ist im Detail
aufgeklärt:

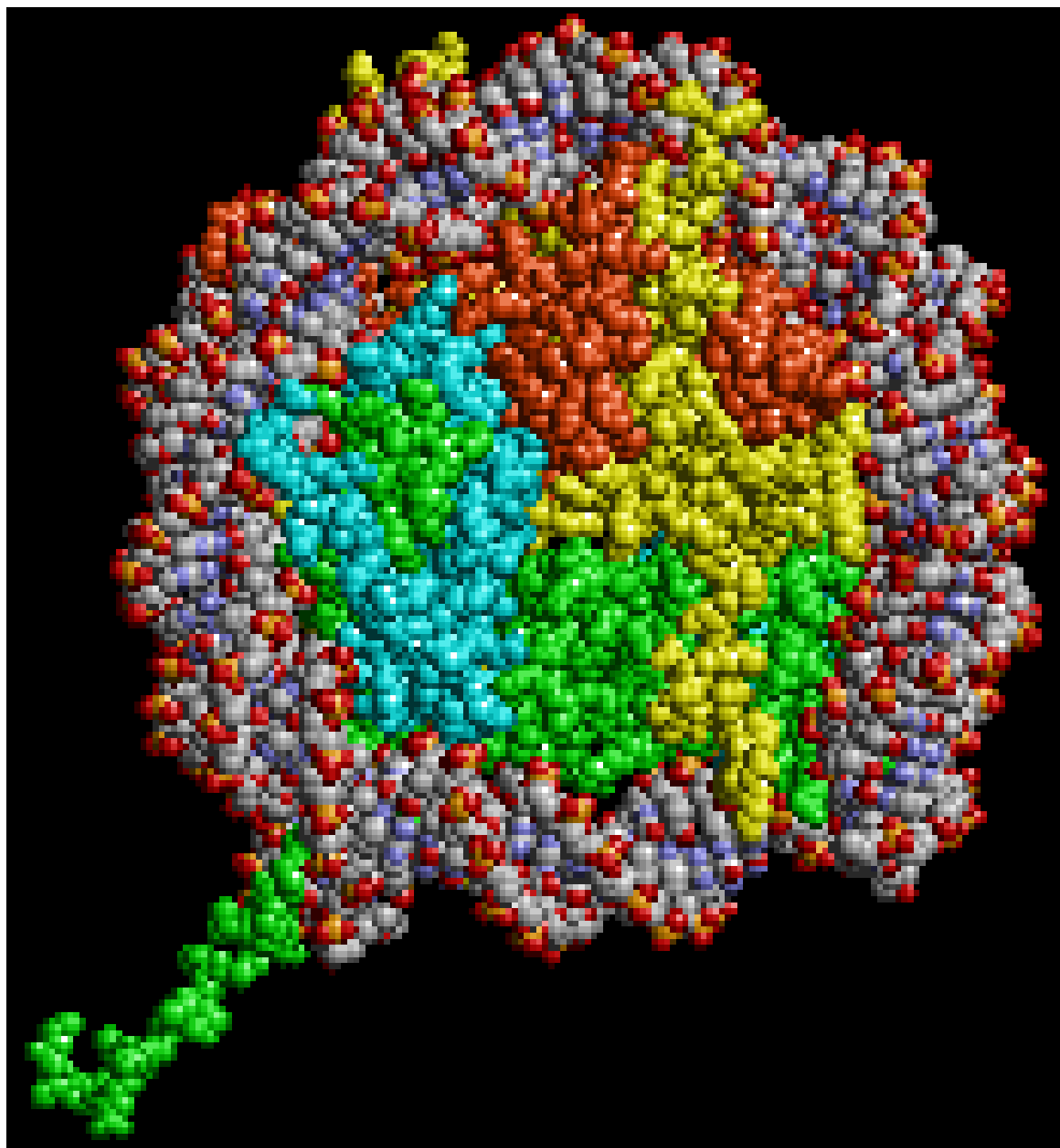


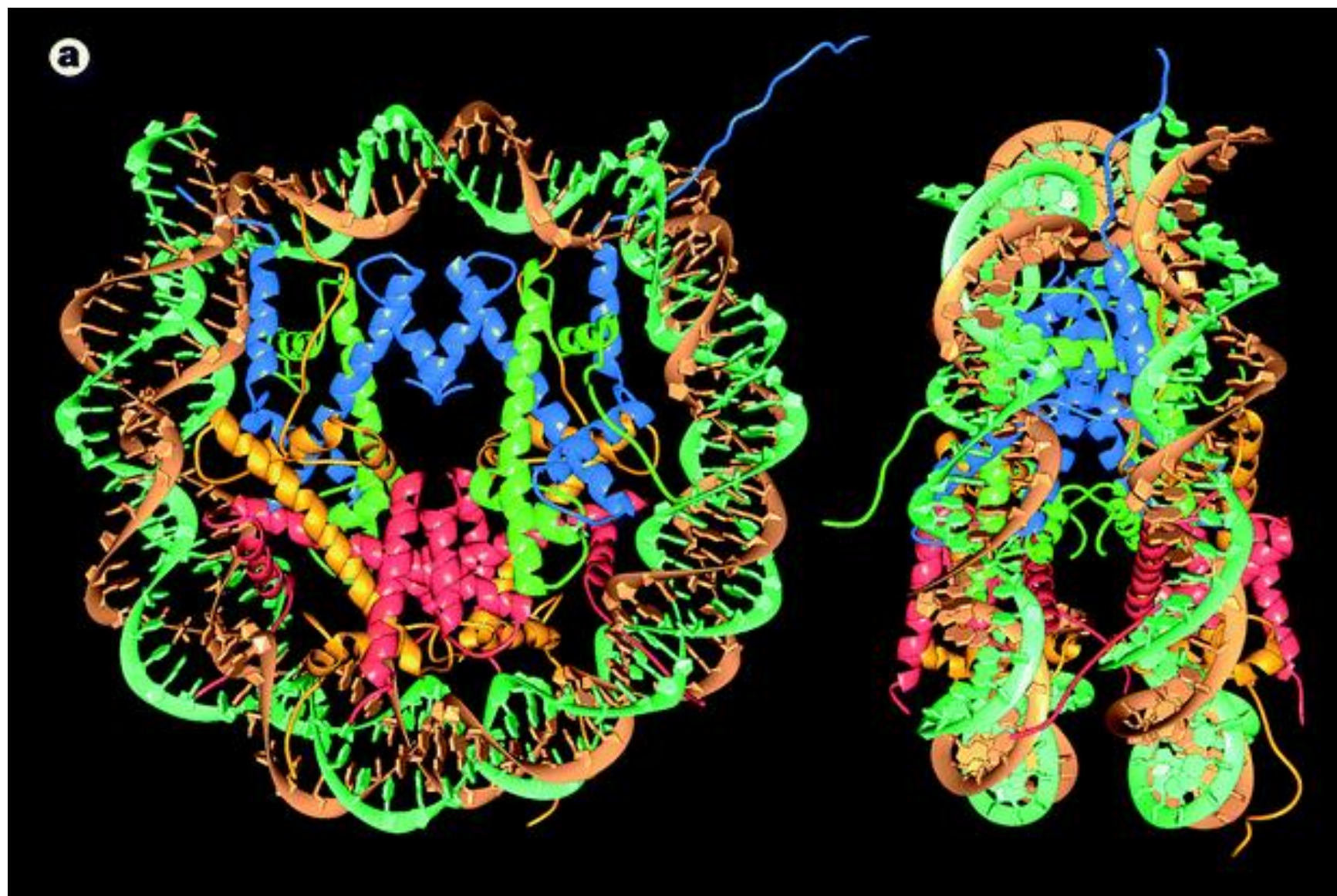
nucleosome core particle



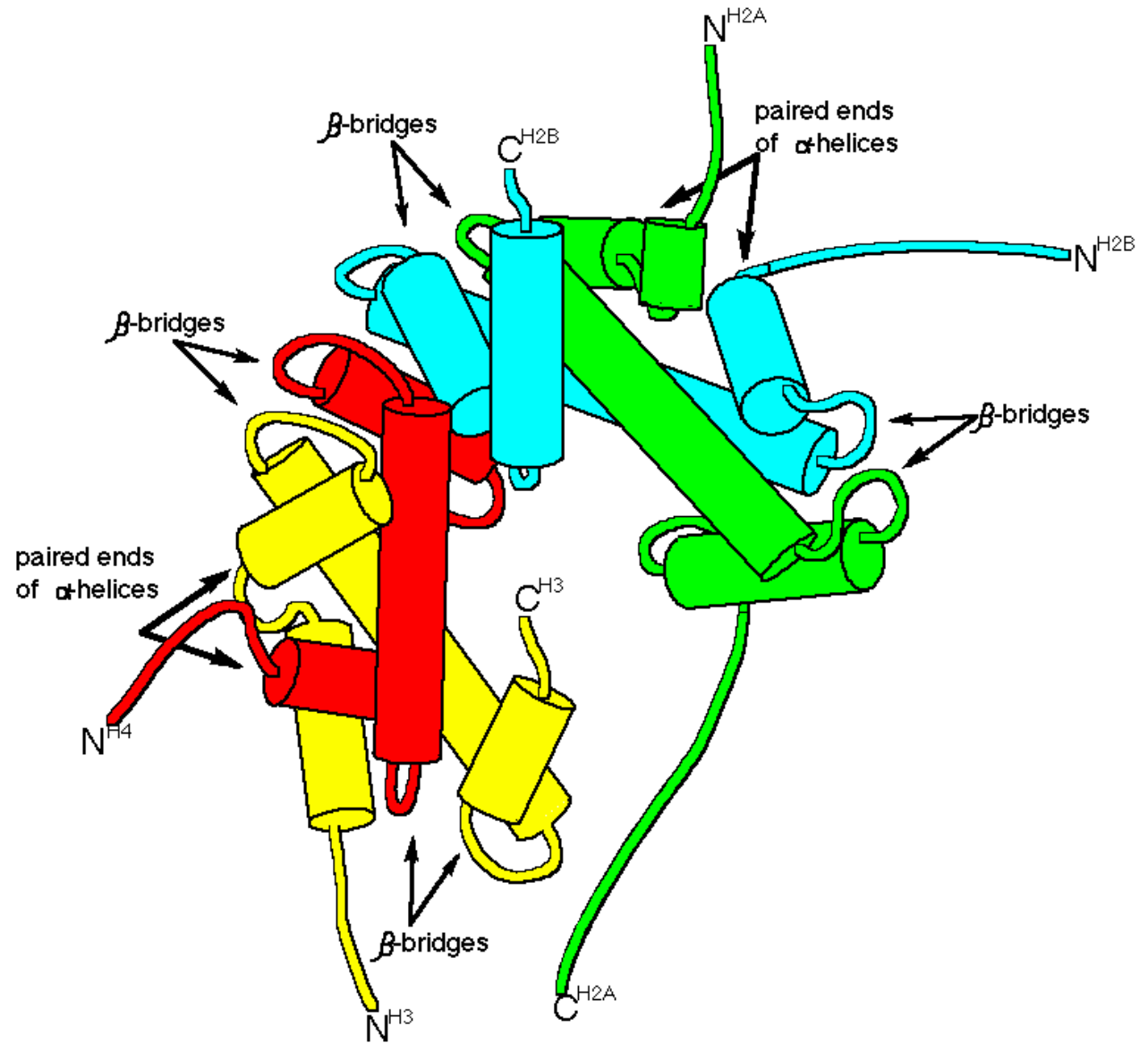
Luger, Mäder, Richmond, Sargent & Richmond, 1997

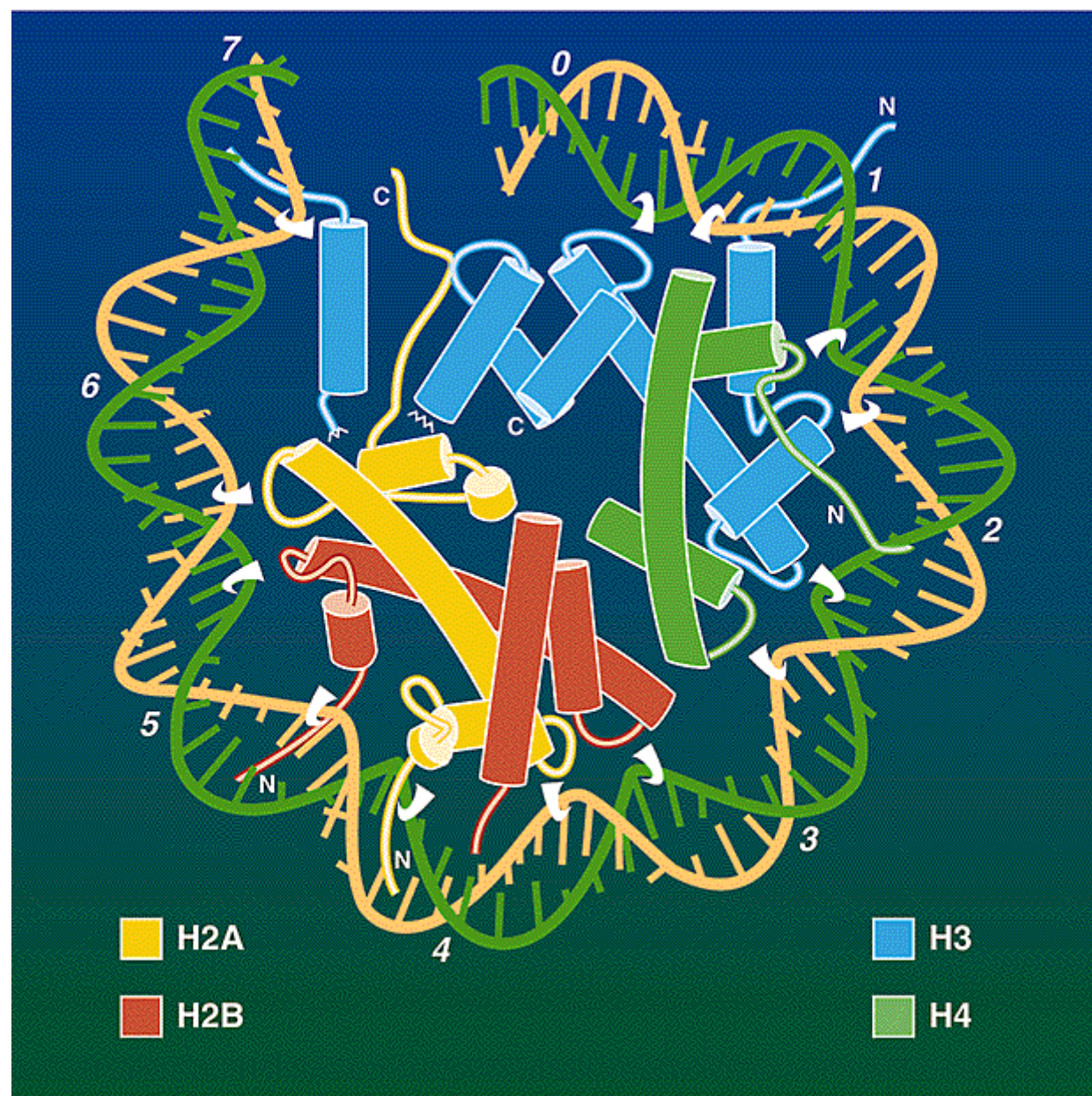




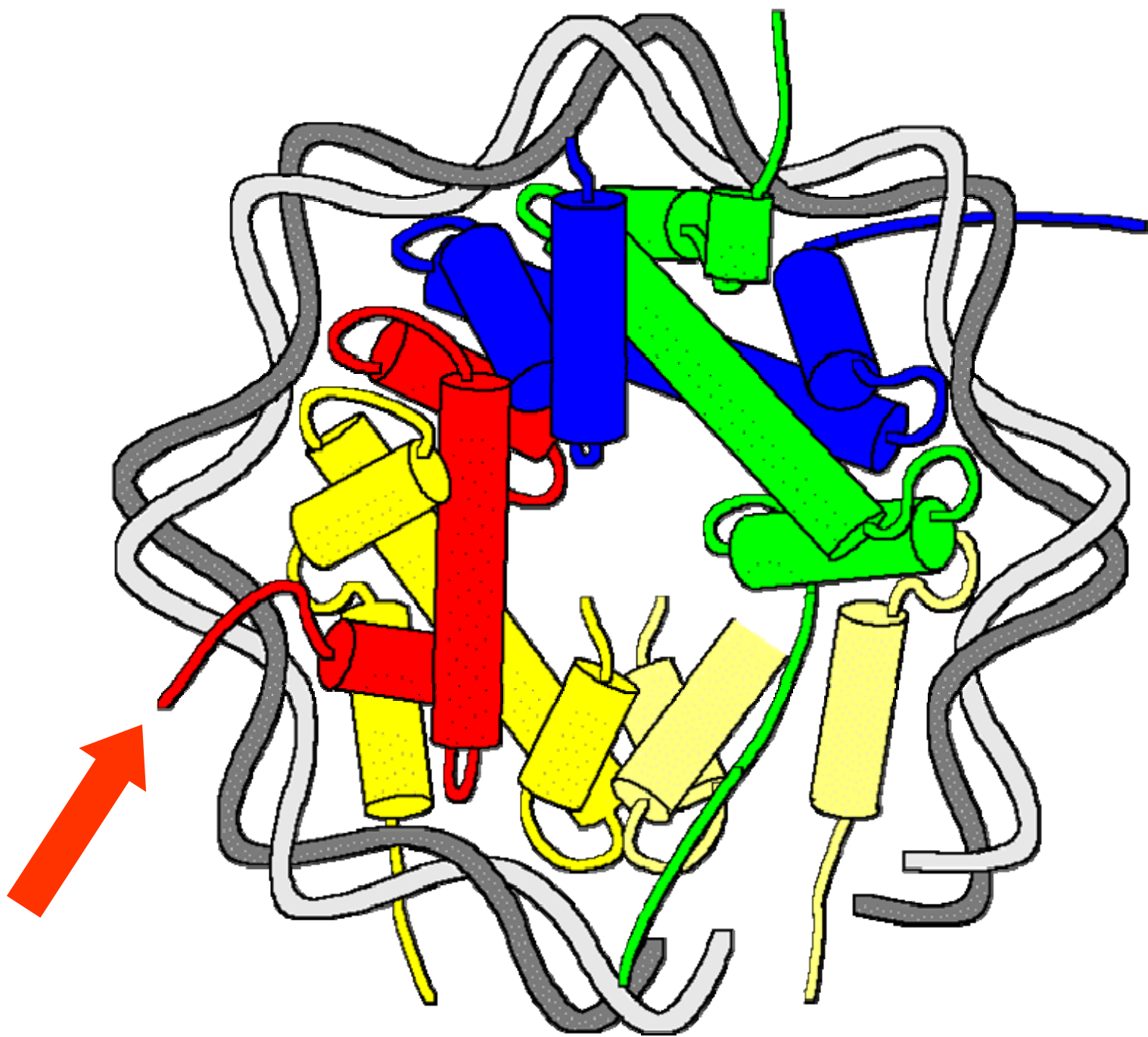


Histonoktamer
die Abbildung
zeigt nur ein
„halbes“
Oktamer



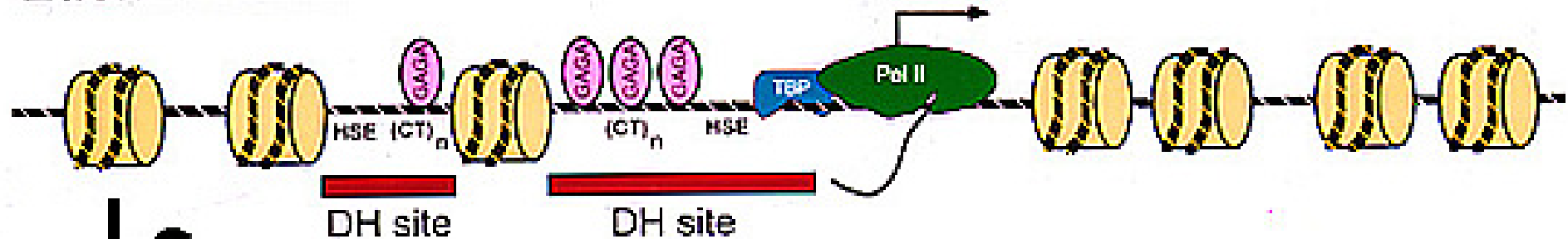




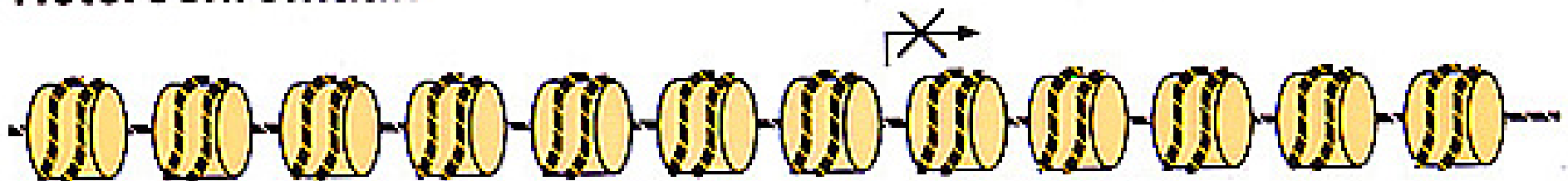


Chromatin und Genaktivität

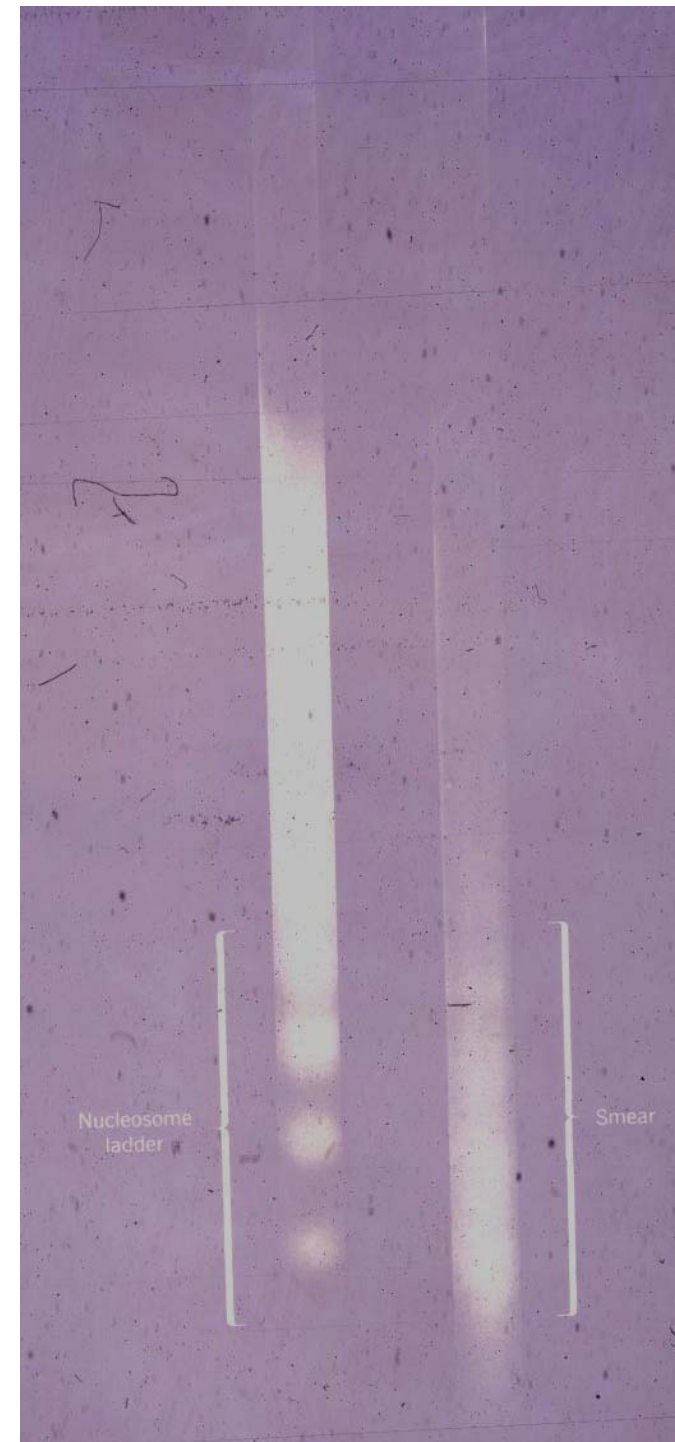
Euchromatin



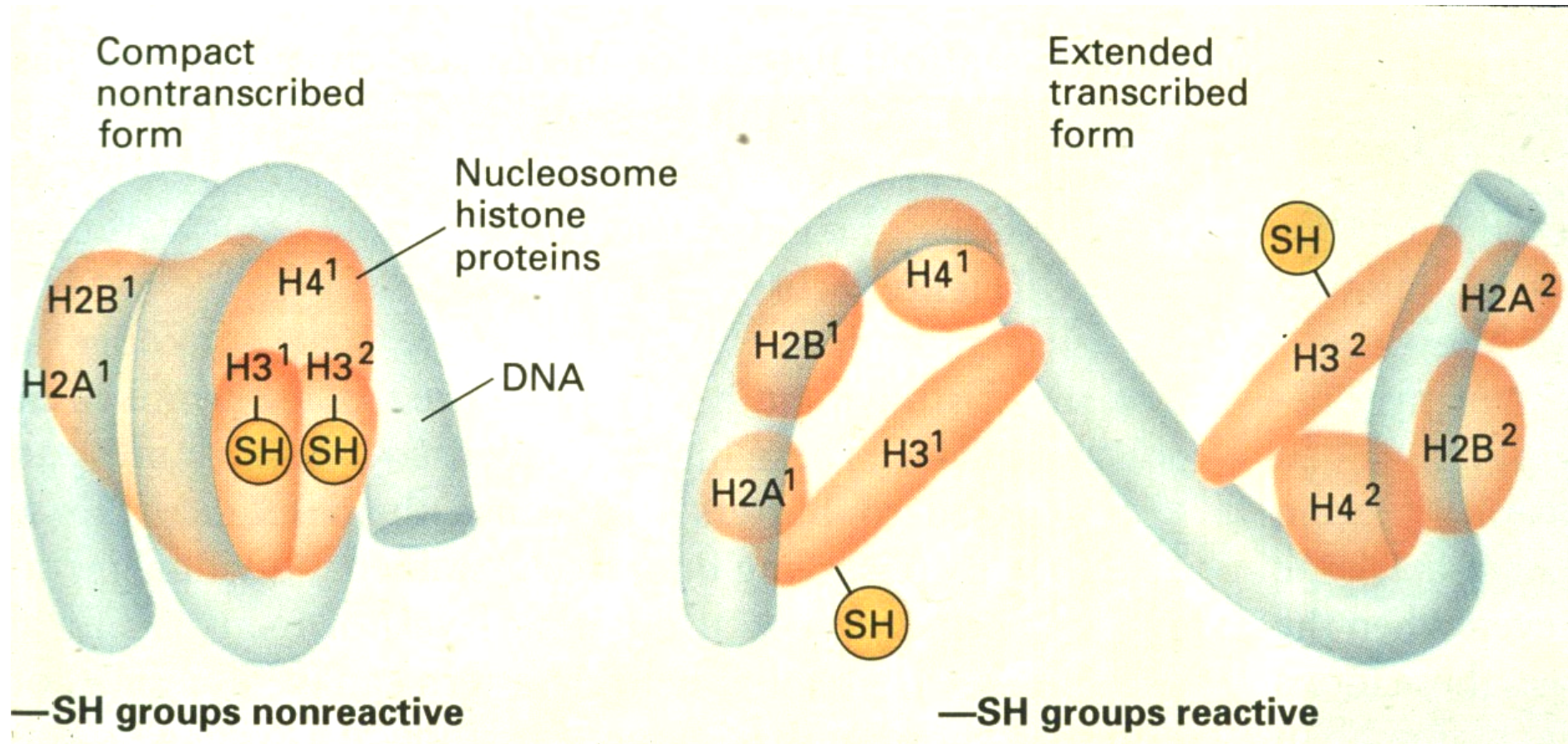
Heterochromatin

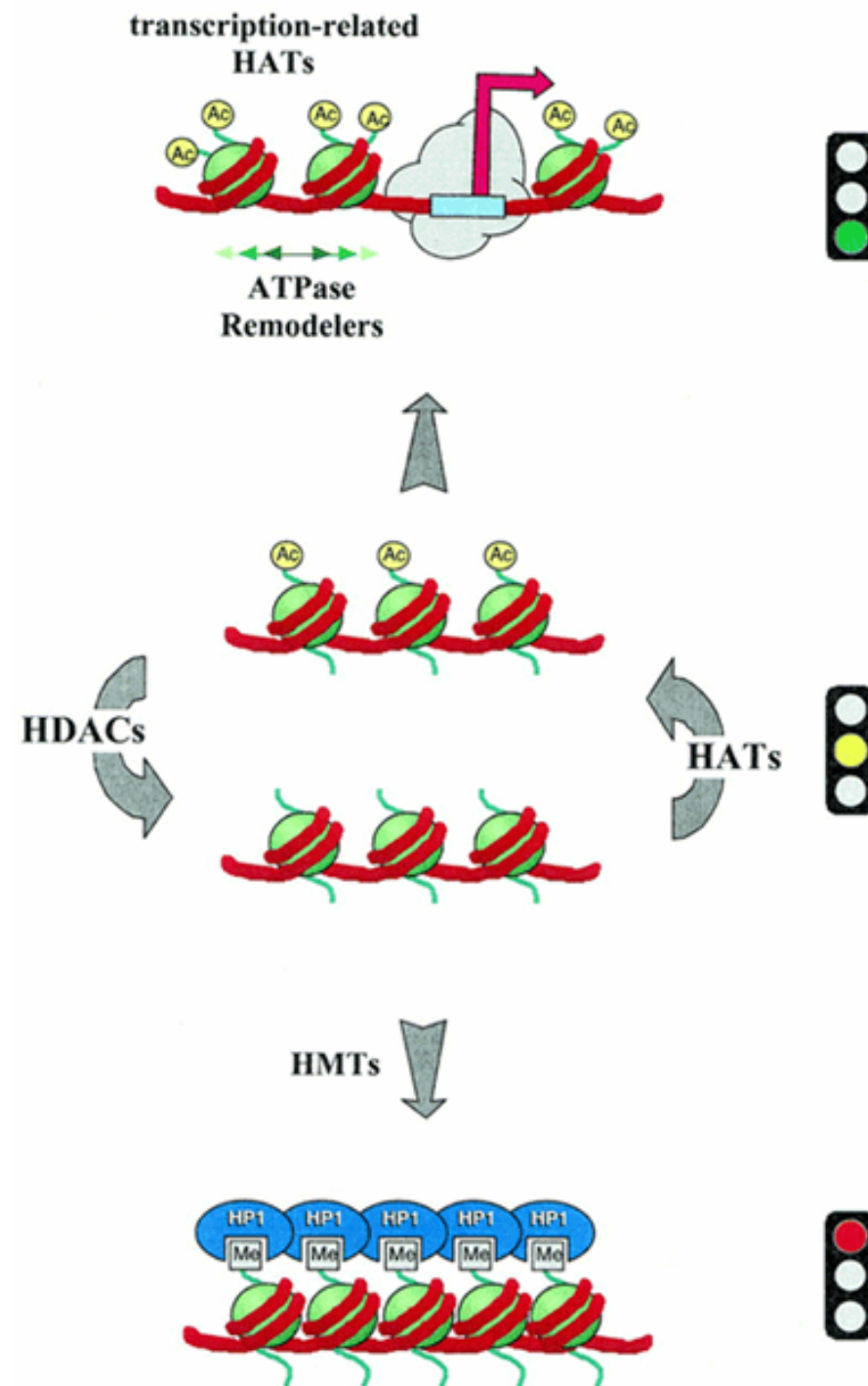


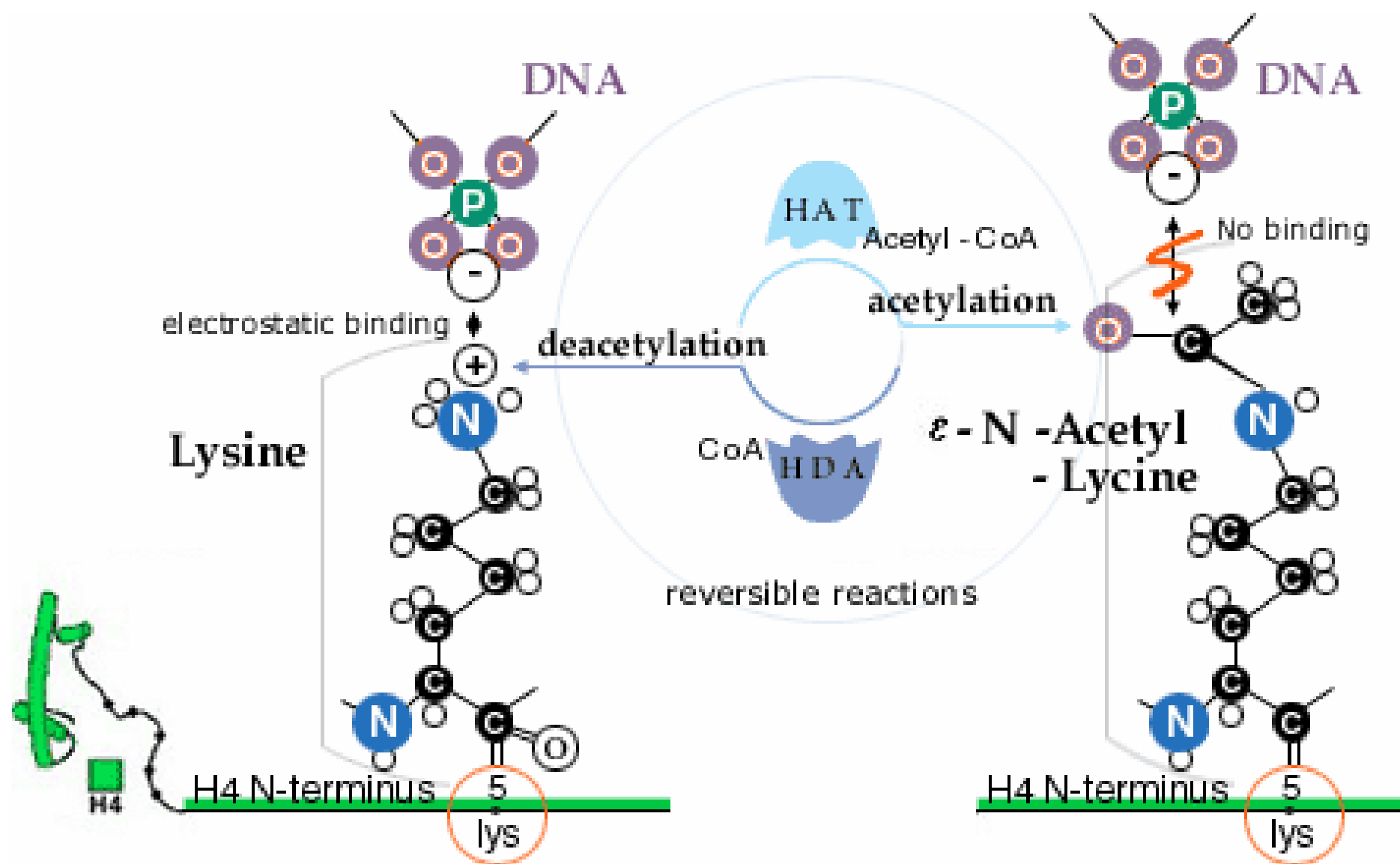
Aktives
Chromatin ist
nicht in
Nukleosomen
organisiert



Inaktive / Aktives Chromatin

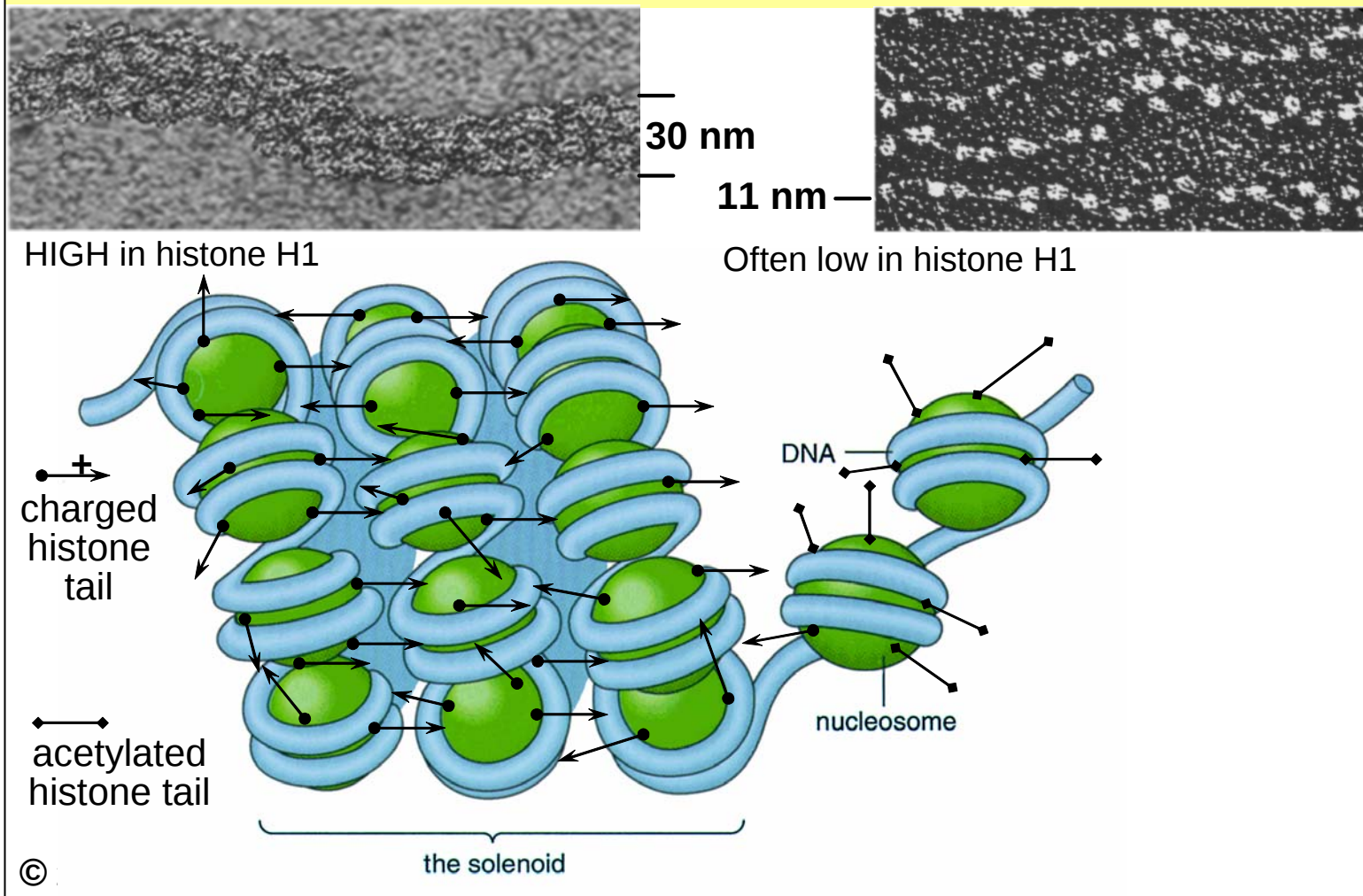






Histon Acetylierung

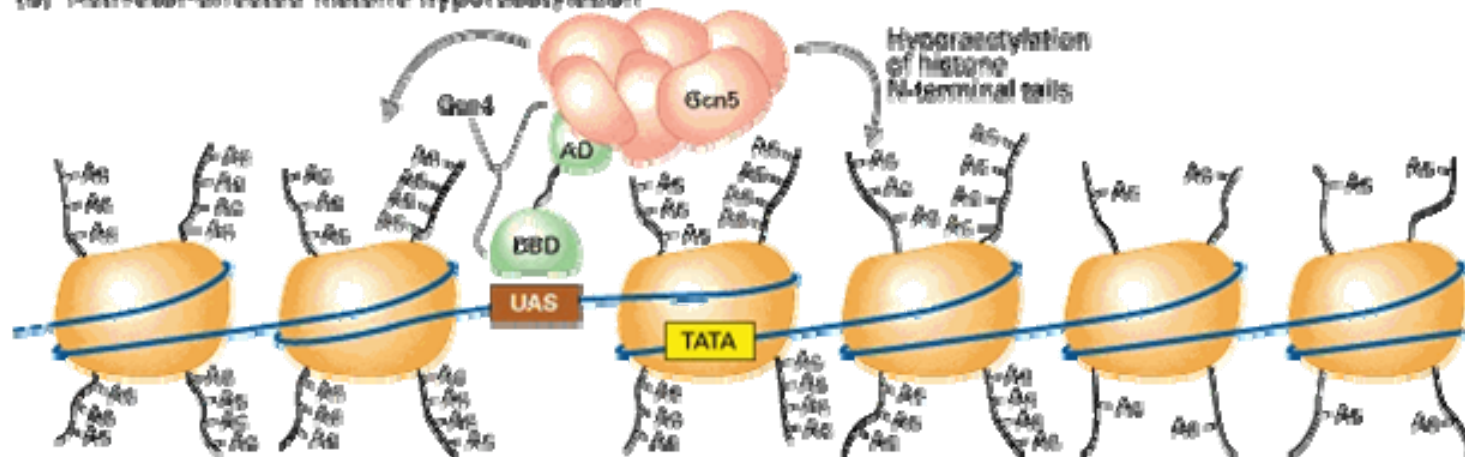
Function of Histone Acetylation



Histone acetylation and gene activation

Model for how HATs might activate gene expression in chromatin

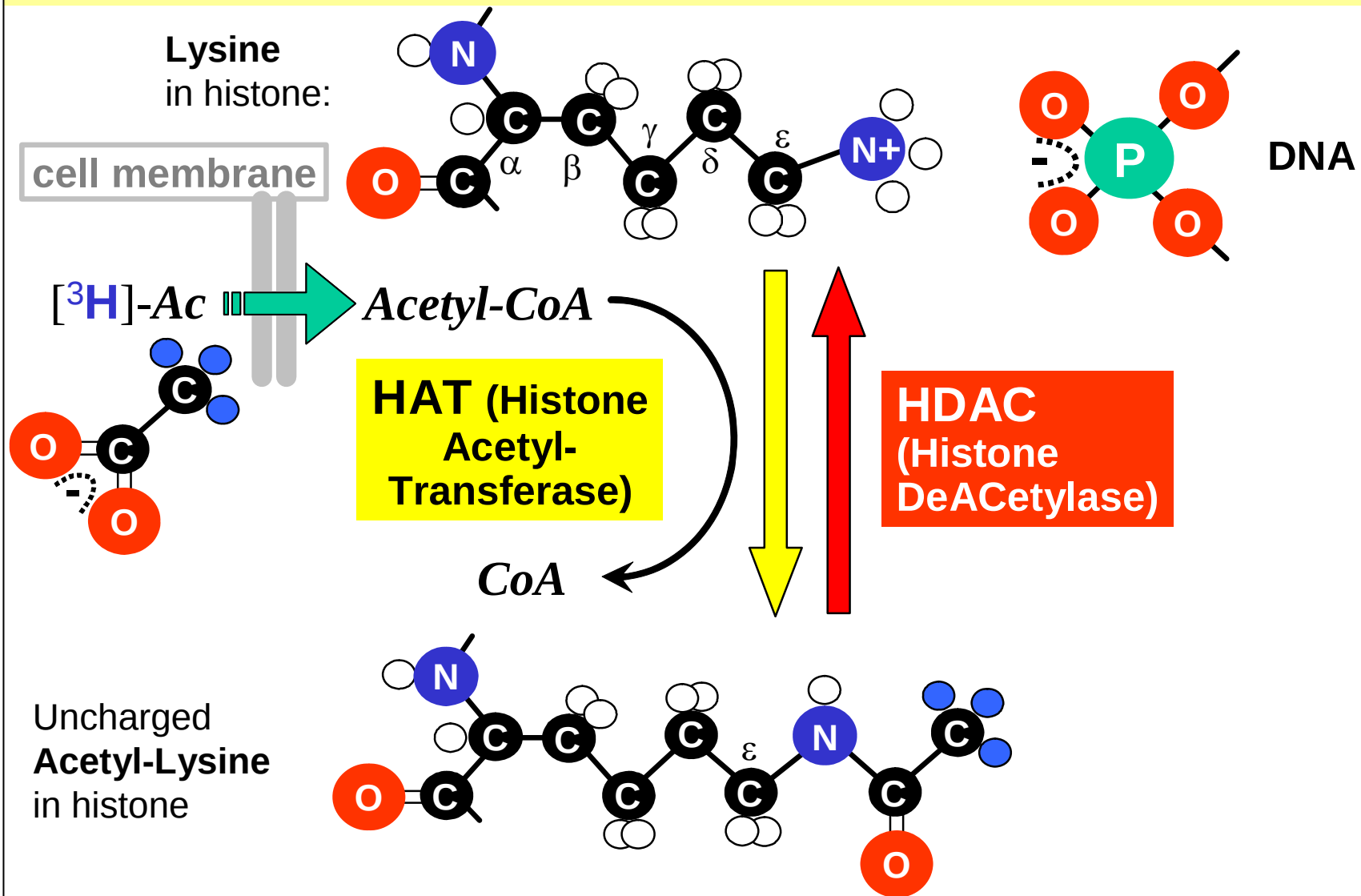
(b) Activator-directed histone hyperacetylation



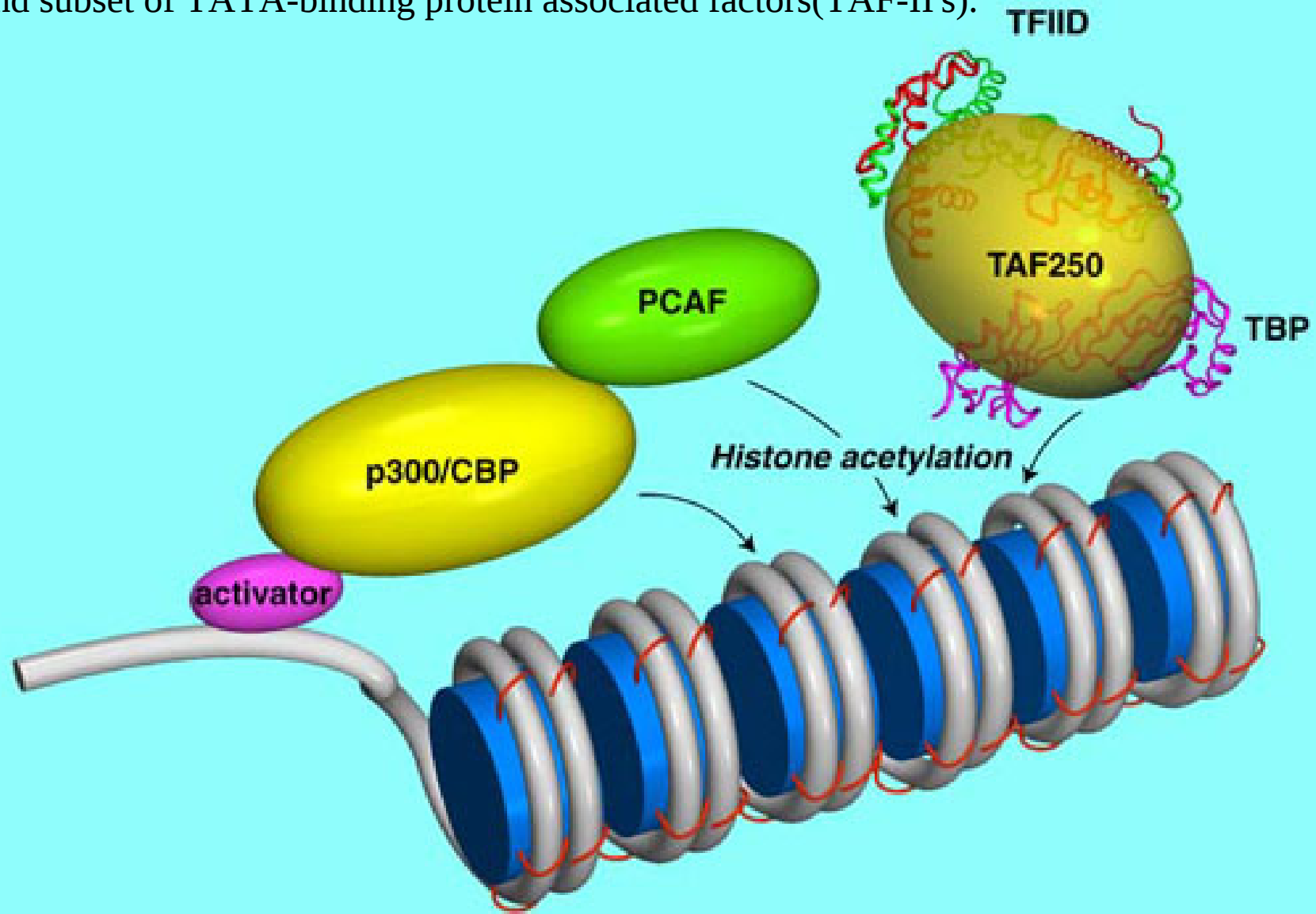
- GCN5, a known co-activator of gene expression in yeast, was shown to be closely related to the first HAT isolated biochemically from human cells
- GCN 5 was shown to be part of a large multi-protein complex
- One of the subunits of the complex was shown to interact directly with GCN4
- GCN4 is a gene-specific TF that binds to a specific DNA sequence in the promoter-proximal region of genes regulated by GCN4

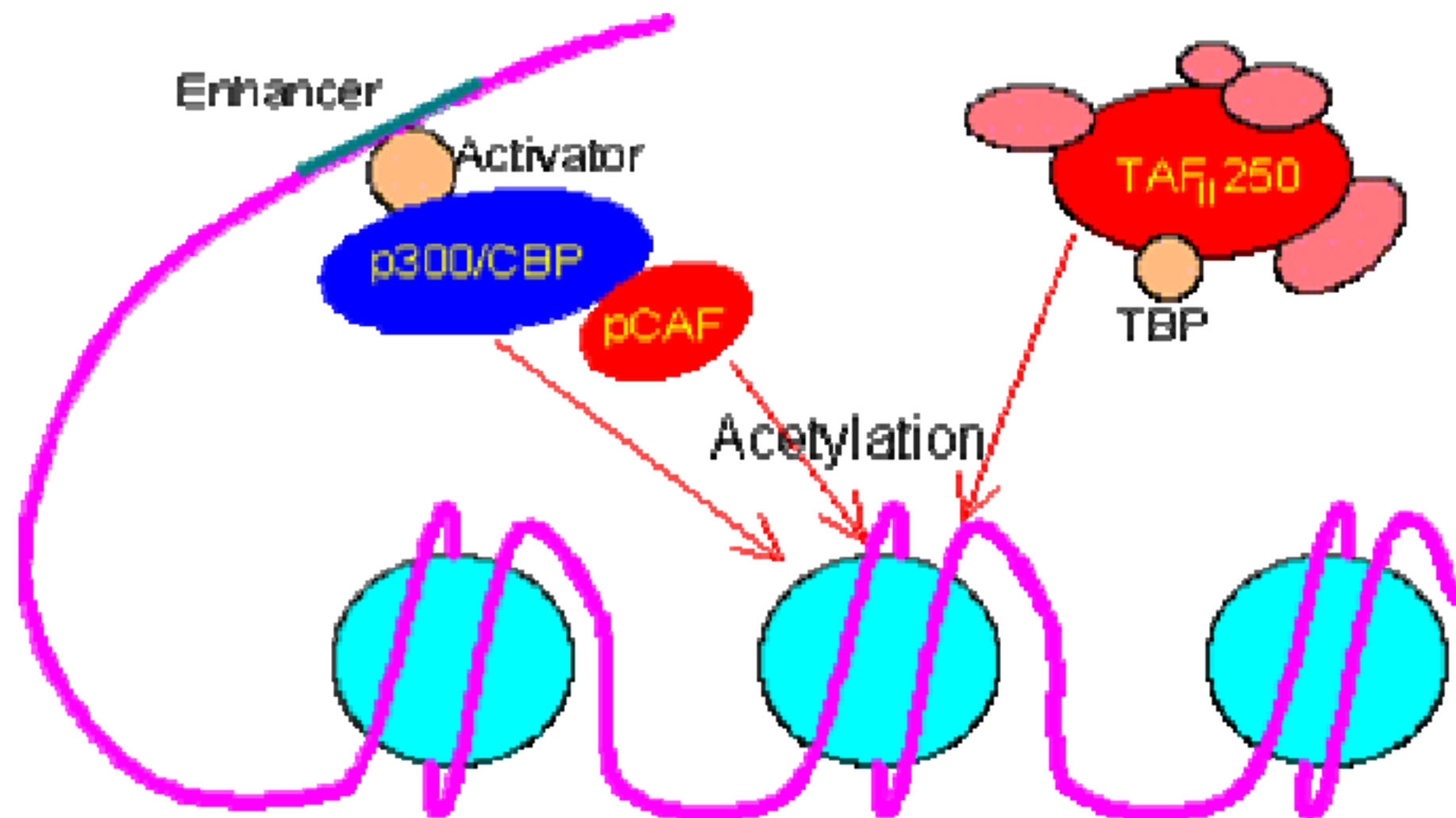
Slide 6 of 12

Histone Acetylation Enzymes



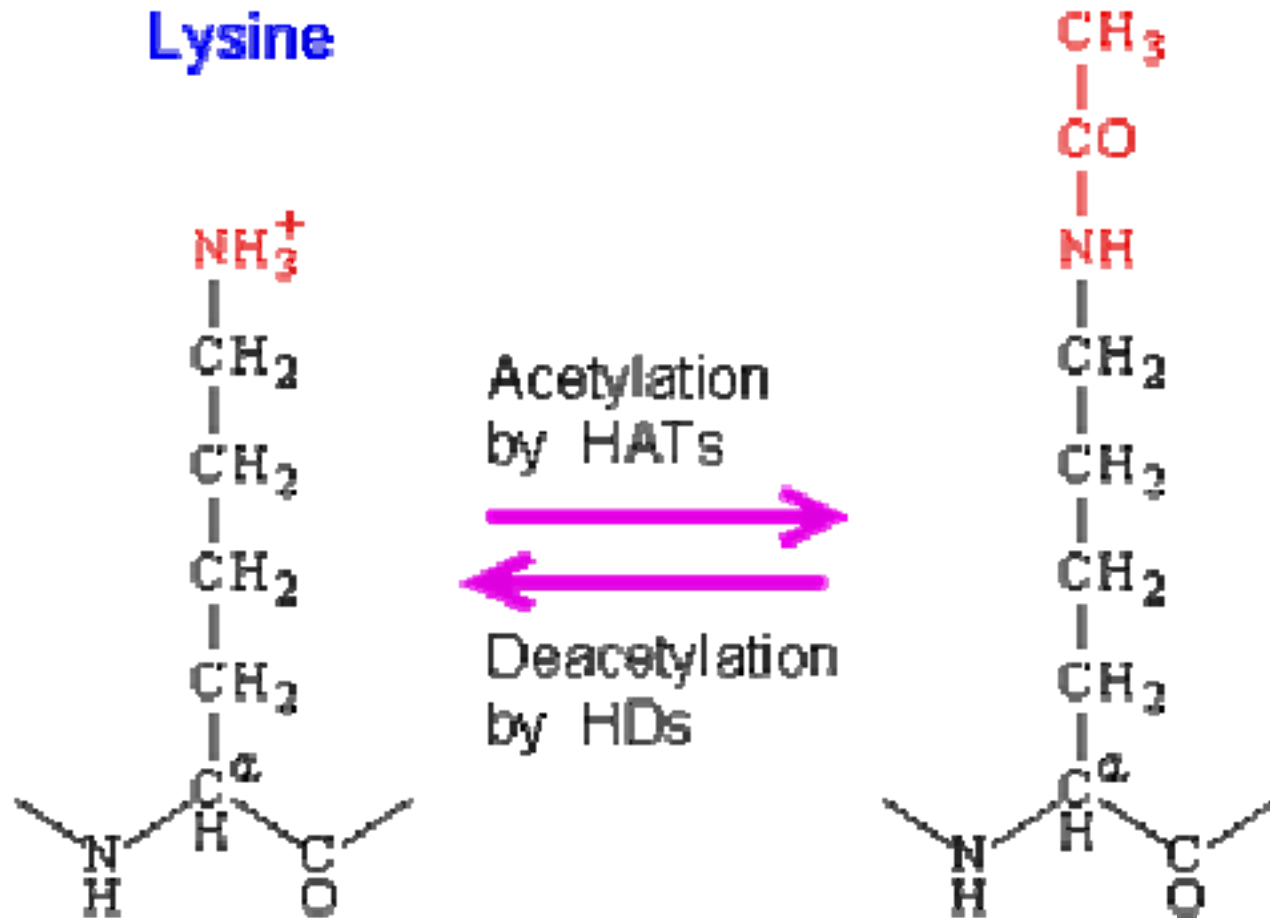
The Protein named PCAF contains both a histone acetyltransferase (HAT) and subset of TATA-binding protein associated factors(TAF-II's).





Acetylierung und Deacetylierung

Lysine



Acetylated Histones – a choice

N-terminus of histone H3

H₃N-ART⁺₅**K**QTAR⁺₉**K**S⁺

TGG⁺₁₄**K**APR⁺₁₈**K**QL⁺

AS⁺₂₃**K**AAR⁺₂₇**K**SAP...⁺

3-6 Ac

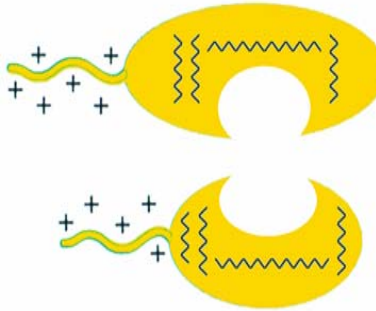
3-6 Ac

H₃N-ART⁺**K**⁺QTAR⁺**K**⁺S
5 9
m m

TGG⁺**K**APR⁺**K**QL⁺
14 18

ASK⁺₂₃AAR⁺₂₇K⁺SAP...

H3 : 3-6Ac



H2B : 3-6Ac

H4 : 4 or 5 Ac

N-terminus of histone H4

Ac-HN-SGRG⁺**K**⁺GG⁺**K**⁺GL G⁺**K**⁺GGAK⁺⁺RR⁺**K**⁺ILR...
5 8 12 16 20 (m)

4 or 5 Ac

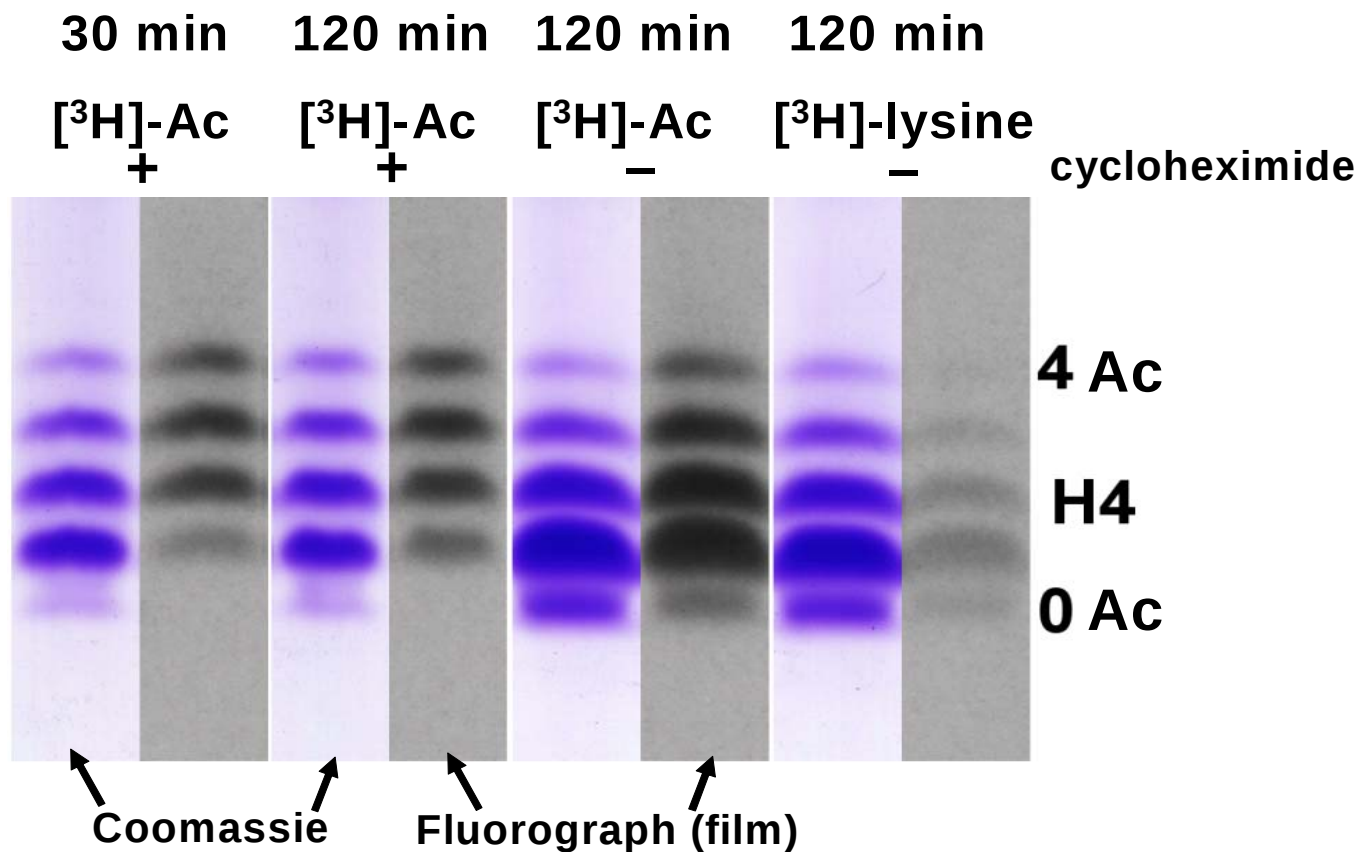
Ac-HN-SGRG⁺₅K⁺G⁺G⁺K⁺GL G⁺₁₂K⁺G⁺G⁺A⁺K⁺₁₆R⁺H⁺R⁺K⁺₂₀ILR...

4 or 5 Ac

Note the “standard” colors for the core histones

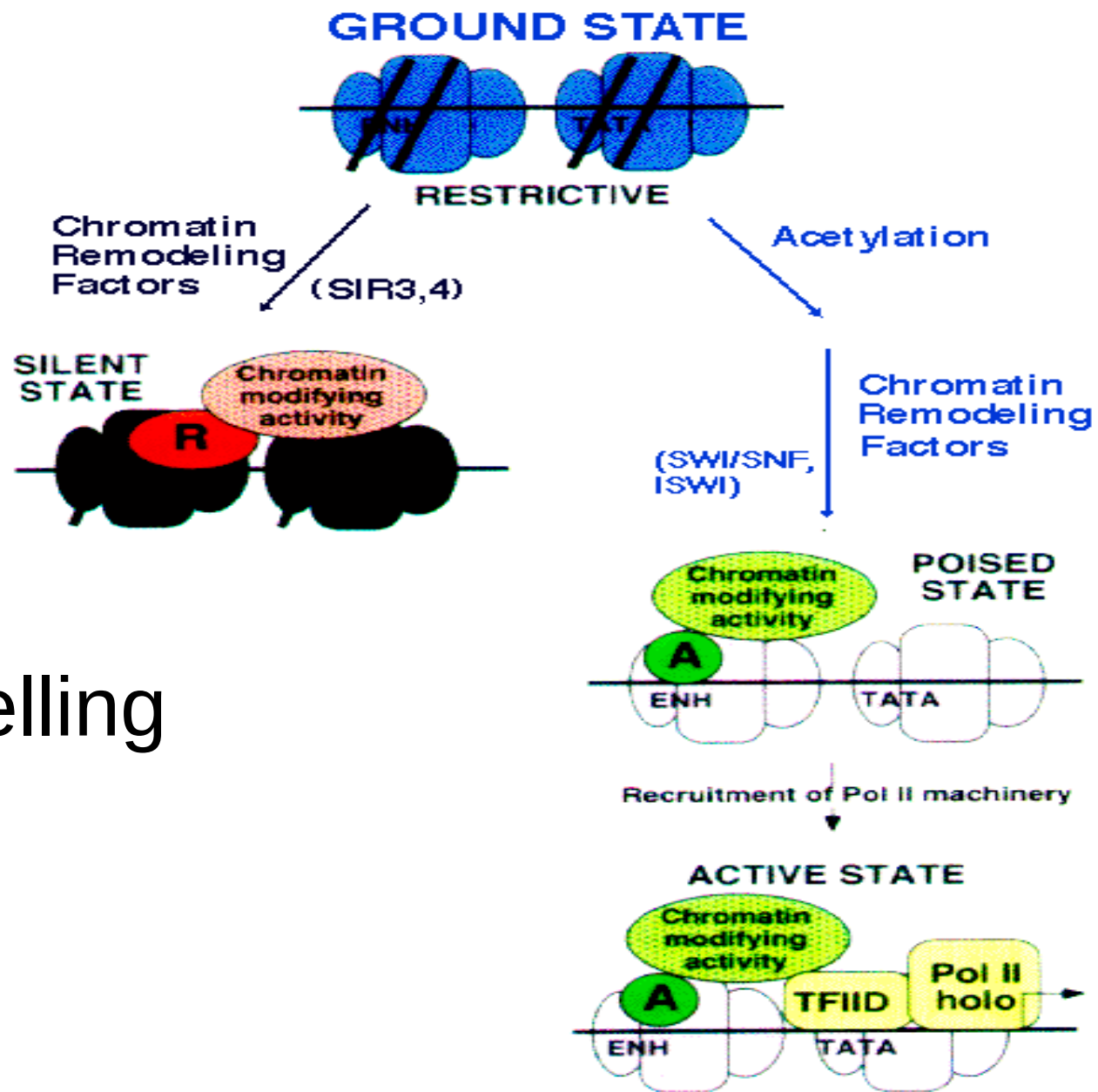
H4-Acetylierung

Acetylation of Yeast histone H4

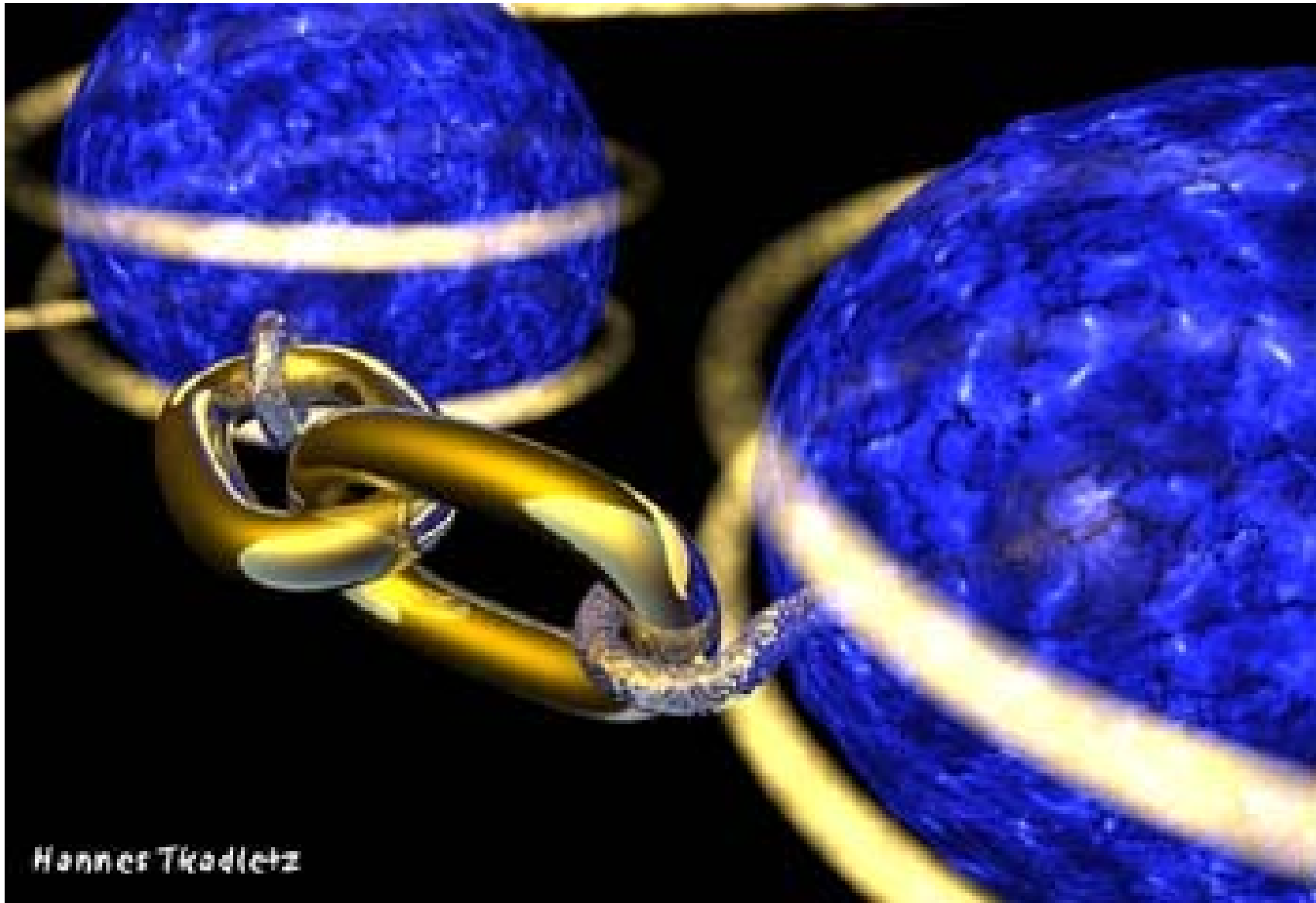


„Remodelling“

“



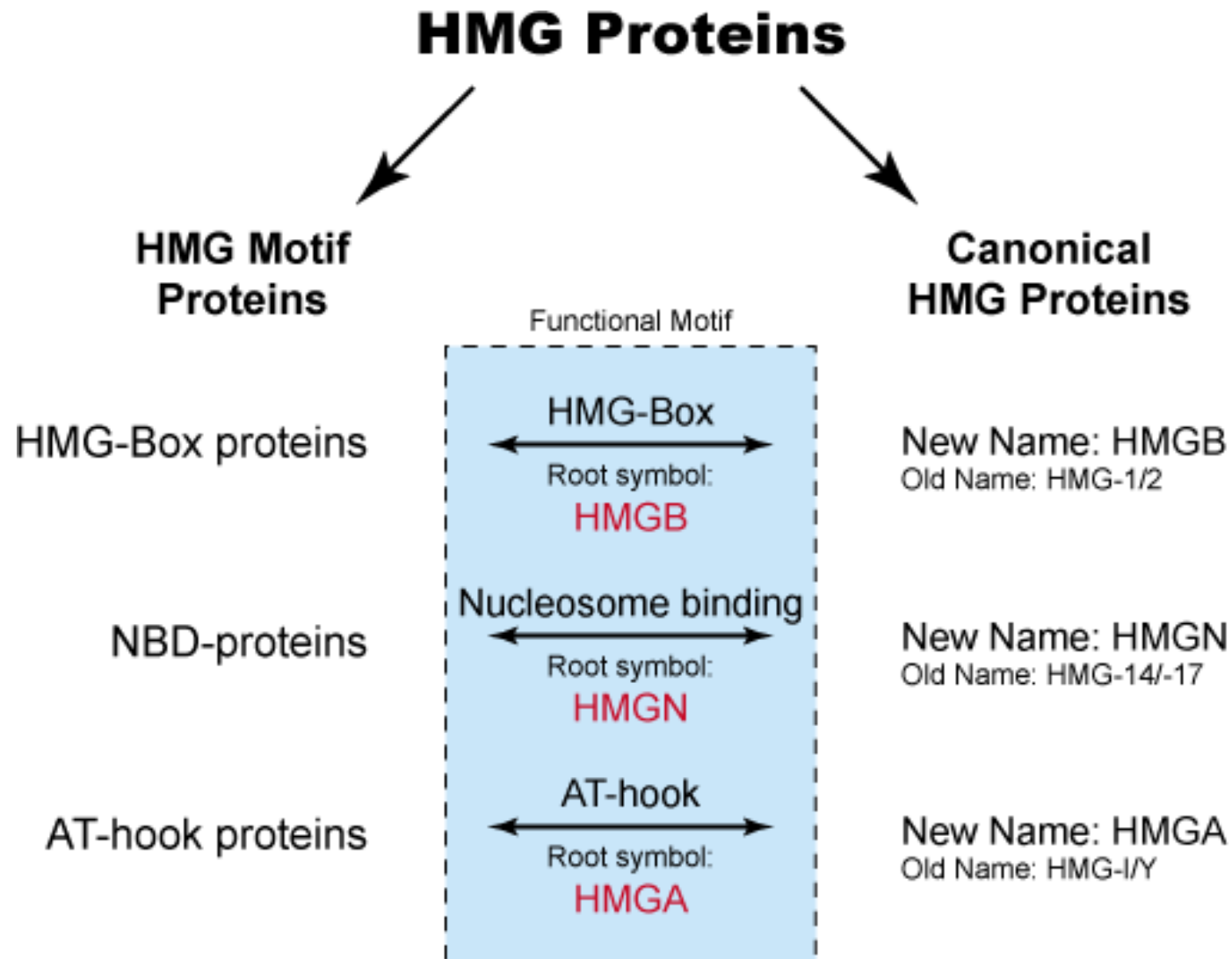
Inaktivierung durch Histonmethylierung



Weiere Modifikationen an Chromatin:

Core histones can be reversibly modified
by acetylation,
methylation, phosphorylation,
ubiquitination or ADP-ribosylation
(Strahl and Allis, 2000)

Weitere Chromatinproteine



Nukleosomenstruktur des Chromatins

informative Internetadressen:

<http://www.average.org/~pruss/nucleosome.html>

http://bama.ua.edu/~hsmithso/class/bsc_495/chromatin/chromatin_web.html

http://www.nature.com/cgi-taf/DynaPage.taf?file=/nature/journal/v389/n6648/full/389231a0_fs.html

http://www.biochem.arizona.edu/classes/bioc462/462a/NOTES/Nucleic_Acids/nucleosome.html

<http://info.bio.cmu.edu/Courses/03438/Nsome/1AOI.htm>

<http://arapaho.nsuok.edu/%7Ebiology/Tutorials/Nucleosome.htm>

<http://medweb.bham.ac.uk/research/chromatin/nucleosome.html>

<http://www.sciencemag.org/cgi/content/abstract/1060118v1>

Chromatin structure: The nucleosome core all wrapped up Histonmethylierung:

Nature **389**, 231 - 232 (1997)

X-Inaktivierung

Barr body in epidermal spinous cell layer

Nuclear appendage in white blood cells

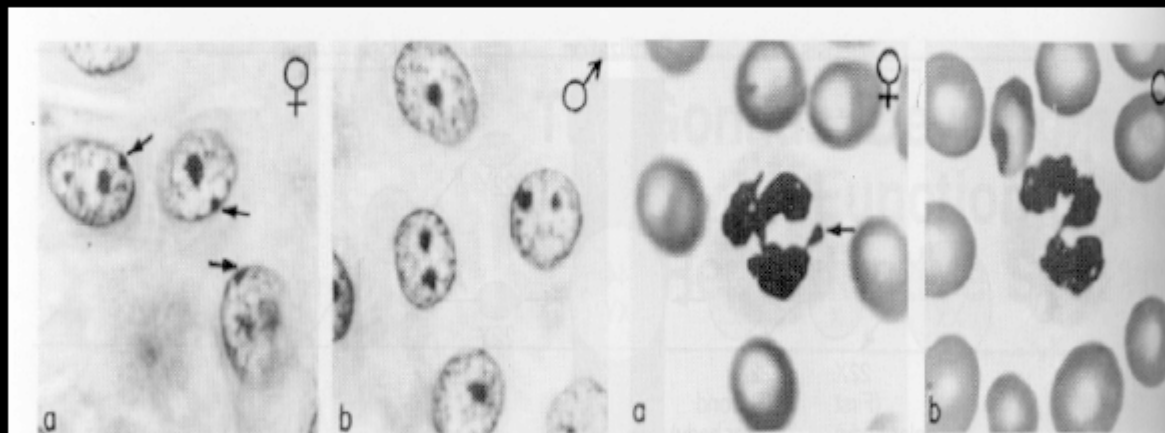


Figure 23-3. *Left:* Barr body (arrows) in the epidermal spinous cell layer. *Right:* Nuclear appendage ("drumstick") identified by arrow in white blood cells. (Reproduced, with permission, from Grumbach MM, Barr ML: Cytologic tests of chromosomal sex in relation to sex anomalies in man. *Recent Prog Horm Res* 1958;**14**:255.)

Mechanism of X-inactivation

Inactivation of an X chromosome requires a gene on that chromosome called *XIST*.

XIST encodes a large molecule of **RNA** (of a type different from those, e.g., mRNA, used in protein synthesis).

XIST RNA accumulates along the X chromosome containing the active *XIST* gene and proceeds to inactivate all (or almost all) of the other hundreds of genes on that chromosome.

XIST RNA does not travel over to any other X chromosome in the nucleus.

Barr bodies are inactive X chromosomes "painted" with *XIST* RNA.

During the first steps of embryonic development of the female, the *XIST* locus on each of her two X chromosomes is expressed but the *XIST* RNA is quickly broken down. Then something happens to tip the balance in favor of one or the other of the X chromosomes.

[transcription](#) **continues** on one of the X chromosomes, leading to an accumulation of *XIST* RNA and converting that chromosome into an inactive Barr body.

transcription of *XIST* **ceases** on the other X chromosome allowing all of its hundreds of other genes to be expressed. The shut-down of the *XIST* locus on the active X chromosome is done by methylating *XIST* regulatory sequences. [DNA methylation](#) usually results in gene repression so methylation permanently blocks *XIST* expression and permits the continued expression of all the other X-linked genes.

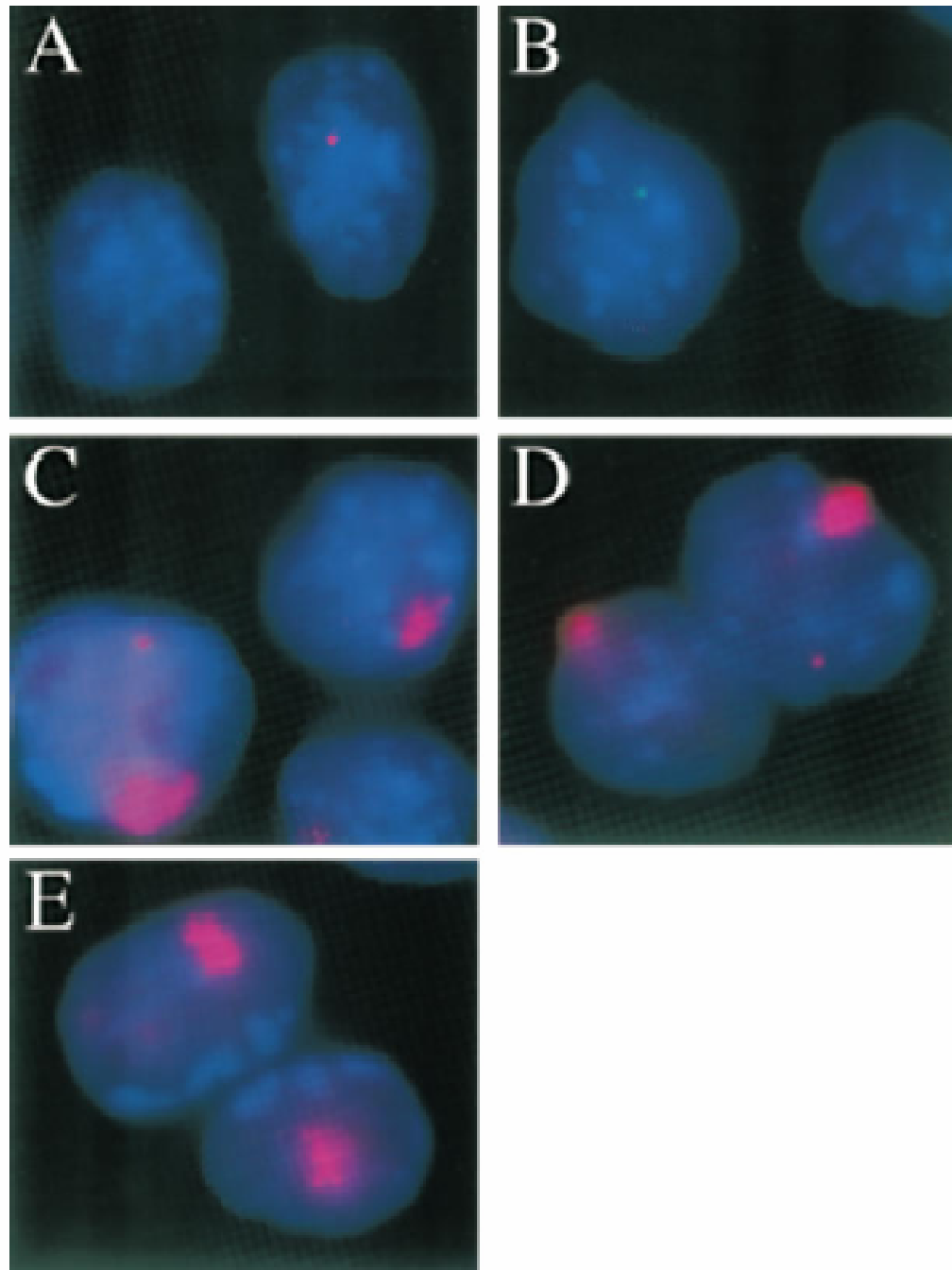
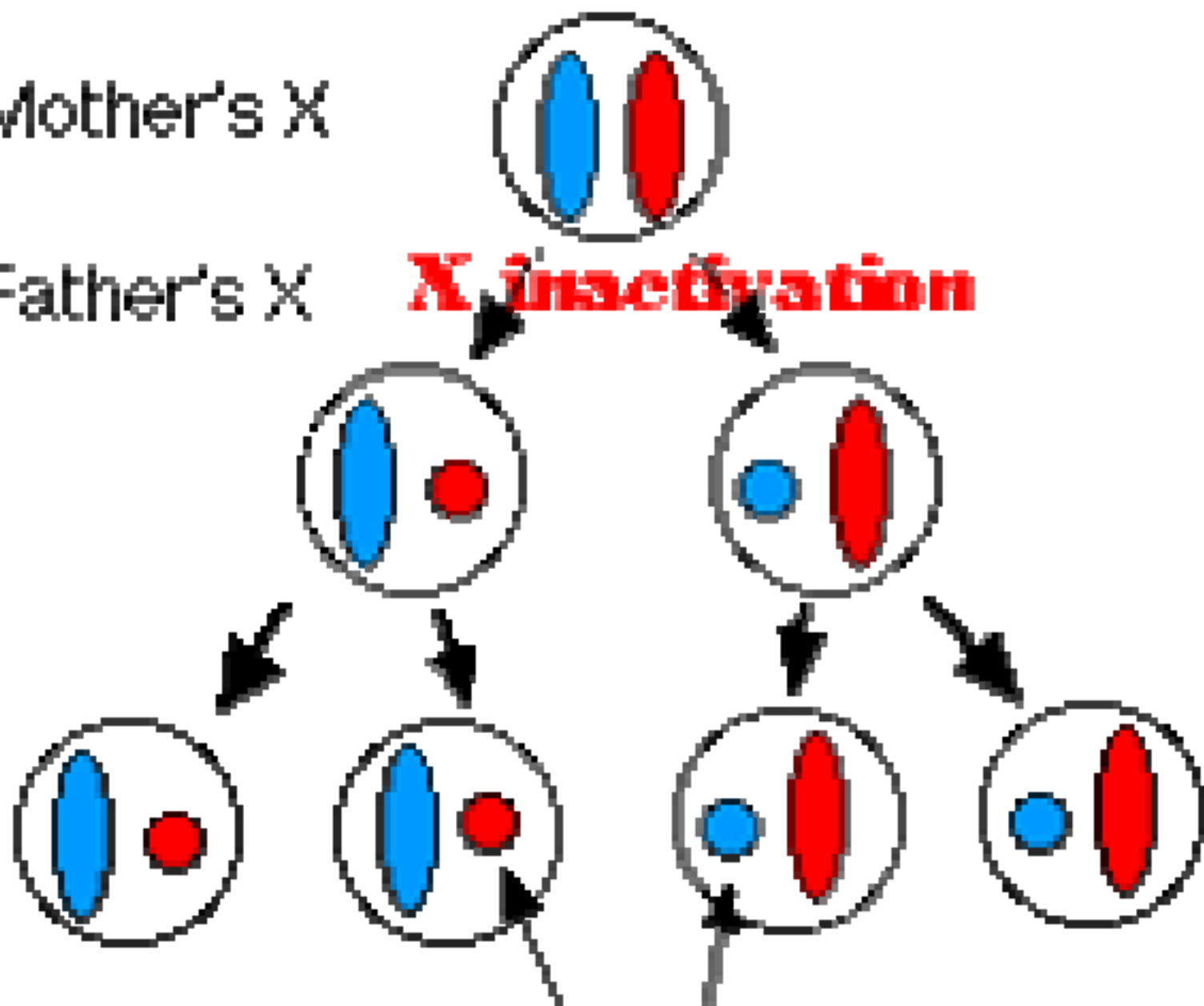


Figure 4. *Xist* Expression in Cells From Female *Xist*^{+/-} Embryos at 7.5 Days of Gestation
Fluorescence in situ hybridization was performed for *Xist* (pink/red) and *Pgk-1* (green) RNA. The full probe was used in (A), (C), and (D); the deletion probe was used in (B) and (E). A probe for *Pgk-1* was also used in (B). (A) *Xist*⁻ male; (B) *Xist*⁻ male; (C) *Xist*^{+/+} female; (D) *Xist*^{+/-} female; (E) *Xist*^{+/-} female.

 = Mother's X

 = Father's X

X inactivation



Barr bodies

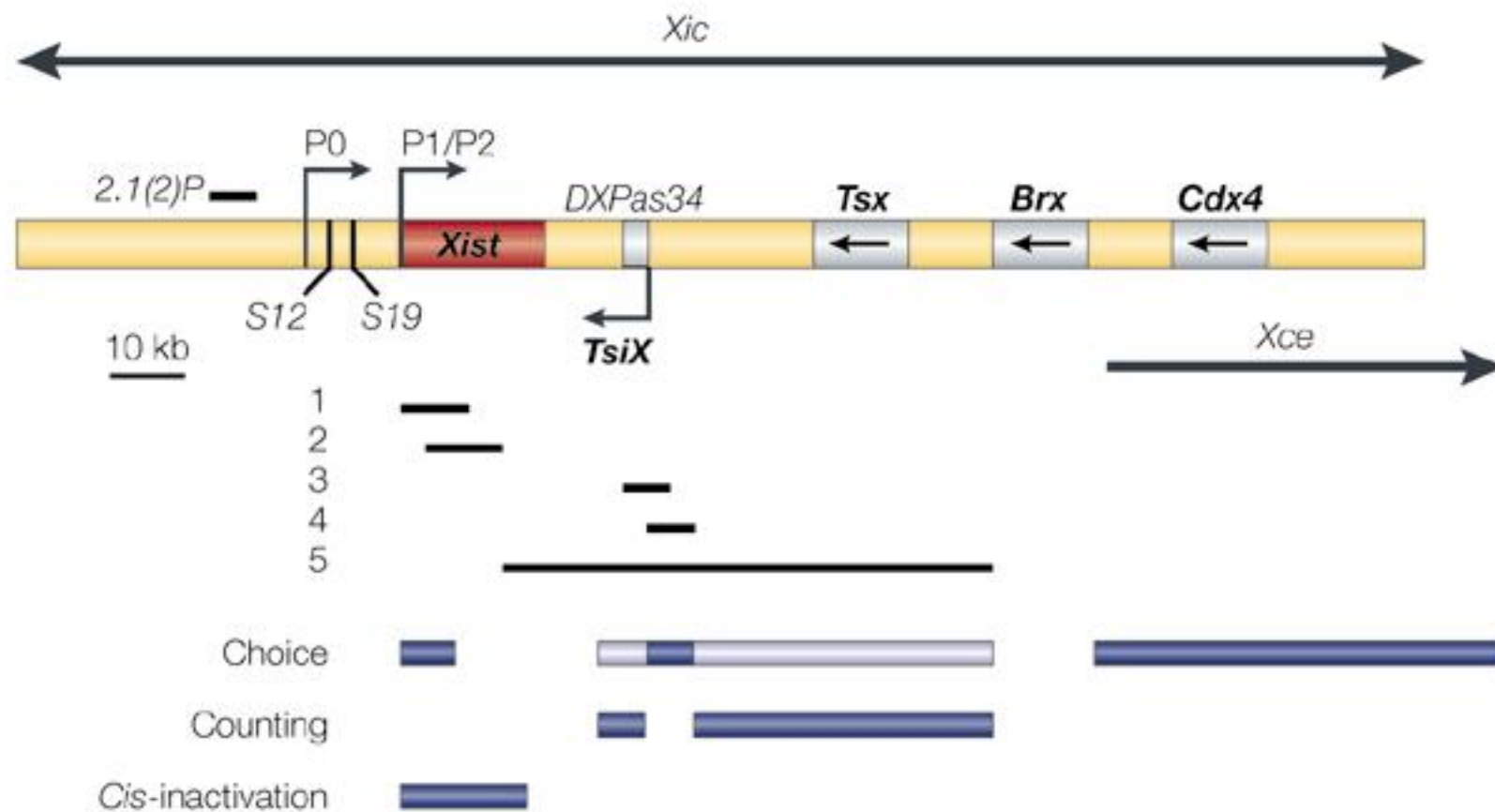
Because of X inactivation, female mammals are mosaic

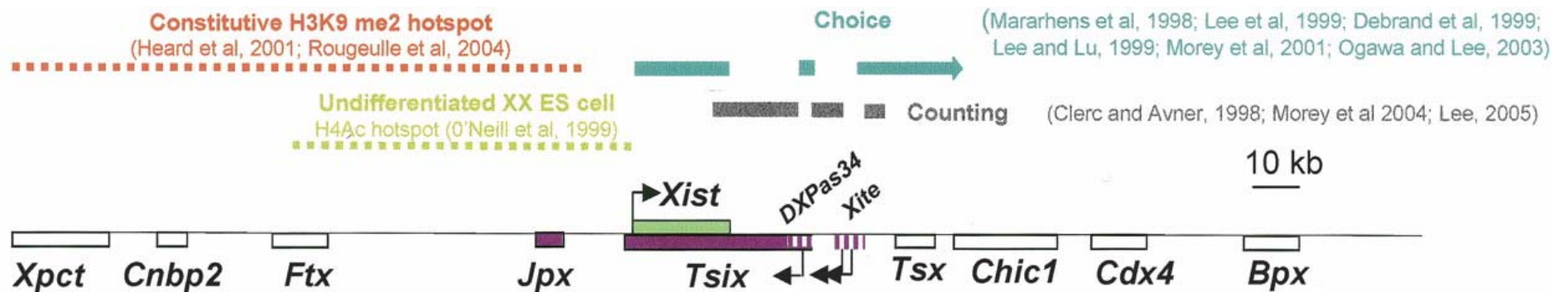


XIC

- signal from X inactivation center, spread along chromosome, promoted by booster elements (Fig 3)
 - signal spreading is less effective in autosomal material than in the X chromosome itself
 - LINEs may represent booster sequences
 - » X chromosome is strikingly LINE-rich
 - » Efficiency of spread into autosomal regions correlates with their LINE content
- inactive X replicates late, the late replication may help maintain the inactive signal, in which late replication prevents transcription and transcription is required for early replication

The X inactivation center





450 kb YAC (Lee et al, 1996)

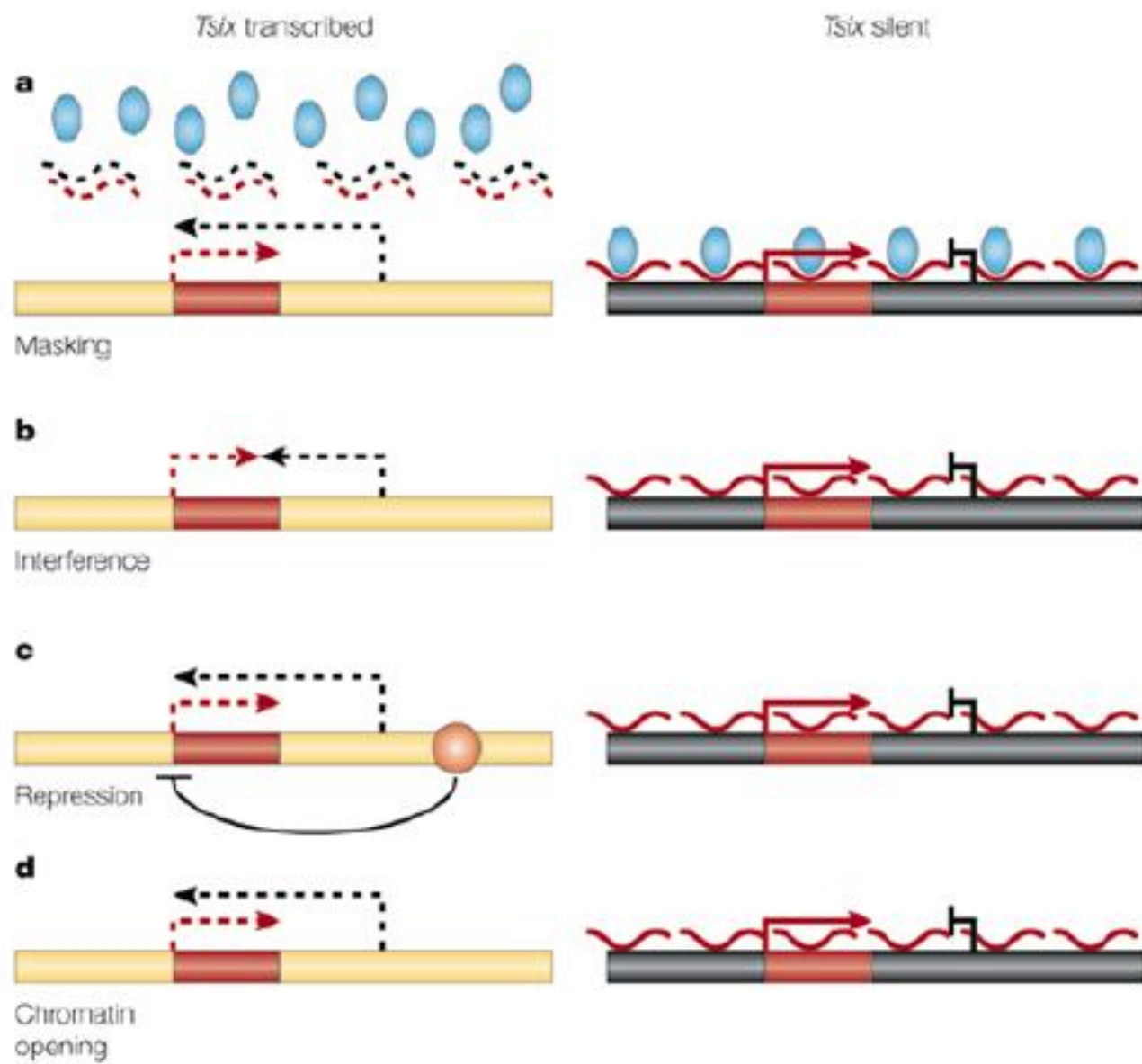
460 kb YAC (Heard et al, 1999)

80 kb BAC (Lee et al, 1999)

35 kb cosmid (Herzing et al, 1997)

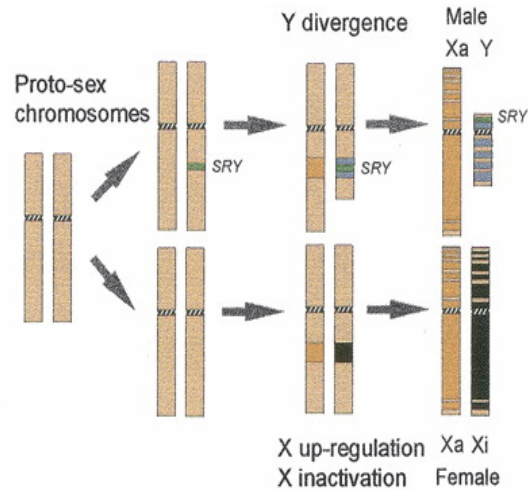
Xist cDNA (Wutz and Jaenisch 2000)

BAC / plasmids (Lee 2005)

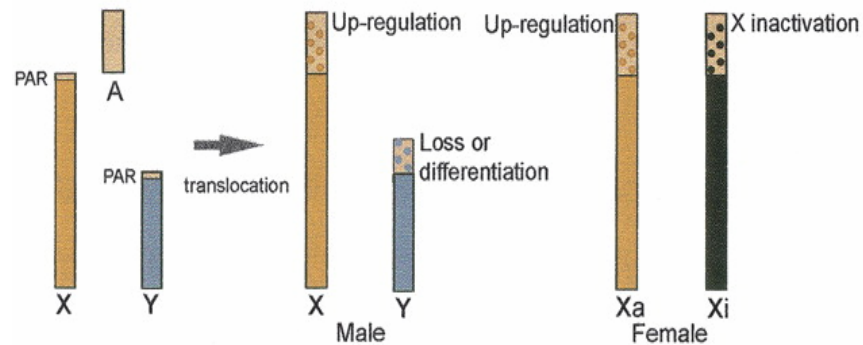


Sex chromosome evolution

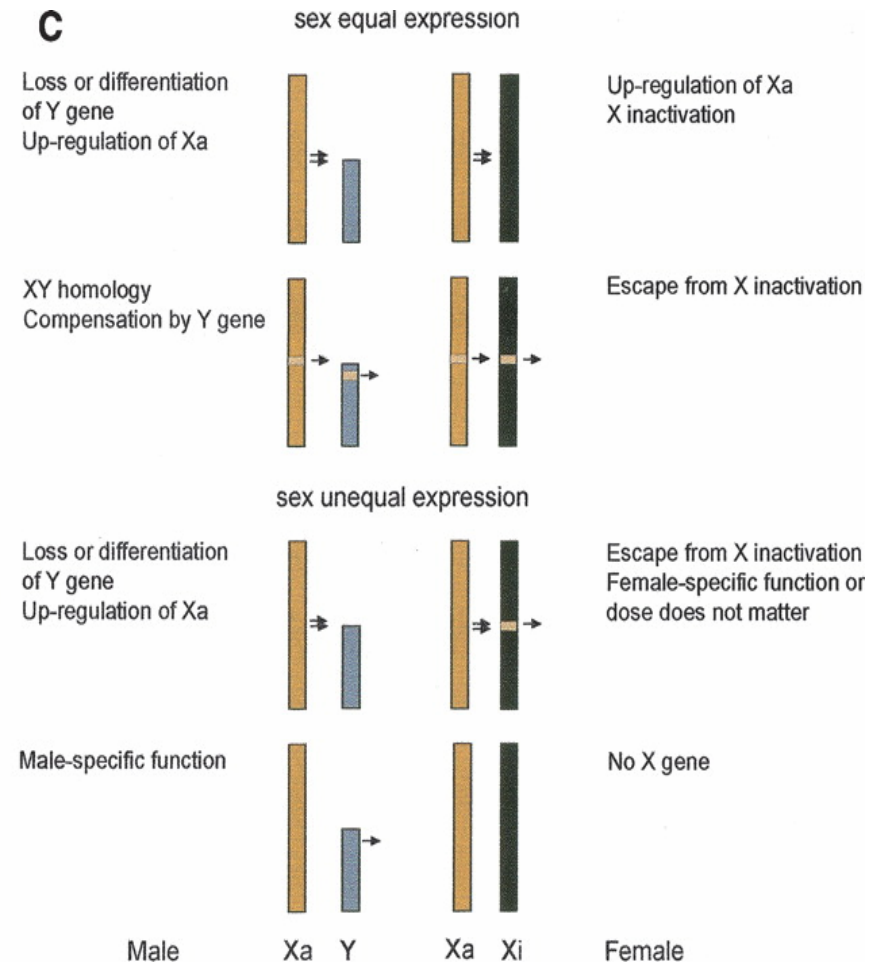
A



B



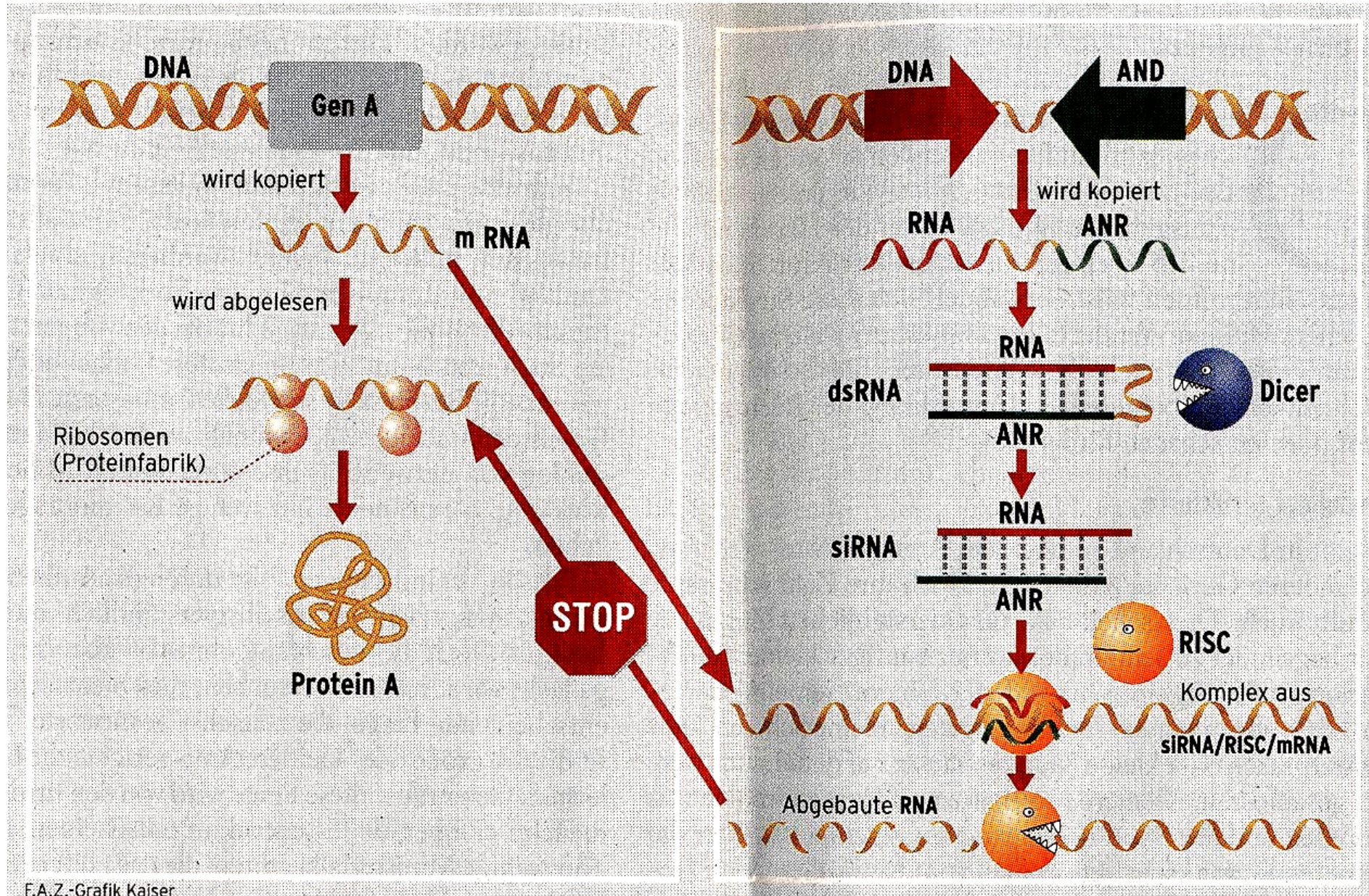
C



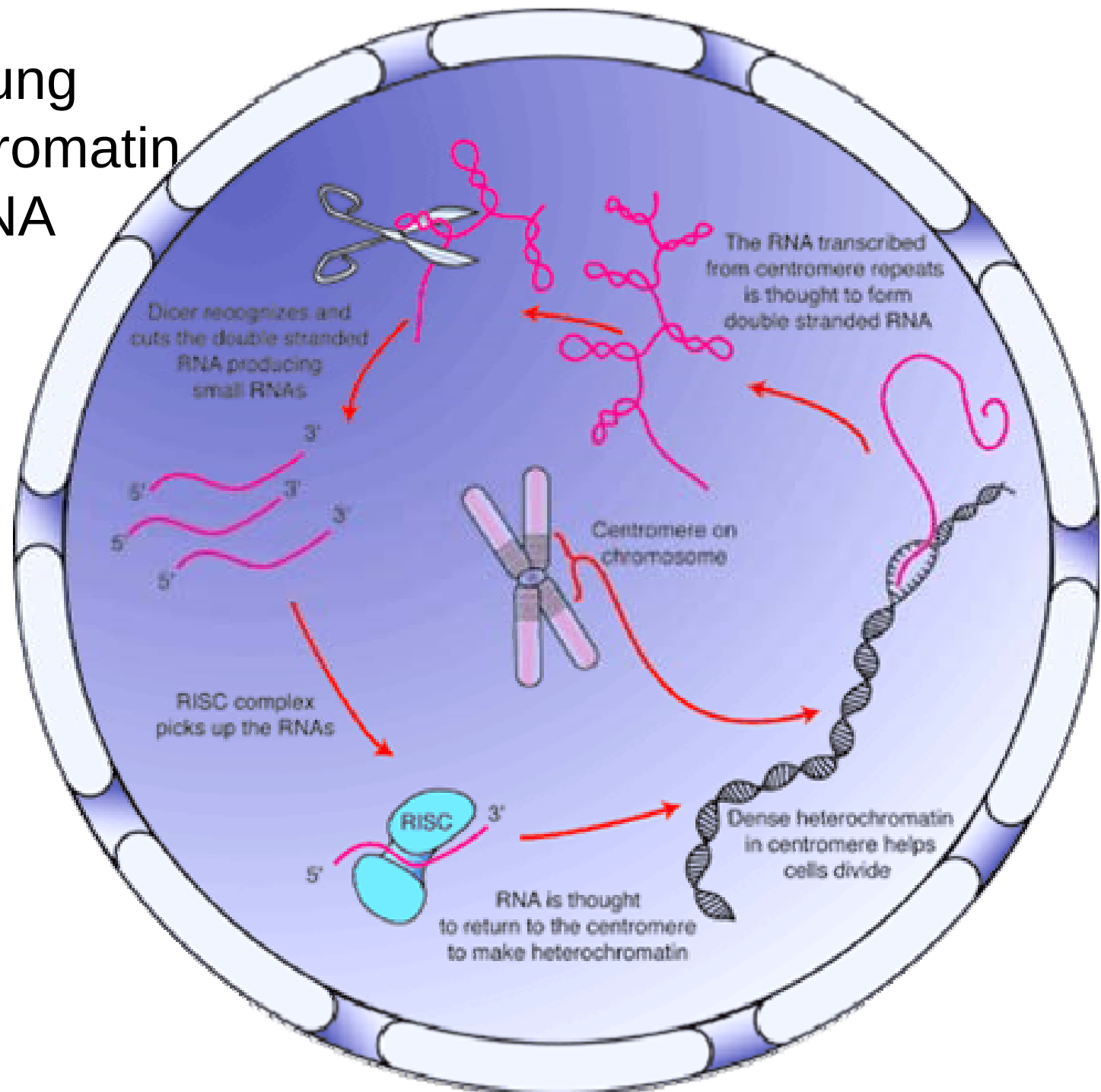
Xist continued...

- RNA is expressed, but does not code for protein
 - longest ORF (open reading frame) is 400 bp
- mouse 15 kb RNA, human 17 kb RNA
- RNA remains trapped in nucleus
- expressed only from the inactive X chromosome
- expressed in all mature female cells, and briefly in testes when an inactive X chromosome is present
- stabilization of Xist RNA mediates initiation of X inactivation

Genregulation durch RNAi

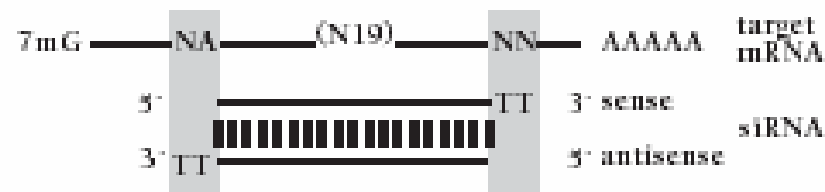


Inaktivierung von Heterochromatin durch RNA



Therapie von Leukämie durch dsRNA

A



B

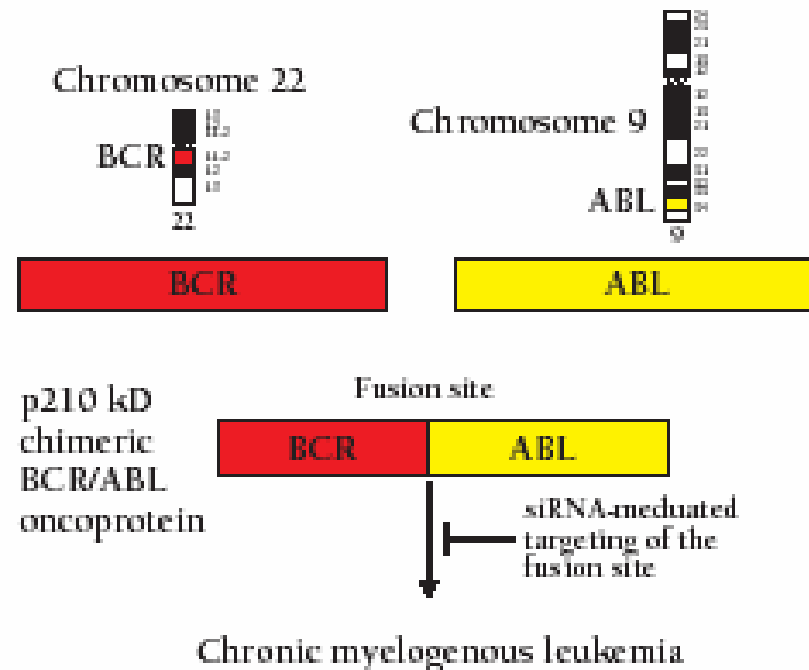
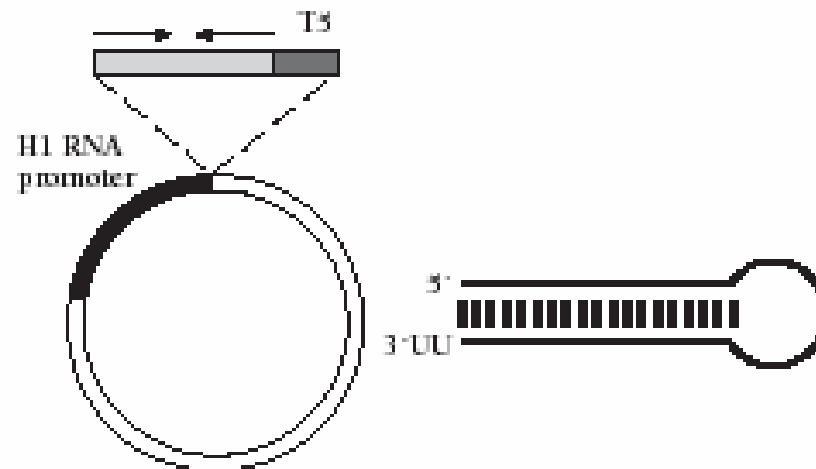


Figure 3. Scheme of the translocation t(9;22) in leukemia. The BCR-ABL fusion mRNA provides a leukemia-specific target that can be cleaved by siRNAs.



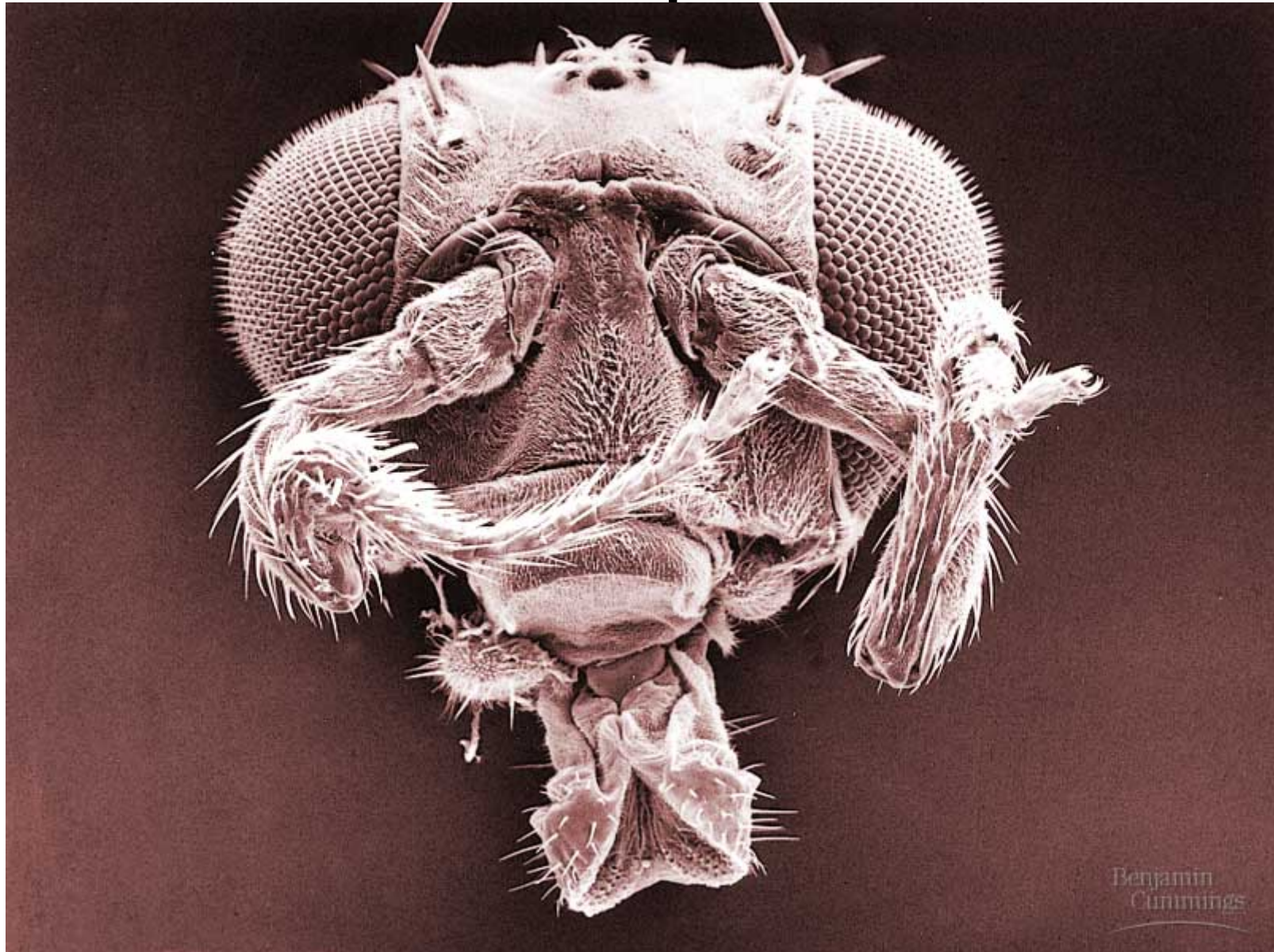
RNA interference (RNAi) represents an evolutionarily conserved cellular defense for controlling the expression of foreign genes in most eukaryotes including humans. RNAi is triggered by double-stranded RNA (dsRNA) and causes sequence-specific mRNA degradation of single-stranded target RNAs homologous in response to dsRNA. The mediators of mRNA degradation are small interfering RNA duplexes (siRNAs), which are produced from long dsRNA by enzymatic cleavage in the cell. siRNAs are approximately twenty-one nucleotides in length, and have a base-paired structure characterized by two-nucleotide 3'-overhangs. Chemically synthesized siRNAs have become powerful reagents for genome-wide analysis of mammalian gene function in cultured somatic cells. Beyond their value for validation of gene function, siRNAs also hold great potential as gene-specific therapeutic agents.

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²Children's University Hospital, Pediatric Hematology and Oncology, 35392 Giessen, Germany

Homöotische Mutante: Antennapedia



Bitte beachten:

- Praktikum beginnt 20. 02. 07 9 Uhr s. t.
- Praktikumsraum Molekulargenetik
(Gebäude SB1, J.-J.-Becherweg 34-36)
- Laborkittel mit bringen
- Praktikumsskript kann ab Montag 12. 2. im
Sekretariat des Institut für
Molekulargenetik abgeholt werden
(kostenpflichtig)